

# Package ‘XINA’

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**Type** Package

**Title** Multiplexed Isobaric Mass Tagged-based Kinetics Data for Network Analysis

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**biocViews** SystemsBiology, Proteomics, RNASeq, Network

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**Description** The aim of XINA is to determine which proteins exhibit similar patterns within and across experimental conditions, since proteins with co-abundance patterns may have common molecular functions. XINA imports multiple datasets, tags dataset in silico, and combines the data for subsequent subgrouping into multiple clusters. The result is a single output depicting the variation across all conditions. XINA, not only extracts coabundance profiles within and across experiments, but also incorporates protein-protein interaction databases and integrative resources such as KEGG to infer interactors and molecular functions, respectively, and produces intuitive graphical outputs.

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add_legend	<i>add_legend</i>
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---

### Description

Add plot legend and locate it outside of a network plot

### Usage

```
add_legend(legend_location = "bottomright", ...)
```

### Arguments

legend_location	
...	Network centrality score matrix Numeric, complex, or logical vectors.

### Value

a legend to a plot

---

alluvial\_enriched      *alluvial\_enriched*


---

## Description

'alluvial\_enriched' draws an alluvial plot and finds comigrated proteins. The comigration is a group of proteins that show the same expression pattern, classified and evaluated by XINA clustering, in at least two conditions. XINA can reduce the dataset complexity by filtering based on the number of comigrated proteins (size, 'comigration\_size' parameter) and perform an enrichment test (P-value of Fisher's exact test, 'pval\_threshold') to determine significance of enriched comigrations. The Fisher's exact test can only be done for two conditions at a time. The following 2x2 table was used to calculate the P-value from the Fisher's exact test. To evaluate significance of co-migrated proteins from cluster #1 in control to cluster #2 in test group,

	-	cluster #1 in control	other clusters in control
cluster #2 in test		65 (TP)	175 (FP)
other clusters in test		35 (FN)	979 (TN)

## Usage

```
alluvial_enriched(clustering_result, selected_conditions,
  comigration_size = 0, pval_threshold = 1, pval_method = "fdr",
  cex = 0.7, alpha = 0.3)
```

## Arguments

**clustering\_result**  
A list containing XINA clustering results. See [xina\\_clustering](#)

**selected\_conditions**  
A vector of condition names used in XINA clustering results. The number of selected conditions should be at least two.

**comigration\_size**  
The number of proteins comigrated together in the selected conditions of XINA clustering results. Default is 0

**pval\_threshold**  
This option is available only when you selected two conditions for comigration search.

**pval\_method**  
Method for p-value adjustment. See [p.adjust](#)

**cex**  
Scaling of fonts of category labels. Default if 0.7. See [alluvial](#)

**alpha**  
Transparency of the stripes. Default if 0.3. See [alluvial](#)

## Value

A data frame containing comigrations and an alluvial plot showing comigrations

**Examples**

```

# load XINA example data
data(xina_example)

# Get the experimental conditions in the example data
classes <- as.vector(example_clusters$condition)

# Get comigrations without any thresholds
all_comigrations <- alluvial_enriched(example_clusters, classes)

# Get comigrations that have >= 5 size (the number of comigrated proteins)
all_cor_enriched <- alluvial_enriched(example_clusters, classes, comigration_size=5)

# Get all the comigrations between Control and Stimulus1
comigrations_Control_Stimulus1 <- alluvial_enriched(example_clusters,
c(classes[1],classes[2]))

# Get comigrations between Control and Stimulus1, that have >=5 size
comigrations_Control_Stimulus1_over5 <- alluvial_enriched(example_clusters,
c(classes[1],classes[2]), comigration_size=5)

# Get comigrations between Control and Stimulus1,
# that have >= 5 size and enrichment FDR <= 0.01
comigrations_Control_Stimulus1_pval0.01_size5 <- alluvial_enriched(example_clusters,
c(classes[1],classes[2]), comigration_size=5, pval_threshold=0.01)

# Get comigrations between Control and Stimulus1,
# that have >= 5 size and enrichment Benjamini & Yekutieli <= 0.01
comigrations_Control_Stimulus1_BY0.01_size5 <- alluvial_enriched(example_clusters,
c(classes[1],classes[2]), comigration_size=5, pval_threshold=0.01, pval_method="BY")

```

---

alluvial\_enrichment\_tests

*alluvial\_enrichment\_tests*


---

**Description**

Fisher's exact test to calculate the significance over all comigrations. The following 2x2 table was used to calculate p-value from Fisher's exact test. To evaluate significance of comigrated proteins from cluster #1 in control to cluster #2 in test condition,

	<b>cluster #1 in control</b>	<b>other clusters in control</b>
<b>cluster #2 in test</b>	65 (TP)	175 (FP)
<b>other clusters in test</b>	35 (FN)	979 (TN)

'alluvial\_enrichment\_tests' also provides another statistical methods including Hypergeometric test and Chi-square test.

**Usage**

```
alluvial_enrichment_tests(count_table, c1, c2, non_cluster = 0,
  test_type = "fisher")
```

**Arguments**

count_table	A data frame generated by using <a href="#">count</a> .
c1	A selected cluster in the first condition.
c2	A selected cluster in the second condition.
non_cluster	The cluster number for proteins that were not detected in a specific sample. Default is 0.
test_type	Enrichment test type. 'fisher' = Fisher's exact test, 'hyper' = Hypergeometric test, 'chisq' = Chi-square test

**Value**

P-value of comigration enrichment test and 2x2 table information

---

calculate\_centrality\_scores  
*calculate\_centrality\_scores*

---

**Description**

'calculate\_centrality\_scores' computes network centrality scores

**Usage**

```
calculate_centrality_scores(net, centrality_type = "Degree")
```

**Arguments**

net	protein-protein interaction network of igraph
centrality_type	the maximum number of clusters

**Value**

A vector of network centrality scores

---

default_size	<i>default_size</i>
--------------	---------------------

---

**Description**

Calculate image size based on the number of clusters

**Usage**

```
default_size(max_cluster)
```

**Arguments**

max\_cluster     the maximum number of clusters

**Value**

A vector of plot width and height

---

draw_alluvial_plot	<i>draw_alluvial_plot</i>
--------------------	---------------------------

---

**Description**

'draw\_alluvial\_plot' draw a alluvial plot

**Usage**

```
draw_alluvial_plot(clustering_result, selected_conditions, count_table,
  alluvia_colors = NULL, cex = 0.7, alpha = 0.3)
```

**Arguments**

clustering\_result     A list containing XINA clustering results. See [xina\\_clustering](#).

selected\_conditions     A vector of condition names used in XINA clustering results. The number of selected conditions should be at least two.

count\_table     A data frame generated by using [count](#).

alluvia\_colors     A vector containing the user-defined colors for each alluvium.

cex     Size of cluster number on block axis. Default if 0.7. See [alluvial](#).

alpha     Transparency of alluvia colors. Default is 0.3. See [alluvial](#).

