Package 'ChIPseeker'

December 19, 2024

Type Package

Title ChIPseeker for ChIP peak Annotation, Comparison, and Visualization

Version 1.43.0

Maintainer Guangchuang Yu <guangchuangyu@gmail.com>

Description This package implements functions to retrieve the nearest genes around the peak, annotate genomic region of the peak, statstical methods for estimate the significance of overlap among ChIP peak data sets, and incorporate GEO database for user to compare the own dataset with those deposited in database. The comparison can be used to infer cooperative regulation and thus can be used to generate hypotheses. Several visualization functions are implemented to summarize the coverage of the peak experiment, average profile and heatmap of peaks binding to TSS regions, genomic annotation, distance to TSS, and overlap of peaks or genes.

Depends R (>= 3.5.0)

Imports AnnotationDbi, aplot, BiocGenerics, boot, dplyr, enrichplot, IRanges, GenomeInfoDb, GenomicRanges, GenomicFeatures, ggplot2, gplots, graphics, grDevices, gtools, magrittr, methods, plotrix, parallel, RColorBrewer, rlang, rtracklayer, S4Vectors, scales, stats, tibble, TxDb.Hsapiens.UCSC.hg19.knownGene, utils, yulab.utils (>= 0.1.5)

Suggests clusterProfiler, ggimage, ggplotify, ggupset, ggVennDiagram, knitr, org.Hs.eg.db, prettydoc, ReactomePA, rmarkdown, testthat

Remotes GuangchuangYu/enrichplot

URL https://yulab-smu.top/contribution-knowledge-mining/

BugReports https://github.com/YuLab-SMU/ChIPseeker/issues

Encoding UTF-8

VignetteBuilder knitr

ByteCompile true

License Artistic-2.0

biocViews Annotation, ChIPSeq, Software, Visualization,
        MultipleComparison

RoxygenNote 7.3.2

git_url https://git.bioconductor.org/packages/ChIPseeker

git_branch devel

git_last_commit 9e54d40

git_last_commit_date 2024-10-29

Repository Bioconductor 3.21

Date/Publication 2024-12-19

Author Guangchuang Yu [aut, cre] (ORCID:
        <https://orcid.org/0000-0002-6485-8781>),
        Ming Li [ctb],
        Qianwen Wang [ctb],
        Yun Yan [ctb],
        Hervé Pagès [ctb],
        Michael Kluge [ctb],
        Thomas Schwarzl [ctb],
        Zhougeng Xu [ctb],
        Chun-Hui Gao [ctb] (ORCID: <https://orcid.org/0000-0002-1445-7939>)

Contents

ChIPseeker-package | *ChIPseeker: ChIPseeker for ChIP peak Annotation, Comparison, and Visualization*

Description

This package implements functions to retrieve the nearest genes around the peak, annotate genomic region of the peak, statstical methods for estimate the significance of overlap among ChIP peak data sets, and incorporate GEO database for user to compare the own dataset with those deposited in database. The comparison can be used to infer cooperative regulation and thus can be used to generate hypotheses. Several visualization functions are implemented to summarize the coverage of the peak experiment, average profile and heatmap of peaks binding to TSS regions, genomic annotation, distance to TSS, and overlap of peaks or genes.

Author(s)

Maintainer: Guangchuang Yu <guangchuangyu@gmail.com> (ORCID)

Other contributors:

- Ming Li <limiang929@gmail.com> [contributor]

- Qianwen Wang <treywea@gmail.com> [contributor]

- Yun Yan <youryanyun@gmail.com> [contributor]

- Hervé Pagès <hpages.on.github@gmail.com> [contributor]

- Michael Kluge <michael.kluge@bio.ifi.lmu.de> [contributor]

- Thomas Schwarzl <schwarzl@embl.de> [contributor]

- Zhougeng Xu <xuzhougeng@163.com> [contributor]

- Chun-Hui Gao <gaospecial@gmail.com> (ORCID) [contributor]

See Also

Useful links:

- https://yulab-smu.top/contribution-knowledge-mining/

- Report bugs at https://github.com/YuLab-SMU/ChIPseeker/issues

. .

Description

capture name of variable

Usage

```
.(..., .env = parent.frame())
```

Arguments

`...`	expression
`.env`	environment

Value

expression

Examples

```
x <- 1
eval(.(x)[[1]])
```

---

annotatePeak *annotatePeak*

Description

Annotate peaks

Usage

```
annotatePeak(
  peak,
  tssRegion = c(-3000, 3000),
  TxDb = NULL,
  level = "transcript",
  assignGenomicAnnotation = TRUE,
  genomicAnnotationPriority = c("Promoter", "5UTR", "3UTR", "Exon", "Intron",
    "Downstream", "Intergenic"),
  annoDb = NULL,
  addFlankGeneInfo = FALSE,
  flankDistance = 5000,
  sameStrand = FALSE,
```

```
    ignoreOverlap = FALSE,
    ignoreUpstream = FALSE,
    ignoreDownstream = FALSE,
    overlap = "TSS",
    verbose = TRUE,
    columns = c("ENTREZID", "ENSEMBL", "SYMBOL", "GENENAME")
)
```