**Value**

An alluvial plot displaying comigrations and the data frame containing the input count\_table with colors.

**Examples**

```
# load XINA example data
data(xina_example)

# get a vector of experimental conditions analyzed in the clustering results
classes <- as.vector(example_clusters$condition)

comigrations_size_over5 <- alluvial_enriched(example_clusters, classes, comigration_size=5)
draw_alluvial_plot(example_clusters, classes, comigrations_size_over5)
```

---

example_clusters	<i>Randomly generated example datasets for XINA users. A dataset containing the XINA clustering results.</i>
------------------	--

---

**Description**

- aligned. XINA clustering results aligned by conditions
- data\_column. Column names for data matrix
- out\_dir. Not available in this example dataset
- nClusters. The number of user-desired clusters. It's 30 in the example.
- max\_cluster. The number of clusters found in the dataset. It's 21 in the example.
- chosen\_model. The chosen covariance model for the example dataset. It's VEI in the example
- optimal\_BIC. BIC at the optimized clustering. It's 29473.57 in the example
- condition. The experimental conditions in the dataset.
- color\_for\_condition. The default color for the conditions that will be used in XINA plot drawing.
- color\_for\_clusters. The default color for the clusters that will be used in XINA clustering plot.
- norm\_method. The used normalization method to standardize the input data. It's "sum\_normalization" in the example.

**Format**

A list with the example XINA clustering result



---

extract\_data\_column    *extract\_data\_column*

---

**Description**

Extract data column names from XINA clustering result

**Usage**

```
extract_data_column(col_head_of_clustering)
```

**Arguments**

col\_head\_of\_clustering  
Column names of XINA clustering result

**Value**

A vector containing column names of data matrix

---

find\_similar\_clusters    *find\_similar\_clusters*

---

**Description**

Compare clusters and find similar ones

**Usage**

```
find_similar_clusters(clustering_result, threshold = 0.95)
```

**Arguments**

clustering\_result  
A list containing XINA clustering results. See [xina\\_clustering](#)

threshold    Pearson's r threshold to find similar ones

**Value**

Write a csv file containing similar clustering information based on the given Pearson's R threshold

---

generate\_count\_table    *generate\_count\_table*

---

### Description

Count the number of comigrated proteins using [count](#)

### Usage

```
generate_count_table(clustering_result, selected_conditions,
                    comigration_size)
```

### Arguments

`clustering_result`  
A list containing XINA clustering results. See [xina\\_clustering](#)

`selected_conditions`  
A vector of condition names used in XINA clustering results.

`comigration_size`  
The number of proteins comigrated together in the selected conditions of XINA clustering results. Default is 0.

### Value

A data frame containing comigrations.

---

generate\_superset    *generate\_superset*

---

### Description

Merge input kinetics files

### Usage

```
generate_superset(f_names, data_column, delim = ",",
                 norm = "sum_normalization")
```

### Arguments

`f_names`            A vector of .csv file paths containing kinetics data

`data_column`        A vector of column names containing data matrix

`delim`                The delimiter of input file (default is ',')

`norm`                 The normalization method. It should be one of `c('sum_normalization', 'zscore')`. Default is `'sum_normalization'`.

**Value**

A data frame containing kinetics data obtained from files in the f\_names vector

---

get_colors	<i>get_colors</i>
------------	-------------------

---

**Description**

Generate color series for XINA graphics

**Usage**

```
get_colors(nClusters, set = "", colorset = NULL)
```

**Arguments**

nClusters	The number of clusters
set	Pre-defined color series set
colorset	manually defined color codes

**Value**

A vector for color code of XINA graphics

---

get_color_for_nodes	<i>get_color_for_nodes</i>
---------------------	----------------------------

---

**Description**

Pre-defined 30 colors

**Usage**

```
get_color_for_nodes()
```

**Value**

A vector for color code of XINA graphics

---

```
get_comigrations_by_name  
    get_comigrations_by_name
```

---

### Description

'*get\_comigrations\_by\_name*' finds proteins comigrated with the given proteins

### Usage

```
get_comigrations_by_name(clustering_result, selected_conditions,  
    protein_list, cex = 0.7, alpha = 0.3)
```

### Arguments

<code>clustering_result</code>	A list containing XINA clustering results. See <a href="#">xina_clustering</a>
<code>selected_conditions</code>	A vector of condition names used in XINA clustering results. The number of selected conditions should be at least two.
<code>protein_list</code>	A vector containing gene names.
<code>cex</code>	Size of cluster number on block axis. Default if 0.7. See <a href="#">alluvial</a>
<code>alpha</code>	Transparency of alluvia colors. Default is 0.3. See <a href="#">alluvial</a>

### Value

An alluvial plot displaying comigrations and the data frame containing comigrations of the input proteins

### Examples

```
# load XINA example data  
data(xina_example)  
  
# the clustering result table  
all_proteins <- as.character(example_clusters$aligned$`Gene name`)  
# get a vector of experimental conditions analyzed in the clustering results  
classes <- as.vector(example_clusters$condition)  
  
comigrated_prots_all <- get_comigrations_by_name(example_clusters, classes, all_proteins[1:3])
```

---

```
get_condition_biased_comigrations
      get_condition_biased_comigrations
```

---

## Description

get comigrations that at least one biased cluster is involved in. Biased clusters are defined by

## Usage

```
get_condition_biased_comigrations(clustering_result, count_table = NULL,
  selected_conditions, condition_composition, threshold_percent = 50,
  color_for_null = "gray", color_for_highly_matched = "red4",
  cex = 0.7, alpha = 0.3)
```