Arguments

`peak`	peak file or GRanges object
`tssRegion`	Region Range of TSS
`TxDb`	TxDb or EnsDb annotation object
`level`	one of transcript and gene
`assignGenomicAnnotation`	
	logical, assign peak genomic annotation or not
`genomicAnnotationPriority`	
	genomic annotation priority
`annoDb`	annotation package
`addFlankGeneInfo`	
	logical, add flanking gene information from the peaks
`flankDistance`	distance of flanking sequence
`sameStrand`	logical, whether find nearest/overlap gene in the same strand
`ignoreOverlap`	logical, whether ignore overlap of TSS with peak
`ignoreUpstream`	logical, if True only annotate gene at the 3' of the peak.
`ignoreDownstream`	
	logical, if True only annotate gene at the 5' of the peak.
`overlap`	one of 'TSS' or 'all', if overlap="all", then gene overlap with peak will be reported as nearest gene, no matter the overlap is at TSS region or not.
`verbose`	print message or not
`columns`	names of columns to be obtained from database

Value

data.frame or GRanges object with columns of:

all columns provided by input.

annotation: genomic feature of the peak, for instance if the peak is located in 5'UTR, it will annotated by 5'UTR. Possible annotation is Promoter-TSS, Exon, 5' UTR, 3' UTR, Intron, and Intergenic.

geneChr: Chromosome of the nearest gene

geneStart: gene start

geneEnd: gene end

geneLength: gene length

geneStrand: gene strand

geneId: entrezgene ID

distanceToTSS: distance from peak to gene TSS

if annoDb is provided, extra column will be included:

ENSEMBL: ensembl ID of the nearest gene

SYMBOL: gene symbol

GENENAME: full gene name

Author(s)

G Yu

See Also

plotAnnoBar plotAnnoPie plotDistToTSS

Examples

```
## Not run:
require(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
peakfile <- system.file("extdata", "sample_peaks.txt", package="ChIPseeker")
peakAnno <- annotatePeak(peakfile, tssRegion=c(-3000, 3000), TxDb=txdb)
peakAnno

## End(Not run)
```

---

as.data.frame.csAnno   *as.data.frame.csAnno*

Description

convert csAnno object to data.frame

Usage

```
## S3 method for class 'csAnno'
as.data.frame(x, row.names = NULL, optional = FALSE, ...)
```

Arguments

x	csAnno object
row.names	row names
optional	should be omitted.
...	additional parameters

Value

data.frame

Author(s)

Guangchuang Yu <https://guangchuangyu.github.io>

---

as.GRanges	*as.GRanges*

Description

convert csAnno object to GRanges

Usage

```
as.GRanges(x)
```

Arguments

x   csAnno object

Value

GRanges object

Author(s)

Guangchuang Yu <https://guangchuangyu.github.io>

---

check_upstream_and_downstream

check upstream and downstream parameter

Description

check_upstream_and_downstream

Usage

```
check_upstream_and_downstream(upstream, downstream)
```

Arguments

upstream   upstream

downstream   downstream

combine_csAnno *combine_csAnno*

Description

Combine csAnno Object

Usage

```
combine_csAnno(x, ...)
```

Arguments

x	csAnno object
...	csAnno objects

Details

https://github.com/YuLab-SMU/ChIPseeker/issues/157

Value

csAnno object

covplot *covplot*

Description

plot peak coverage

Usage

```
covplot(
  peak,
  weightCol = NULL,
  xlab = "Chromosome Size (bp)",
  ylab = "",
  title = "ChIP Peaks over Chromosomes",
  chrs = NULL,
  xlim = NULL,
  lower = 1,
  fill_color = "black"
)
```

Arguments

`peak`	peak file or GRanges object
`weightCol`	weight column of peak
`xlab`	xlab
`ylab`	ylab
`title`	title
`chrs`	selected chromosomes to plot, all chromosomes by default
`xlim`	ranges to plot, default is whole chromosome
`lower`	lower cutoff of coverage signal
`fill_color`	specify the color/palette for the plot. Order matters

Value

ggplot2 object

Author(s)

G Yu

`csAnno-class`	*Class "csAnno" This class represents the output of ChIPseeker Annotation*

Description

Class "csAnno" This class represents the output of ChIPseeker Annotation

Slots

`anno` annotation

`tssRegion` TSS region

`level` transcript or gene

`hasGenomicAnnotation` logical

`detailGenomicAnnotation` Genomic Annotation in detail

`annoStat` annotation statistics

`peakNum` number of peaks

Author(s)

Guangchuang Yu https://guangchuangyu.github.io

See Also

annotatePeak

downloadGEObedFiles *downloadGEObedFiles*

Description

download all BED files of a particular genome version

Usage

```
downloadGEObedFiles(genome, destDir = getwd())
```

Arguments

genome   genome version

destDir   destination folder

Author(s)

G Yu

downloadGSMbedFiles *downloadGSMbedFiles*

Description

download BED supplementary files of a list of GSM accession numbers

Usage

```
downloadGSMbedFiles(GSM, destDir = getwd())
```

Arguments

GSM   GSM accession numbers

destDir   destination folder

Author(s)

G Yu

---

dropAnno   *dropAnno*

Description

dropAnno

Usage

```
dropAnno(csAnno, distanceToTSS_cutoff = 10000)
```

Arguments

csAnno   output of annotatePeak

distanceToTSS_cutoff

                      distance to TSS cutoff

Details

drop annotation exceeding distanceToTSS_cutoff

Value

csAnno object

Author(s)

Guangchuang Yu

---

enrichAnnoOverlap   *enrichAnnoOverlap*

Description

calcuate overlap significant of ChIP experiments based on their nearest gene annotation

Usage

```
enrichAnnoOverlap(
  queryPeak,
  targetPeak,
  TxDb = NULL,
  pAdjustMethod = "BH",
  chainFile = NULL,
  distanceToTSS_cutoff = NULL
)
```

Arguments

queryPeak	query bed file
targetPeak	target bed file(s) or folder containing bed files
TxDb	TxDb
pAdjustMethod	pvalue adjustment method
chainFile	chain file for liftOver
distanceToTSS_cutoff	
	restrict nearest gene annotation by distance cutoff

Value

data.frame

Author(s)

G Yu

---

enrichPeakOverlap   *enrichPeakOverlap*

Description

calculate overlap significant of ChIP experiments based on the genome coordinations

Usage

```
enrichPeakOverlap(
  queryPeak,
  targetPeak,
  TxDb = NULL,
  pAdjustMethod = "BH",
  nShuffle = 1000,
  chainFile = NULL,
  pool = TRUE,
  mc.cores = detectCores() - 1,
  verbose = TRUE
)
```

Arguments

queryPeak	query bed file or GRanges object
targetPeak	target bed file(s) or folder that containing bed files or a list of GRanges objects
TxDb	TxDb
pAdjustMethod	pvalue adjustment method

nShuffle	shuffle numbers
chainFile	chain file for liftOver
pool	logical, whether pool target peaks
mc.cores	number of cores, see [mclapply](#)
verbose	logical

Value

data.frame

Author(s)

G Yu

---

getAnnoStat	*getAnnoStat*

Description

getting status of annotation

Usage

```
getAnnoStat(x)
```

Arguments

x   csAnno object

---

getBioRegion	*getBioRegion*

Description

prepare a bioregion of selected feature

Usage

```
getBioRegion(
  TxDb = NULL,
  upstream = 1000,
  downstream = 1000,
  by = "gene",
  type = "start_site"
)
```

Arguments

`TxDb`	TxDb
`upstream`	upstream from start site or end site
`downstream`	downstream from start site or end site
`by`	one of 'gene', 'transcript', 'exon', 'intron' , '3UTR' , '5UTR', 'UTR'
`type`	one of "start_site", "end_site", "body"

Details

this function combined previous functions getPromoters(), getBioRegion() and getGeneBody() in order to solve the following issues.

(1) https://github.com/GuangchuangYu/ChIPseeker/issues/16

(2) https://github.com/GuangchuangYu/ChIPseeker/issues/87

The getBioRegion() function can prevoid a region of interest from `txdb` object. There are three kinds of regions, `start_site`, `end_site` and body.

We take transcript region to expain the differences of these three regions. tx: chr1 1000 1400.

body region refers to the 1000-1400bp.

`start_site` region with `upstream` = 100, `downstream` = 100 refers to 900-1100bp.

`end_site` region with `upstream` = 100, `downstream` = 100 refers to 1300-1500bp.

Value

GRanges object

Author(s)

Guangchuang Yu, Ming L

---

getGeneAnno	*getGeneAnno*

Description

get gene annotation, symbol, gene name etc.

Usage

```
getGeneAnno(annoDb, geneID, type, columns)
```

Arguments

`annoDb`	annotation package
`geneID`	query geneID
`type`	gene ID type
`columns`	names of columns to be obtained from database

Value

data.frame

Author(s)

G Yu

---

getGenomicAnnotation   *getGenomicAnnotation*

Description

get Genomic Annotation of peaks

Usage

```
getGenomicAnnotation(
  peaks,
  distance,
  tssRegion = c(-3000, 3000),
  TxDb,
  level,
  genomicAnnotationPriority,
  sameStrand = FALSE
)
```

Arguments

peaks	peaks in GRanges object
distance	distance of peak to TSS
tssRegion	tssRegion, default is -3kb to +3kb
TxDb	TxDb object
level	one of gene or transcript
genomicAnnotationPriority	
	genomic Annotation Priority
sameStrand	whether annotate gene in same strand

Value

character vector

Author(s)

G Yu

getGEOgenomeVersion *getGEOgenomeVersion*

Description

get genome version statistics collecting from GEO ChIPseq data

Usage

```
getGEOgenomeVersion()
```

Value

data.frame

Author(s)

G Yu

getGEOInfo *getGEOInfo*

Description

get subset of GEO information by genome version keyword

Usage

```
getGEOInfo(genome, simplify = TRUE)
```

Arguments

genome	genome version
simplify	simplify result or not

Value

data.frame

Author(s)

G Yu

---

getGEOspecies *getGEOspecies*

Description

accessing species statistics collecting from GEO database

Usage

```
getGEOspecies()
```

Value

data.frame

Author(s)

G Yu

---

getNearestFeatureIndicesAndDistances
getNearestFeatureIndicesAndDistances

Description

get index of features that closest to peak and calculate distance

Usage

```
getNearestFeatureIndicesAndDistances(
  peaks,
  features,
  sameStrand = FALSE,
  ignoreOverlap = FALSE,
  ignoreUpstream = FALSE,
  ignoreDownstream = FALSE,
  overlap = "TSS"
)
```