## Arguments

clustering_result	A list containing XINA clustering results. See <a href="#">xina_clustering</a>
count_table	A data frame generated by using <a href="#">count</a> . If count_table is NULL (by default), XINA will consider all the comigrations.
selected_conditions	A vector of condition names used in XINA clustering results. The number of selected conditions should be at least two.
condition_composition	The resulting data frame of 'plot_condition_compositions'. See <a href="#">plot_condition_compositions</a> .
threshold_percent	Default is 50. The percentage threshold for finding condition-biased clusters
color_for_null	A color for non-condition-biased comigrations. Default is 'gray'
color_for_highly_matched	A color for comigrations that are involved with more than two condition-biased clusters. Default is 'red4'
cex	Size of cluster number on block axis. Default if 0.7. See <a href="#">alluvial</a> .
alpha	Transparency of alluvia colors. Default is 0.3. See <a href="#">alluvial</a> .

## Value

An alluvial plot displaying comigrations and the data frame containing condition-biased comigrations.

## Examples

```
# load XINA example data
data(xina_example)

# get a vector of experimental conditions analyzed in the clustering results
```

```
conditions <- as.vector(example_clusters$condition)

# get condition composition information
condition_composition <- plot_condition_compositions(example_clusters)

comigrations_size10 <- alluvial_enriched(example_clusters, conditions, comigration_size=10)
# Finding condition-biased comigrations by 50% threshold
condition_biased_comigrations <-
get_condition_biased_comigrations(clustering_result=example_clusters,
count_table=comigrations_size10, selected_conditions=conditions,
condition_composition=condition_composition)

# Finding condition-biased comigrations by 70% threshold
condition_biased_comigrations <-
get_condition_biased_comigrations(clustering_result=example_clusters,
count_table=comigrations_size10, selected_conditions=conditions,
condition_composition=condition_composition,
threshold_percent=70)
```

---

`get_layout`

*get\_layout*

---

## Description

Get igraph layout by the number of nodes

## Usage

```
get_layout(subnet_condition)
```

## Arguments

subnet\_condition  
A igraph sub-network

## Value

igraph network layout

---

```
get_mTOR_proteins      get_mTOR_proteins
```

---

**Description**

Get mTOR pathway genes

**Usage**

```
get_mTOR_proteins(time_points, conditions)
```

**Arguments**

time_points	A vector containing time points of the data matrix
conditions	A vector containing condition information, for example normal, disease and drug treated disase.

**Value**

A vector containing mTOR pathway gene names

---

```
get_random_data      get_random_data
```

---

**Description**

Get randomized time-series data

**Usage**

```
get_random_data(time_points, conditions, num_total, percent.sign = 0.1,
  equal = TRUE)
```

**Arguments**

time_points	A vector containing time points of the data matrix
conditions	A vector containing condition information, for example normal, disease and drug treated disase.
num_total	The number of total proteins to be generated
percent.sign	Percentage of differentially expressed proteins. Ignored when equal=FALSE.
equal	If equal is TRUE, all the conditions will have numbers between 0 and 1. If it is FALSE, the first three conditions will have different ranges. First condition will have numbers from 0.3 to 0.4. Second condition will have numbers from 0.6 to 0.8. Third condition will have numbers from 0.3 to 0.5. Other conditions will have numbers from 0 to 1.

**Value**

A list containing randomly generated data matrix

---

<code>get_stats</code>	<i>get_stats</i>
------------------------	------------------

---

**Description**

Calculate statistics of the given data for XINA network analysis

**Usage**

```
get_stats(centrality_results, na.rm = FALSE)
```

**Arguments**

<code>centrality_results</code>	Network centrality score data frame calculated by XINA network module
<code>na.rm</code>	If it is FALSE, no exclusion of NA values.

**Value**

A data frame containing statistics of XINA network centrality scores

---

<code>get_theme_blank</code>	<i>get_theme_blank</i>
------------------------------	------------------------

---

**Description**

Predefined ggplot theme for removing ticks, titles and labels of X and Y axis

**Usage**

```
get_theme_blank()
```

**Value**

A ggplot theme



---

```
get_unknown_ppi_nodes get_unknown_ppi_nodes
```

---

**Description**

Get proteins with no known interactions within the cluster based on the used protein-protein interaction database source

**Usage**

```
get_unknown_ppi_nodes(xina_result, cl)
```

**Arguments**

`xina_result` A list containing XINA network analysis results. See [xina\\_analysis](#)  
`cl` the clustering number of XINA clustering results. See [xina\\_clustering](#)

**Value**

A data frame containing proteins with no known interactions within the cluster based on the used protein-protein interaction database source

**Examples**

```
# load XINA example data
data(xina_example)

# load the previously processed XINA analysis results
# if you want to learn how to run 'xina_analysis', please see \link[XINA]{xina_analysis}
data(xina_result_example)

# Extract unknown PPI nodes in the cluster #1
get_unknown_ppi_nodes(xina_result_example, 1)
```

---

<code>gn</code>	<i>A character vector containing 19,396 human genes This is for the random data generation of XINA</i>
-----------------	--

---

**Description**

- Characters of human genes

**Format**

A character vector containing 19,396 human genes

**Source**

<https://www.ncbi.nlm.nih.gov/gene>

---

gn_desc	<i>A character vector containing 19,396 human gene descriptions This is for the randome data generation of XINA</i>
---------	---

---

**Description**

- Human gene description corresponding to 'gn' vector

**Format**

A character vector containing 19,396 human gene descriptions

**Source**

<https://www.ncbi.nlm.nih.gov/gene>

---

hprd_ppi	<i>Protein-protein interaction resource downloaded from HPRD DB A data frame containing HRPD protein-protein interaction data</i>
----------	---

---

**Description**

- gene\_symbol\_1. Gene name interacting with gene name in 'gene\_symbol\_2'
- gene\_symbol\_2. Gene name interacting with gene name in 'gene\_symbol\_1'
- Experiment\_type. Experimental or computational methods supporting the interaction

**Format**

A data frame containing HRPD protein-protein interaction data

**Source**

<http://www.hprd.org/>

---

length2	<i>length2</i>
---------	----------------

---

**Description**

Customized function for vector length calculation

**Usage**

```
length2(x, na.rm = FALSE)
```

**Arguments**

x	A vector
na.rm	If it is FALSE, no exclusion of NA values.

**Value**

A vector length

---

load_previous_results	<i>load_previous_results</i>
-----------------------	------------------------------

---

**Description**

Get previous XINA clustering results to R space

**Usage**

```
load_previous_results(clustering_dir = getwd(), data_column = NULL,
  fp_clusters = "xina_clusters.csv")
```

**Arguments**

clustering_dir	The directory path of XINA clustering results
data_column	A vector containing column names of data matrix
fp_clusters	File path of XINA clustering results

**Value**

Comma-separated file containing aligned XINA clustering results.

**Examples**

```
# Load XINA's example data
data(xina_example)
write.csv(example_clusters$aligned,"xina_clusters_aligned.csv")
write.csv(example_clusters$clusters,"xina_clusters.csv")

# Reload the clustering result
example_clusters_reloaded <- load_previous_results(".")
```

---

```
make_random_xina_data  make_random_xina_data
```

---

**Description**

Generate random proteomics dataset for testing XINA 'make\_random\_xina\_data' will make random proteomics data for XINA test. The generated data will have three conditions and seven time points, c("0hr", "2hr", "6hr", "12hr", "24hr", "48hr", "72hr").

**Usage**

```
make_random_xina_data(n = 500, mtor = TRUE, time_points = c("0hr",
  "2hr", "6hr", "12hr", "24hr", "48hr", "72hr"),
  conditions = c("Control", "Stimulus1", "Stimulus2"))
```

**Arguments**

n	The number of proteins for one condition. Default is 500.
mtor	If it is TRUE (default), mTOR pathway genes will be significant. If it is FALSE, randomly selected genes will be significant in first three conditions.
time_points	A vector containing time points of the data matrix
conditions	A vector containing condition information, for example normal, disease and drug treated disease.

**Value**

Three comma-separated files containing time-series data for XINA

**Examples**

```
make_random_xina_data()
g1 <- read.csv("Control.csv", check.names=FALSE,
stringsAsFactors = FALSE)
g2 <- read.csv("Stimulus1.csv", check.names=FALSE,
stringsAsFactors = FALSE)
g3 <- read.csv("Stimulus2.csv", check.names=FALSE,
stringsAsFactors = FALSE)
```

```
head(g1)
head(g2)
head(g3)
```

---

```
mutate_colors      mutate_colors
```

---

### Description

'mutate\_colors' generates new color scheme for XINA clustering plot based on condition composition results ([plot\\_condition\\_compositions](#)). If any clusters have higher percentage than the 'threshold\_percent', XINA will assign new colors in accordance to 'color\_for\_condition'. If not, XINA will give 'gray' color or user-defined color via 'null\_color' parameter.

### Usage

```
mutate_colors(condition_composition, color_for_condition,
              null_color = "gray", threshold_percent = 50)
```

### Arguments

condition\_composition  
A data frame generated by [plot\\_condition\\_compositions](#)

color\_for\_condition  
A vector like 'color\_for\_condition' of [xina\\_clustering](#)

null\_color  
Default is 'gray'. This color is for clusters that are not biased to any of experimental conditions

threshold\_percent  
Default is 50. The percentage threshold for giving new colors

### Value

A data frame containing statistics of XINA network centrality scores

### Examples

```
# load XINA example data
data(xina_example)

# Plot condition composition pie-chart with default option
condition_composition <- plot_condition_compositions(example_clusters)
example_clusters$color_for_clusters <- mutate_colors(condition_composition,
example_clusters$color_for_condition)
plot_clusters(example_clusters, xval=c(0,2,6,12,24,48,72), xylab=FALSE)
```

---

organize_clusters	<i>organize_clusters</i>
-------------------	--------------------------

---

**Description**

Organize XINA clustering information by gene name

**Usage**

```
organize_clusters(clustering_dir = getwd(), super_ds, file_out = TRUE)
```

**Arguments**

clustering_dir	The directory path of XINA clustering results
super_ds	XINA clusters
file_out	If it is TRUE, it writes the aligned clustering informaion to "xina_clusters_aligned.csv" file.

**Value**

Comma-separated file containing aligned XINA clustering results.

---

plot_clusters	<i>plot_clusters</i>
---------------	----------------------

---

**Description**

Draw all the clustering results. 'plot\_clusters' draws two plots, scaled and unscaled line graphs. Scaled graphs have same y limits that are 0 to 1 by default, but can be changed via 'y\_lim' parameter.

**Usage**

```
plot_clusters(clustering_result, y_lim = NULL, xval = NULL,
             xtickmark = NULL, xylab = TRUE, ggplot_theme = NULL)
```

**Arguments**

clustering_result	A list containing XINA clustering results. See <a href="#">xina_clustering</a>
y_lim	Y axis limit. If you set y_lim=c(0,1), 'plot_clusters' will plot line graphs scaled from 0 to 1 in y-axis Default is NULL, which means unscaled line graphs.
xval	XINA basically considers time points as a ordinary variable, like 1,2,3,4...n. You can make the time points as a continuous variable using xval.
xtickmark	Change X axis tick marks. Default is data_column of the clustering result list.
xylab	If it is FALSE, x and y labels will be blank. If it is TRUE (default), x and y labels will be shown.
ggplot_theme	This is ggplot theme to modify XINA clustering plot.

**Value**

Line graphs of all the clusters

**Examples**

```
library(ggplot2)

# load XINA example data
data(xina_example)

# Draw clustering plots
plot_clusters(example_clusters)

# Apply theme to the clustering plot
theme1 <- theme(title=element_text(size=8, face='bold'),
axis.text.x = element_text(size=7),
axis.text.y = element_blank(),
axis.ticks.x = element_blank(),
axis.ticks.y = element_blank(),
axis.title.x = element_blank(),
axis.title.y = element_blank())
plot_clusters(example_clusters, ggplot_theme=theme1)
```

---

plot\_clusters\_all      *plot\_clusters\_all*

---

**Description**

Draw line graphs of all the proteins in the given dataset

**Usage**

```
plot_clusters_all(clustering_result, selected_condition = NULL)
```

**Arguments**

clustering\_result

A list containing XINA clustering results. See [xina\\_clustering](#)

selected\_condition

A condition name to draw the kinetics plot

**Value**

a list containing clustering results and pdf file containing a BIC plot in current working directory.