Arguments

`peaks`	peak in GRanges
`features`	features in GRanges
`sameStrand`	logical, whether find nearest gene in the same strand
`ignoreOverlap`	logical, whether ignore overlap of TSS with peak
`ignoreUpstream`	logical, if True only annotate gene at the 3' of the peak.
`ignoreDownstream`	
	logical, if True only annotate gene at the 5' of the peak.
`overlap`	one of "TSS" or "all"

Value

list

Author(s)

G Yu

---

getPromoters *getPromoters*

Description

prepare the promoter regions

Usage

```
getPromoters(TxDb = NULL, upstream = 1000, downstream = 1000, by = "gene")
```

Arguments

`TxDb`	TxDb
`upstream`	upstream from TSS site
`downstream`	downstream from TSS site
`by`	one of gene or transcript

Value

GRanges object

---

getSampleFiles *getSampleFiles*

Description

get filenames of sample files

Usage

```
getSampleFiles()
```

Value

list of file names

Author(s)

G Yu

---

getTagMatrix *getTagMatrix*

Description

calculate the tag matrix

Usage

```
getTagMatrix(
  peak,
  upstream,
  downstream,
  windows,
  type,
  by,
  TxDb = NULL,
  weightCol = NULL,
  nbin = NULL,
  verbose = TRUE,
  ignore_strand = FALSE
)
```

Arguments

`peak`	peak peak file or GRanges object
`upstream`	the distance of upstream extension
`downstream`	the distance of downstream extension
`windows`	a collection of region
`type`	one of "start_site", "end_site", "body"
`by`	one of 'gene', 'transcript', 'exon', 'intron', '3UTR' , '5UTR', or specified by users
`TxDb`	TxDb or self-made granges object, served as txdb
`weightCol`	column name of weight, default is NULL
`nbin`	the amount of nbines
`verbose`	print message or not
`ignore_strand`	ignore the strand information or not

Details

`getTagMatrix()` function can produce the matrix for visualization. `peak` stands for the peak file. `window` stands for a collection of regions that users want to look into. Users can use `window` to capture the peak of interest. There are two ways to input `window`.

The first way is that users can use getPromoters()/getBioRegion()/makeBioRegionFromGranges() to get `window` and put it into getTagMatrix().

The second way is that users can use getTagMatrix() to call getPromoters()/getBioRegion()/makeBioRegionFromGran In this way users do not need to input `window` parameter but they need to input `txdb`.

`txdb` is a set of packages contained annotation of regions of different genomes. Users can get the regions of interest through specific functions. These specific functions are built in getPromoters()/getBioRegion(). Many regions can not be gain through `txdb`, like insulator and enhancer regions. Users can provide these regions in the form of granges object. These self-made granges object will be passed to `TxDb` parameter and they will be passed to makeBioRegionFromGranges() to produce the `window`. In a word, `TxDb` parameter is a reference information. Users can pass `txdb object` or self-made granges into it.

Details see [getPromoters](#),[getBioRegion](#) and [makeBioRegionFromGranges](#)

`upstream` and `downstream` parameter have different usages:

(1) window parameter is provided,

if type == 'body', upstream and downstream can use to extend the flank of body region.

if type == 'start_site'/'end_site', upstream and downstream do not play a role in getTagMatrix() function.

(2) window parameter is missing,

if type == 'body', upstream and downstream can use to extend the flank of body region.

if type == 'start_site'/'end_site', upstream and downstream refer to the upstream and downstream of the start_site or the end_site.

`weightCol` refers to column in peak file. This column acts as a weight vaule. Details see https://github.com/YuLab-SMU/ChIPseeker/issues/15

`nbin` refers to the number of bins. getTagMatrix() provide a binning method to get the tag matrix.

Value

tagMatrix

---

getTagMatrix.binning.internal

getTagMatrix.binning.internal

Description

calculate the tagMatrix by binning the idea was derived from the function of deeptools https://deeptools.readthedocs.io/en/dev

Usage

```
getTagMatrix.binning.internal(
  peak,
  weightCol = NULL,
  windows,
  nbin = 800,
  upstream = NULL,
  downstream = NULL,
  ignore_strand = FALSE
)
```

Arguments

peak	peak peak file or GRanges object
weightCol	weightCol column name of weight, default is NULL
windows	windows a collection of region with equal or not equal size, eg. promoter region, gene region.
nbin	the amount of nbines needed to be splited and it should not be more than min_body_length
upstream	rel object, NULL or actual number
downstream	rel object, NULL or actual number
ignore_strand	ignore the strand information or not

Value

tagMatrix

---

getTagMatrix.internal *getTagMatrix.internal*

Description

calculate the tag matrix

Usage

```
getTagMatrix.internal(peak, weightCol = NULL, windows, ignore_strand = FALSE)
```

Arguments

peak	peak file or GRanges object
weightCol	column name of weight, default is NULL
windows	a collection of region with equal size, eg. promoter region.
ignore_strand	ignore the strand information or not

Value

tagMatrix

Author(s)

G Yu

---

getTagMatrix2 *getTagMatrix2*

Description

Nested function for getTagMatrix() to deal with multiple windows

Usage

```
getTagMatrix2(
  peak,
  upstream,
  downstream,
  windows_name,
  type,
  by,
  TxDb = NULL,
  weightCol = NULL,
  nbin = NULL,
```

```
  verbose = TRUE,
  ignore_strand = FALSE
)
```

Arguments

`peak`	peak peak file or GRanges object
`upstream`	the distance of upstream extension
`downstream`	the distance of downstream extension
`windows_name`	the names of windows
`type`	one of "start_site", "end_site", "body"
`by`	one of 'gene', 'transcript', 'exon', 'intron', '3UTR' , '5UTR', or specified by users
`TxDb`	TxDb or self-made granges object, served as txdb
`weightCol`	column name of weight, default is NULL
`nbin`	the amount of nbines
`verbose`	print message or not
`ignore_strand`	ignore the strand information or not

Details

This is an internal function.

Value

tagMatrix

`getTagMatrix2.binning.internal`
internal function

Description

internal function

Usage

```
getTagMatrix2.binning.internal(
  peak,
  weightCol = NULL,
  windows,
  windows_name,
  nbin = 800,
  upstream = NULL,
  downstream = NULL,
  ignore_strand = FALSE
)
```

Arguments

`peak`	peak peak file or GRanges object
`weightCol`	column name of weight, default is NULL
`windows`	a collection of region
`windows_name`	the name of windows
`nbin`	the amount of nbines
`upstream`	the distance of upstream extension
`downstream`	the distance of downstream extension
`ignore_strand`	ignore the strand information or not

`getTagMatrix2.internal`

getTagMatrix2.internal

Description

getTagMatrix2.internal

Usage

```
getTagMatrix2.internal(
  peak,
  weightCol = NULL,
  windows,
  windows_name,
  ignore_strand = FALSE
)
```

Arguments

`peak`	peak peak file or GRanges object
`weightCol`	column name of weight, default is NULL
`windows`	a collection of region
`windows_name`	the name of windows
`ignore_strand`	ignore the strand information or not

---

info *Information Datasets*

Description

ucsc genome version, precalcuated data and gsm information

makeBioRegionFromGranges

makeBioRegionFromGranges

Description

make windows from granges object

Usage

```
makeBioRegionFromGranges(gr, by, type, upstream = 1000, downstream = 1000)
```

Arguments

gr	a grange object contain region of interest
by	specify be users, e.g. gene, insulator, enhancer
type	one of "start_site", "end_site", "body"
upstream	upstream from start site or end site, can be NULL if the type == 'body'
downstream	downstream from start site or end site, can be NULL if the type == 'body'

Details

makeBioRegionFromGranges() function can make bioregion from granges object.

The differences between makeBioRegionFromGranges() and getBioRegion() is that getBioRegion() get the region object from txdb object but makeBioRegionFromGranges() get the region from the granges object provided by users. For example, txdb object do not contain insulator or enhancer regions. Users can provide these regions through self-made granges object

There are three kinds of regions, start_site, end_site and body.

We take enhancer region to explain the differences of these three regions. enhancer: chr1 1000 1400.

body region refers to the 1000-1400bp.

start_site region with upstream = 100, downstream = 100 refers to 900-1100bp.

end_site region with upstream = 100, downstream = 100 refers to 1300-1500bp.

In makeBioRegionFromGranges(), upstream and downstream can be NULL if the type == 'body'. by should be specified by users and can not be omitted. by parameter will be used to made labels. type should also be specified.

<https://github.com/YuLab-SMU/ChIPseeker/issues/189>

Value

GRanges object

---

overlap *overlap*

Description

calculate the overlap matrix, which is useful for vennplot

Usage

```
overlap(Sets)
```

Arguments

Sets   a list of objects

Value

data.frame

Author(s)

G Yu

---

peakHeatmap *peakHeatmap*

Description

plot the heatmap of peaks

Usage

```
peakHeatmap(
  peak,
  weightCol = NULL,
  TxDb = NULL,
  upstream = 1000,
  downstream = 1000,
  xlab = "",
  ylab = "",
  title = NULL,
  palette = NULL,
  verbose = TRUE,
  by = "gene",
  type = "start_site",
  nbin = NULL,
```

```
    ignore_strand = FALSE,
    windows,
    ncol = NULL,
    nrow = NULL
)
```

Arguments

peak	peak file or GRanges object
weightCol	column name of weight
TxDb	TxDb object
upstream	upstream position
downstream	downstream position
xlab	xlab
ylab	ylab
title	title
palette	palette to be filled in,details see [scale_colour_brewer](#)
verbose	print message or not
by	one of 'gene', 'transcript', 'exon', 'intron' , '3UTR' , '5UTR', 'UTR'
type	one of "start_site", "end_site", "body"
nbin	the amount of nbines
ignore_strand	ignore the strand information or not
windows	a collection of region
ncol	the ncol of plotting a list of peak
nrow	the nrow of plotting a list of peak

Value

figure

Author(s)

G Yu

peakHeatmap_multiple_Sets

peakHeatmap

Description

plot the heatmap of peaks align to a sets of regions

Usage

```
peakHeatmap_multiple_Sets(
  peak,
  weightCol = NULL,
  TxDb = NULL,
  upstream = 1000,
  downstream = 1000,
  xlab = "",
  ylab = "",
  title = NULL,
  palette = NULL,
  verbose = TRUE,
  by = "gene",
  type = "start_site",
  nbin = NULL,
  ignore_strand = FALSE,
  windows_name = NULL,
  ncol = NULL,
  nrow = NULL,
  facet_label_text_size = 12
)
```