## Examples

```
# load XINA example data
data(xina_example)

# Plot kinetics of all the proteins in Control
plot_clusters_all(example_clusters, selected_condition="Control")

# Plot kinetics of all the proteins in Stimulus1
plot_clusters_all(example_clusters, selected_condition="Stimulus1")

# Plot kinetics of all the proteins in Stimulus2
plot_clusters_all(example_clusters, selected_condition="Stimulus2")

# Plot kinetics of all the proteins in three data
plot_clusters_all(example_clusters)
```

---

```
plot_condition_compositions
      plot_condition_compositions
```

---

## Description

computes condition composition of the XINA clustering results and draws pie-charts.

## Usage

```
plot_condition_compositions(clustering_result, bullseye = FALSE,
  ggplot_theme = NULL)
```

## Arguments

clustering_result	A list containing XINA clustering results. See <a href="#">xina_clustering</a>
bullseye	If it is TRUE, draw bullseye plot instead of the pie-chart. Default is FALSE
ggplot_theme	This is ggplot theme to modify condition composition pie-chart and bulles eye plots.

## Value

A condition composition plot and a data frame containing condition compositions of the clusters



**Examples**

```
# load XINA example data
data(xina_example)

# Plot condition composition pie-chart with default option
plot_condition_compositions(example_clusters)

# Make a new color code for conditions
condition_colors <- c("tomato","steelblue1","gold")
names(condition_colors) <- example_clusters$condition
example_clusters$color_for_condition <- condition_colors

# Draw condition composition pie-chart with the new color code
plot_condition_compositions(example_clusters)

# Draw condition composition bullseye plot
plot_condition_compositions(example_clusters, bullseye = TRUE)
```

---

```
plot_enrichment_results
      plot_enrichment_results
```

---

**Description**

Plot GO and KEGG enrichment results

**Usage**

```
plot_enrichment_results(enriched_results,
  term_description = "term_description", sig_score = "pvalue",
  num_terms = 0, get_log = TRUE, fill_color = "darkgray")
```

**Arguments**

enriched_results	GO or KEGG enrichment results. See <a href="#">xina_enrichment</a> and <a href="#">xina_enrichment</a>
term_description	Description of terms to be drawn on Y axis. Default is "term_description" of XINA enrichment results.
sig_score	significant score to plot on X axis. Default is "pvalue".
num_terms	The number of terms to be plotted. Default is 0, which means no limit.
get_log	If this is TRUE, 'plot_enrichment_results' will take -log10 of p-values.
fill_color	Default is 'darkgray'. You can change color of bars.

**Value**

ggplot bar graph

**Examples**

```
## Not run:
library(STRINGdb)

# load XINA example data
data(xina_example)

# Get STRING database for protein-protein intereaction information
string_db <- STRINGdb$new( version="10", species=9606,
score_threshold=0, input_directory="" )
string_db

# XINA analysis with STRING DB
xina_result <- xina_analysis(example_clusters, string_db)

# Select proteins that showed cluster #1 in the Stimulus2 condition
subgroup <- subset(example_clusters$aligned, Stimulus2==1)
protein_list <- as.vector(subgroup$`Gene name`)

# Enrichment test and get significantly enriched functional terms
# that have adjusted p-value less than 0.1
kegg_enriched <- xina_enrichment(string_db, protein_list,
enrichment_type = "KEGG", pval_threshold=0.1)
plot_enrichment_results(kegg_enriched$KEGG, num_terms=10)

## End(Not run)
```

---

plot\_NA

*plot\_NA*

---

**Description**

Draw NULL plot

**Usage**

```
plot_NA()
```

**Value**

a empty plot

---

rank centrality	<i>rank centrality</i>
-----------------	------------------------

---

**Description**

Give ranks based on network centrality scores

**Usage**

```
rank centrality(centrality_score, type, num_breaks = 5)
```

**Arguments**

centrality_score	Network centrality score matrix
type	Network centrality score type, such as 'Eigenvector'
num_breaks	The number of ranks

**Value**

A vector containing ranks

---

string_example	<i>Protein-protein interaction resource downloaded from STRING DB for XINA's example dataset A data frame containing protein-protein interactions</i>
----------------	---

---

**Description**

- gene\_symbol\_1. Gene name interacting with gene name in 'gene\_symbol\_2'
- gene\_symbol\_2. Gene name interacting with gene name in 'gene\_symbol\_1'
- PPI\_Source. Data original source

**Format**

A data frame containing STRING protein-protein interaction data

**Source**

<https://string-db.org/>

---

xina\_analysis                      *xina\_analysis*

---

## Description

xina\_analysis is to analyze protein-protein interaction(PPI) networks using STRINGdb and igraph R package. This module computes PPI networks within each XINA clusters.

## Usage

```
xina_analysis(clustering_result, ppi_db, is_stringdb = TRUE,
              flag_simplify = TRUE, node_shape = "sphere",
              num_clusters_in_row = 5, img_size = NULL, img_qual = 300)
```

## Arguments

**clustering\_result**                      A list containing XINA clustering results. See [xina\\_clustering](#)

**ppi\_db**                                      STRINGdb object

**is\_stringdb**                                If it is TRUE (default), XINA will process 'ppi\_db' as STRINGdb, but it is FALSE, XINA will accepts your 'ppi\_db' as it is. You can make your own igraph network using customized PPI information instead of STRINGdb.

**flag\_simplify**                              If it is TRUE (default), XINA will exclude unconnected proteins

**node\_shape**                                You can choose node shape. Default is "sphere". See [shapes](#)

**num\_clusters\_in\_row**                      The number of clusters in a row on the XINA network plot. Default is 5.

**img\_size**                                  Set the image size. For width=1000 and height=1500, it is `img_size=c(1000,1500)`.

**img\_qual**                                  Set the image resolution. Default is 300.

## Value

A PNG file (XINA\_Cluster\_Networks.png) displaying PPI network plots of all the clusters and a list containing XINA network analysis results.