Arguments

peak	peak file or GRanges object
weightCol	column name of weight
TxDb	TxDb object
upstream	upstream position
downstream	downstream position
xlab	xlab
ylab	ylab
title	title
palette	palette to be filled in,details see scale_colour_brewer
verbose	print message or not
by	one of 'gene', 'transcript', 'exon', 'intron' , '3UTR' , '5UTR', 'UTR'

type	one of "start_site", "end_site", "body"
nbin	the amount of nbines
ignore_strand	ignore the strand information or not
windows_name	the name for each window, which will also be showed in the picture as labels
ncol	the ncol of plotting a list of peak
nrow	the nrow of plotting a list of peak
facet_label_text_size	
	the size of facet label text

Value

figure

---

peak_Profile_Heatmap   *peak_Profile_Heatmap*

Description

plot peak heatmap and profile in a picture

Usage

```
peak_Profile_Heatmap(
  peak,
  weightCol = NULL,
  TxDb = NULL,
  upstream = 1000,
  downstream = 1000,
  xlab = "",
  ylab = "",
  title = NULL,
  palette = NULL,
  verbose = TRUE,
  by = "gene",
  type = "start_site",
  nbin = NULL,
  ignore_strand = FALSE,
  windows_name = NULL,
  ncol = NULL,
  nrow = NULL,
  facet_label_text_size = 12,
  conf,
  facet = "row",
  free_y = TRUE,
  height_proportion = 4
)
```

Arguments

peak	peak file or GRanges object
weightCol	column name of weight
TxDb	TxDb object
upstream	upstream position
downstream	downstream position
xlab	xlab
ylab	ylab
title	title
palette	palette to be filled in,details see [scale_colour_brewer](#)
verbose	print message or not
by	one of 'gene', 'transcript', 'exon', 'intron' , '3UTR' , '5UTR', 'UTR'
type	one of "start_site", "end_site", "body"
nbin	the amount of nbines
ignore_strand	ignore the strand information or not
windows_name	the name for each window, which will also be showed in the picture as labels
ncol	the ncol of plotting a list of peak
nrow	the nrow of plotting a list of peak
facet_label_text_size	
	the size of facet label text
conf	confidence interval
facet	one of 'none', 'row' and 'column'
free_y	if TRUE, y will be scaled by AvgProf
height_proportion	
	the proportion of profiling picture and heatmap

---

plotAnnoBar	*plotAnnoBar method generics*

Description

plotAnnoBar method for csAnno instance

Usage

```
plotAnnoBar(
  x,
  xlab = "",
  ylab = "Percentage(%)",
  title = "Feature Distribution",
  ...
)

## S4 method for signature 'list'
plotAnnoBar(
  x,
  xlab = "",
  ylab = "Percentage(%)",
  title = "Feature Distribution",
  ...
)

plotAnnoBar(x, xlab="", ylab='Percentage(%)',title="Feature Distribution", ...)
```

Arguments

x	csAnno instance
xlab	xlab
ylab	ylab
title	title
...	additional paramter

Value

plot

Author(s)

Guangchuang Yu https://guangchuangyu.github.io

---

plotAnnoBar.data.frame

plotAnnoBar.data.frame

Description

plot feature distribution based on their chromosome region

Usage

```
plotAnnoBar.data.frame(
  anno.df,
  xlab = "",
  ylab = "Percentage(%)",
  title = "Feature Distribution",
  categoryColumn
)
```

Arguments

`anno.df`	annotation stats
`xlab`	xlab
`ylab`	ylab
`title`	plot title
`categoryColumn`	category column

Details

plot chromosome region features

Value

bar plot that summarize genomic features of peaks

Author(s)

Guangchuang Yu <https://yulab-smu.top>

See Also

`annotatePeak` `plotAnnoPie`

`plotAnnoPie`   *plotAnnoPie method generics*

Description

plotAnnoPie method for `csAnno` instance

Usage

```
plotAnnoPie(
  x,
  ndigit = 2,
  cex = 0.9,
  col = NA,
  legend.position = "rightside",
  pie3D = FALSE,
  radius = 0.8,
  ...
)

plotAnnoPie(x,ndigit=2,cex=0.9,col=NA,legend.position="rightside",pie3D=FALSE,radius=0.8,...)
```

Arguments

`x`	csAnno instance
`ndigit`	number of digit to round
`cex`	label cex
`col`	color
`legend.position`	
	topright or other.
`pie3D`	plot in 3D or not
`radius`	radius of the pie
`...`	extra parameter

Value

plot

Author(s)

Guangchuang Yu <https://guangchuangyu.github.io>

`plotAnnoPie.csAnno` *plotAnnoPie*

Description

pieplot from peak genomic annotation

Usage

```
plotAnnoPie.csAnno(
  x,
  ndigit = 2,
  cex = 0.8,
  col = NA,
  legend.position = "rightside",
  pie3D = FALSE,
  radius = 0.8,
  ...
)
```

Arguments

x	csAnno object
ndigit	number of digit to round
cex	label cex
col	color
legend.position	
	topright or other.
pie3D	plot in 3D or not
radius	radius of Pie
...	extra parameter

Value

pie plot of peak genomic feature annotation

Author(s)

Guangchuang Yu https://yulab-smu.top

See Also

annotatePeak plotAnnoBar

Examples

```
## Not run:
require(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
peakfile <- system.file("extdata", "sample_peaks.txt", package="chipseeker")
peakAnno <- annotatePeak(peakfile, TxDb=txdb)
plotAnnoPie(peakAnno)

## End(Not run)
```

---

plotAvgProf   *plotAvgProf*

Description

plot the profile of peaks

Usage

```
plotAvgProf(
  tagMatrix,
  xlim,
  xlab = "Genomic Region (5'->3')",
  ylab = "Peak Count Frequency",
  conf,
  facet = "none",
  free_y = TRUE,
  origin_label = "TSS",
  verbose = TRUE,
  ...
)
```

Arguments

tagMatrix	tagMatrix or a list of tagMatrix
xlim	xlim
xlab	x label
ylab	y label
conf	confidence interval
facet	one of 'none', 'row' and 'column'
free_y	if TRUE, y will be scaled by AvgProf
origin_label	label of the center
verbose	print message or not
...	additional parameter

Value

ggplot object

Author(s)

G Yu; Y Yan

---

plotAvgProf.binning *plotAvgProf.binning*

Description

plot the profile of peaks by binning

Usage

```
plotAvgProf.binning(
  tagMatrix,
  xlab = "Genomic Region (5'->3')",
  ylab = "Peak Count Frequency",
  conf,
  facet = "none",
  free_y = TRUE,
  upstream = NULL,
  downstream = NULL,
  label,
  ...
)
```

Arguments

tagMatrix	tagMatrix or a list of tagMatrix
xlab	x label
ylab	y label
conf	confidence interval
facet	one of 'none', 'row' and 'column'
free_y	if TRUE, y will be scaled
upstream	rel object reflects the percentage of flank extension, e.g rel(0.2) integer reflects the actual length of flank extension or TSS region NULL reflects the gene body with no extension
downstream	rel object reflects the percentage of flank extension, e.g rel(0.2) integer reflects the actual length of flank extension or TSS region NULL reflects the gene body with no extension
label	label
...	additional parameter

Value

ggplot object

---

plotAvgProf2   *plotAvgProf*

Description

plot the profile of peaks that align to flank sequences of TSS

Usage

```
plotAvgProf2(
  peak,
  weightCol = NULL,
  TxDb = NULL,
  upstream = 1000,
  downstream = 1000,
  xlab = "Genomic Region (5'->3')",
  ylab = "Peak Count Frequency",
  conf,
  facet = "none",
  free_y = TRUE,
  verbose = TRUE,
  ignore_strand = FALSE,
  ...
)
```

Arguments

peak	peak file or GRanges object
weightCol	column name of weight
TxDb	TxDb object
upstream	upstream position
downstream	downstream position
xlab	xlab
ylab	ylab
conf	confidence interval
facet	one of 'none', 'row' and 'column'
free_y	if TRUE, y will be scaled by AvgProf
verbose	print message or not
ignore_strand	ignore the strand information or not
...	additional parameter

Details

This function is the old function of plotPeakProf2. It can only plot the start site region of gene.

Value

ggplot object

Author(s)

G Yu, Ming L

---

plotDistToTSS   *plotDistToTSS method generics*

Description

plotDistToTSS method for csAnno instance

Usage

```
plotDistToTSS(
  x,
  distanceColumn = "distanceToTSS",
  xlab = "",
  ylab = "Binding sites (%) (5'->3')",
  title = "Distribution of transcription factor-binding loci relative to TSS",
  ...
)

## S4 method for signature 'list'
plotDistToTSS(
  x,
  distanceColumn = "distanceToTSS",
  xlab = "",
  ylab = "Binding sites (%) (5'->3')",
  title = "Distribution of transcription factor-binding loci relative to TSS",
  distanceBreaks = c(0, 1000, 3000, 5000, 10000, 1e+05),
  palette = NULL,
  ...
)

plotDistToTSS(x,distanceColumn="distanceToTSS", xlab="",
ylab="Binding sites (%) (5'->3')",
title="Distribution of transcription factor-binding loci relative to TSS",...)
```

Arguments

x	csAnno instance
distanceColumn	distance column name
xlab	xlab

ylab	ylab
title	title
...	additional parameter
distanceBreaks	breaks of distance, default is 'c(0, 1000, 3000, 5000, 10000, 100000)'
palette	palette name for coloring different distances. Run 'RColorBrewer::display.brewer.all()' to see all applicable values.

Value

plot

Author(s)

Guangchuang Yu <https://guangchuangyu.github.io>

---

plotDistToTSS.data.frame

plotDistToTSS.data.frame

Description

plot feature distribution based on the distances to the TSS

Usage

```
plotDistToTSS.data.frame(
  peakDist,
  distanceColumn = "distanceToTSS",
  distanceBreaks = c(0, 1000, 3000, 5000, 10000, 1e+05),
  palette = NULL,
  xlab = "",
  ylab = "Binding sites (%) (5'->3')",
  title = "Distribution of transcription factor-binding loci relative to TSS",
  categoryColumn = ".id"
)
```

Arguments

peakDist	peak annotation
distanceColumn	column name of the distance from peak to nearest gene
distanceBreaks	default is 'c(0, 1000, 3000, 5000, 10000, 100000)'
palette	palette name for coloring different distances. Run 'RColorBrewer::display.brewer.all()' to see all applicable values.
xlab	x label
ylab	y lable
title	figure title
categoryColumn	category column, default is ".id"

Value

bar plot that summarize distance from peak to TSS of the nearest gene.