Item	Description
All_network	PPI network of all the input proteins
Sub_network	A list containing PPI networks of each clusters
Data	XINA clustering results. See <a href="#">xina_clustering</a>
Nodes	A list of proteins in each cluster
Conditions	A list of experimental condition of proteins in each cluster
Titles	A list of plot titles for XINA plotting
out_dir	A directory path storing XINA network analysis results
is_stringdb	False = different PPI DB and TRUE = STRING DB

**Examples**

```
## Not run:
# load XINA example data
data(xina_example)

# use the following code for utilizing up-to-date STRING DB
tax_id <- 9606 # for human
# tax_id <- 10090 # for mouse
library(STRINGdb)
library(igraph)
string_db <- STRINGdb$new( version='10', species=tax_id, score_threshold=0, input_directory='' )
string_db
xina_result <- xina_analysis(example_clusters, string_db, flag_simplify=FALSE)

# Run XINA with a protein-protein interaction edgelist
data(HPRD)
net_all <- simplify(graph_from_data_frame(d=hprd_ppi, directed=FALSE),
  remove_multiple = FALSE, remove_loops = TRUE)
xina_result <- xina_analysis(example_clusters, net_all, is_stringdb=FALSE, flag_simplify=FALSE)

## End(Not run)
```

---

xina_clustering	<i>xina_clustering</i>
-----------------	------------------------

---

**Description**

Clustering multiplexed time-series omics data to find co-abundance profiles

**Usage**

```
xina_clustering(f_names, data_column, out_dir = getwd(),
  nClusters = 20, norm = "sum_normalization", chosen_model = "")
```

**Arguments**

f_names	A vector containing input file (.csv) paths
data_column	A vector containing column names (1st row of the input file) of data matrix
out_dir	A directory path for saving clustering results. (default: out_dir=getwd())
nClusters	The number of desired maximum clusters
norm	Default is "sum_normalization". Sum-normalization is to divide the data matrix by row sum. If you want to know more about sum-normalization, see <a href="https://www.ncbi.nlm.nih.gov/pubmed/19861354">https://www.ncbi.nlm.nih.gov/pubmed/19861354</a> . "zscore" is to calculate Z score for each protein. See <a href="#">scale</a> .
chosen_model	You can choose a specific model rather than testing all the models that are available in mclust. <a href="#">mclustModelNames</a> If you want k-means clustering instead of the model-based clustering, use "kmeans" here.

**Value**

a plot containing a BIC plot in current working directory and a list containing below information:

<b>Item</b>	<b>Description</b>
clusters	XINA clustering results
aligned	XINA clustering results aligned by ID
data_column	Data matrix column names
out_dir	The directory path containing XINA results
nClusters	The number of clusters desired by user
max_cluster	The number of clusters optimized by BIC
chosen_model	The used covariance model for model-based clustering
optimal_BIC	BIC of the optimized covariance model
condition	Experimental conditions of the user input data
color_for_condition	Colors assigned to each experimental conditions which is used for condition composition plot
color_for_clusters	Colors assigned to each clusters which is used for XINA clustering plot
norm_method	Used normalization method

**Examples**

```
# Generate random multiplexed time-series data
random_data_info <- make_random_xina_data()

# Data files
data_files <- paste(random_data_info$conditions, ".csv", sep='')

# time points of the data matrix
data_column <- random_data_info$time_points

# mclust requires the fixed random seed to get reproduce the clustering results
set.seed(0)

# Run the model-based clustering to find co-abundance profiles
example_clusters <- xina_clustering(data_files, data_column=data_column,
nClusters=30)

# Run k-means clustering to find co-abundance profiles
example_clusters <- xina_clustering(data_files, data_column=data_column,
nClusters=30,
chosen_model="kmeans")
```

---

xina\_enrichment

*xina\_enrichment*


---

**Description**

xina\_enrichment conducts functional enrichment tests using gene ontology or KEGG pathway terms for a given protein list

**Usage**

```
xina_enrichment(string_db, protein_list, enrichment_type = "GO",
  pval_threshold = 0.05, methodMT = "fdr")
```

**Arguments**

string_db	STRINGdb object
protein_list	A vector of gene names to draw protein-protein interaction network.
enrichment_type	A functional annotation for the enrichment test. 'enrichment_type' should be one of 'GO' and 'KEGG'.
pval_threshold	P-value threshold to get significantly enriched terms from the given proteins
methodMT	Method for p-value adjustment. See <a href="#">get_enrichment</a> . Default is 'fdr'.

**Value**

A list of data frames containing enrichment results

**Examples**

```
## Not run:
library(STRINGdb)
library(Biobase)

# load XINA example data
data(xina_example)

# Get STRING database for protein-protein intereaction information
string_db <- STRINGdb$new( version="10", species=9606, score_threshold=0, input_directory="" )
string_db

# XINA analysis with STRING DB
xina_result <- xina_analysis(example_clusters, string_db)

# Select proteins that showed cluster #1 in the Stimulus2 condition
subgroup <- subset(example_clusters$aligned, Stimulus2==1)
protein_list <- as.vector(subgroup$`Gene name`)

# Enrichment test using KEGG pathway terms that have adjuseted p-value less than 0.1
kegg_enriched <- xina_enrichment(string_db, protein_list,
  enrichment_type = "KEGG", pval_threshold=0.1)
plot_enrichment_results(kegg_enriched$KEGG, num_terms=10)

# Enrichment test using GO terms that have adjuseted p-value less than 0.1
go_enriched <- xina_enrichment(string_db, protein_list,
  enrichment_type = "GO", pval_threshold=0.1)
plot_enrichment_results(go_enriched$Component, num_terms=10)

## End(Not run)
```

---

xina\_plot\_all                      *xina\_plot\_all*

---

## Description

xina\_plot\_all is to draw protein-protein interaction network plots of all the clusters

## Usage

```
xina_plot_all(xina_result, clustering_result, condition = "all",
              centrality_type = NULL, flag_simplify = TRUE, num_breaks = 5,
              layout_specified = "", vertex_label_flag = FALSE,
              vertex.label.color = "black", vertex.color = "", edge.color = NULL,
              vertex.label.dist = 0.6, vertex.label.cex = 0.8,
              edge.arrow.size = 0.4, vertex.size = 10, vertex.shape = "sphere",
              legend_location = "bottom", num_clusters_in_row = 5,
              flag_unknown_only = FALSE, img_size = NULL, img_qual = 300)
```

## Arguments

**xina\_result**      A list containing XINA network analysis results. See [xina\\_analysis](#)

**clustering\_result**  
A list containing XINA clustering results. See [xina\\_clustering](#)

**condition**      Default is 'all', which means use all the proteins to draw graphs. If you specify the experimental condition name used for XINA clustering, xina\_plot\_all will draw graphs using specific condition's proteins.

**centrality\_type**  
'centrality\_type' should be one of c('Degree', 'Eigenvector', 'Hub', 'Authority', 'Closeness', 'Betweenness')

Centrality score	igraph function
Degree	<a href="#">degree</a>
Eigenvector	<a href="#">eigen_centrality</a>
Hub	<a href="#">hub_score</a>
Authority	<a href="#">authority_score</a>
Closeness	<a href="#">closeness</a>
Betweenness	<a href="#">betweenness</a>

**flag\_simplify**    If it is TRUE (default), XINA will exclude unconnected proteins

**num\_breaks**      'num\_breaks' is the number of ranks based on network centrality. Default is 5.

**layout\_specified**  
This can change network layout. 'layout\_specified' should be one of c('sphere', 'star', 'gem', 'tree', 'circle', 'random', 'nicely'). XINA's layouts are based on igraph's layout. See [layout\\_](#)



Layout	igraph layout name
sphere	<a href="#">layout_on_sphere</a>
star	<a href="#">layout_as_star</a>
gem	<a href="#">layout_with_gem</a>
tree	<a href="#">layout_as_tree</a>
circle	<a href="#">layout_in_circle</a>
random	<a href="#">layout_randomly</a>
nicely	<a href="#">layout_nicely</a>

Default is 'layout\_nicely' of igraph

vertex_label_flag	If vertex_label_flag is TRUE (default), igraph network graphs will be labeled by gene names. If vertex_label_flag is FALSE, igraph network graphs will be drawn without labels.
vertex.label.color	Color of labels. Default is black.
vertex.color	Color of nodes. Default is pink.
edge.color	Color of edges. Default is pink.
vertex.label.dist	Distance between node and label. Default is 0.6.
vertex.label.cex	Size of labels. Default is 0.8.
edge.arrow.size	Size of edges. Default is 0.4.
vertex.size	Size of nodes. Default is 10.
vertex.shape	You can choose node shape. Default is 'sphere'. See <a href="#">shapes</a> .
legend_location	If centrality_type is chosen, xina_plot_single add the color legend guiding rank of nodes based on the centrality score. Default is 'bottomright', but you can choose one of these 'bottomright', 'bottom', 'bottomleft', 'left', 'topleft', 'top', 'topright', 'right' and 'center'.
num_clusters_in_row	The number of clusters in a row on the XINA network plot. Default is 5.
flag_unknown_only	If this is TRUE, 'xina_plot_all' will plot proteins that do not have any protein-protein interaction in the given database.
img_size	Set the image size. For width=1000 and height=1500, it is img_size=c(1000,1500). Default is c(3000,3000).
img_qual	Set the image resolution. Default is 300.

### Value

PNG images of PPI network plots of all the clusters

**Examples**

```

## the following code is to show how it works quickly
## load XINA example data
data(xina_example)

## load the previously processed XINA analysis results
# if you want to learn how to run 'xina_analysis', please see \link[XINA]{xina_analysis}
data(xina_result_example)

# XINA network plots
xina_plot_all(xina_result_example, example_clusters)

# XINA network plots for Control condition
xina_plot_all(xina_result_example, example_clusters, condition='Control')

```

---

xina\_plot\_bycluster    *xina\_plot\_bycluster*

---

**Description**

xina\_plot\_bycluster is to draw protein-protein interaction network plots of each cluster

**Usage**

```

xina_plot_bycluster(xina_result, clustering_result, cl = NULL,
  condition = "all", flag_legend = TRUE, centrality_type = NULL,
  flag_simplify = TRUE, layout_specified = "",
  vertex_label_flag = TRUE, vertex.label.dist = 0.6,
  vertex.label.cex = 0.8, edge.arrow.size = 0.4, vertex.size = 10,
  vertex.shape = "sphere", vertex.color = "",
  edge.color = "darkgray", legend_location = "bottom",
  flag_unknown_only = FALSE)

```

**Arguments**

xina_result	A list containing XINA network analysis results. See <a href="#">xina_analysis</a>
clustering_result	A list containing XINA clustering results. See <a href="#">xina_clustering</a>
cl	Cluster number in the XINA clustering results
condition	Default is 'all', which means use all the proteins to draw graphs. If you specify the experimental condition name used for XINA clustering,
flag_legend	If it is TRUE, a legend will be printed out together.
centrality_type	'centrality_type' should be one of c('Degree', 'Eigenvector', 'Hub', 'Authority', 'Closeness', 'Betweenness')

Centrality score	igraph function
Degree	<a href="#">degree</a>
Eigenvector	<a href="#">eigen_centrality</a>
Hub	<a href="#">hub_score</a>
Authority	<a href="#">authority_score</a>
Closeness	<a href="#">closeness</a>
Betweenness	<a href="#">betweenness</a>

`flag_simplify` If it is TRUE (default), XINA will exclude unconnected proteins

`layout_specified`

This can change network layout. 'layout\_specified' should be one of c('sphere', 'star', 'gem', 'tree', 'circle', 'random', 'nicely'). XINA's layouts are based on igraph's layout. See [layout\\_](#)

Layout	igraph layout name
sphere	<a href="#">layout_on_sphere</a>
star	<a href="#">layout_as_star</a>
gem	<a href="#">layout_with_gem</a>
tree	<a href="#">layout_as_tree</a>
circle	<a href="#">layout_in_circle</a>
random	<a href="#">layout_randomly</a>
nicely	<a href="#">layout_nicely</a>

Default is 'layout\_nicely' of igraph

`vertex_label_flag`

If `vertex_label_flag` is TRUE (default), igraph network graphs will be labeled by gene names If `vertex_label_flag` is FALSE, igraph network graphs will be drawn without labels

`vertex.label.dist`

Distance between node and label. Default is 0.6

`vertex.label.cex`

Size of labels Default is 0.8

`edge.arrow.size`

Size of edges Default is 0.4

`vertex.size`

Size of nodes Default is 10

`vertex.shape`

You can choose node shape. Default is 'sphere'. See [shapes](#)

`vertex.color`

Color of nodes. Default is pink.

`edge.color`

Color of edges. Default is pink.

`legend_location`

If `centrality_type` is chosen, `xina_plot_single` add the color legend guiding rank of nodes based on the centrality score. Default is 'bottomright', but you can choose one of these 'bottomright', 'bottom', 'bottomleft', 'left', 'topleft', 'top', 'topright', 'right' and 'center'.

`flag_unknown_only`

If this is TRUE, 'xina\_plot\_bycluster' will plot proteins that do not have any protein-protein interaction in the given database

**Value**

A PNG file (XINA\_Cluster\_Networks.png) displaying protein-protein interaction network plots of all the clusters and a list containing XINA network analysis results

PNG images of PPI network plots of all the clusters

**Examples**

```
## the following code is to show how it works quickly
## load XINA example data
data(xina_example)

## load the previously processed XINA analysis results
# if you want to learn how to run 'xina_analysis', please see \link[XINA]{xina_analysis}
data(xina_result_example)

# plot cluster #1
xina_plot_bycluster(xina_result_example, example_clusters, cl=1)

# plot PPI network of Control condition in cluster #1
xina_plot_bycluster(xina_result_example, example_clusters, cl=1, condition='Control')
```

---

xina\_plot\_single      *xina\_plot\_single*

---

**Description**

xina\_plot\_single draws protein-protein interaction network plot for given 'protein\_list'.

**Usage**

```
xina_plot_single(xina_result, protein_list, centrality_type = NULL,
  layout_specified = "", vertex_label_flag = TRUE, main = NULL,
  vertex.label.color = "black", vertex.color = NA,
  edge.color = "darkgray", vertex.label.dist = 0.6,
  vertex.label.cex = 0.8, edge.arrow.size = 0.4, vertex.size = 10,
  vertex.shape = "sphere", legend_location = "bottom",
  num_breaks = 5, digits_round_up = 5, flag_simplify = TRUE,
  flag_legend = TRUE)
```

**Arguments**

xina\_result      A list containing XINA network analysis results. See [xina\\_analysis](#)

protein\_list     A vector of gene names to draw a protein-protein interaction network graph.

centrality\_type

'centrality\_type' should be one of c('Degree', 'Eigenvector', 'Hub', 'Authority', 'Closeness', 'Betweenness')

Centrality score	igraph function
Degree	<a href="#">degree</a>
Eigenvector	<a href="#">eigen_centrality</a>
Hub	<a href="#">hub_score</a>
Authority	<a href="#">authority_score</a>
Closeness	<a href="#">closeness</a>
Betweenness	<a href="#">betweenness</a>

layout\_specified

This can change network layout. 'layout\_specified' should be one of c('sphere', 'star', 'gem', 'tree', 'circle', 'random', 'nicely'). XINA's layouts are based on igraph's layout. See [layout\\_](#)

Layout	igraph layout name
sphere	<a href="#">layout_on_sphere</a>
star	<a href="#">layout_as_star</a>
gem	<a href="#">layout_with_gem</a>
tree	<a href="#">layout_as_tree</a>
circle	<a href="#">layout_in_circle</a>
random	<a href="#">layout_randomly</a>
nicely	<a href="#">layout_nicely</a>

Default is 'layout\_nicely' of igraph

vertex\_label\_flag

If vertex\_label\_flag is TRUE (default), igraph network graphs will be labeled by gene names. If vertex\_label\_flag is FALSE, igraph network graphs will be drawn without labels.

main

Title of network figure. IF it is NULL (default), it will be the number of plotted proteins.

vertex.label.color

Color of labels. Default is black.

vertex.color

Color of nodes. Default is pink.

edge.color

Color of edges. Default is pink.

vertex.label.dist

Distance between node and label. Default is 0.6.

vertex.label.cex

Size of labels. Default is 0.8.

edge.arrow.size

Size of edges. Default is 0.4.

vertex.size

Size of nodes. Default is 10.

vertex.shape

You can choose node shape. Default is 'sphere'. See [shapes](#)

legend_location	If centrality_type is chosen, 'xina_plot_single' adds the color legend guiding rank of nodes based on the centrality score. Default is 'bottomright', but you can choose one of these 'bottomright', 'bottom', 'bottomleft', 'left', 'topleft', 'top', 'topright', 'right' and 'center'.
num_breaks	'num_breaks' is the number of ranks based on network centrality. Default is 5.
digits_round_up	See <a href="#">Round</a>
flag_simplify	If it is TRUE (default), XINA will exclude unconnected proteins
flag_legend	If it is TRUE, a legend will be printed out together.

### Value

A PNG file (XINA\_Cluster\_Networks.png) displaying protein-protein interaction network plots of all the clusters and a list containing XINA network analysis results

### Examples

```
## the following code is to show how it works quickly
## load XINA example data
data(xina_example)

## load the previously processed XINA analysis results
# if you want to learn how to run 'xina_analysis', please see \link[XINA]{xina_analysis}
data(xina_result_example)

# get gene names that are clustered to #21 in "Stimulus2" condition
subgroup <- subset(example_clusters$aligned, Stimulus2==21)
protein_list <- subgroup$`Gene name`

# Calculate protein-protein interaction network
xina_plot_single(xina_result_example, protein_list)

# Calculate protein-protein interaction network and Eigenvector centrality
eigen_info <- xina_plot_single(xina_result_example, protein_list, centrality_type='Eigenvector')
```

---

xina_result_example	<i>Previously processed xina analysis using XINA's random example data A list containing 'xina_analysis' results</i>
---------------------	--

---

### Description

- All\_network. PPI network of all the input proteins
- Sub\_network. A list containing PPI networks of each clusters
- Data. XINA clustering results. See [xina\\_clustering](#)
- Nodes. A list of proteins in each cluster

- Conditions. A list of experimental condition of proteins in each cluster
- Titles. A list of plot titles for XINA plotting
- out\_dir. A directory path storing XINA network analysis results
- is\_stringdb. False = different PPI DB and TRUE = STRING DB

**Format**

A data frame containing STRING protein-protein interaction data

**Source**

XINA

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