Author(s)

Guangchuang Yu https://guangchuangyu.github.io

See Also

annotatePeak

Examples

```
## Not run:
require(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
peakfile <- system.file("extdata", "sample_peaks.txt", package="ChIPseeker")
peakAnno <- annotatePeak(peakfile, TxDb=txdb)
plotDistToTSS(peakAnno)

## End(Not run)
```

---

plotMultiProf   *internal function for plotPeakProf_MultiWindows*

Description

internal function for plotPeakProf_MultiWindows

Usage

```
plotMultiProf(
  tagMatrix,
  conf,
  xlab = "Genomic Region (5'->3')",
  ylab = "Peak Count Frequency",
  facet = "none",
  free_y = TRUE,
  ...
)
```

Arguments

tagMatrix	tagMatrix
conf	confidence interval
xlab	xlab
ylab	ylab

facet	one of 'none', 'row' and 'column'
free_y	if TRUE, y will be scaled by AvgProf
...	additional parameter

---

plotMultiProf.binning *internal function*

Description

internal function

Usage

```
plotMultiProf.binning(
  tagMatrix,
  xlab = "Genomic Region (5'->3')",
  ylab = "Peak Count Frequency",
  conf,
  facet = "none",
  free_y = TRUE,
  upstream = NULL,
  downstream = NULL,
  label,
  ...
)
```

Arguments

tagMatrix	tagMatrix
xlab	xlab
ylab	ylab
conf	confidence interval
facet	one of 'none', 'row' and 'column'
free_y	if TRUE, y will be scaled by AvgProf
upstream	the upstream extension
downstream	the downstream extension
label	the label of the center
...	additional parameter

plotMultiProf.binning.internal
                    internal function

Description

internal function

Usage

```
plotMultiProf.binning.internal(
  tagMatrix,
  conf,
  xlab = "Genomic Region (5'->3')",
  ylab = "Peak Count Frequency",
  facet = "none",
  free_y = TRUE,
  upstream = NULL,
  downstream = NULL,
  label,
  ...
)
```

Arguments

tagMatrix	tagMatrix
conf	confidence interval
xlab	xlab
ylab	ylab
facet	one of 'none', 'row' and 'column'
free_y	if TRUE, y will be scaled by AvgProf
upstream	the upstream extension
downstream	the downstream extension
label	the label of the center
...	additional parameter

---

plotMultiProf.normal   *internal function*

Description

internal function

Usage

```
plotMultiProf.normal(
  tagMatrix,
  xlim,
  xlab = "Genomic Region (5'->3')",
  ylab = "Peak Count Frequency",
  conf,
  facet = "none",
  free_y = TRUE,
  origin_label = "TSS",
  verbose = TRUE,
  ...
)
```

Arguments

tagMatrix	tagMatrix
xlim	xlim
xlab	xlab
ylab	ylab
conf	confidence interval
facet	one of 'none', 'row' and 'column'
free_y	if TRUE, y will be scaled by AvgProf
origin_label	the label of the center
verbose	print message or not
...	additional parameter

---

plotMultiProf.normal.internal
internal function

Description

internal function

Usage

```
plotMultiProf.normal.internal(
  tagMatrix,
  conf,
  xlim = c(-3000, 3000),
  xlab = "Genomic Region (5'->3')",
  ylab = "Peak Count Frequency",
  facet = "row",
  free_y = TRUE,
  origin_label,
  ...
)
```

Arguments

tagMatrix	tagMatrix
conf	confidence interval
xlim	xlim
xlab	xlab
ylab	ylab
facet	one of 'none', 'row' and 'column'
free_y	if TRUE, y will be scaled by AvgProf
origin_label	the label of the center
...	additional parameter

---

plotPeakProf   *plotPeakProf_MultiWindows*

Description

plot the profile of peaks ' plotPeakProf_MultiWindows() is almost the same as plotPeakProf2(), having the main difference of accepting two or more granges objects. Accepting more granges objects can help compare the same peaks in different windows.

Usage

```
plotPeakProf(
  tagMatrix = NULL,
  peak,
  upstream,
  downstream,
  conf,
  by,
  type,
  windows_name = NULL,
  weightCol = NULL,
  TxDb = NULL,
  xlab = "Genomic Region (5'->3')",
  ylab = "Peak Count Frequency",
  facet = "row",
  free_y = TRUE,
  verbose = TRUE,
  nbin = NULL,
  ignore_strand = FALSE,
  ...
)
```

Arguments

tagMatrix	tagMatrix or a list of tagMatrix
peak	peak file or GRanges object
upstream	upstream position
downstream	downstream position
conf	confidence interval
by	feature of interest
type	one of "start_site", "end_site", "body"
windows_name	the name for each window, which will also be showed in the picture as labels
weightCol	column name of weight
TxDb	TxDb object or self-made granges objects
xlab	xlab
ylab	ylab
facet	one of 'none', 'row' and 'column'
free_y	if TRUE, y will be scaled by AvgProf
verbose	print message or not
nbin	the amount of bines
ignore_strand	ignore the strand information or not
...	additional parameter

Details

TxDb parameter can accept txdb object. But many regions can not be obtained by txdb object. In this case, Users can provide self-made granges served the same role as txdb object and pass to TxDb object.

by the features of interest.

(1) if users use txdb, by can be one of 'gene', 'transcript', 'exon', 'intron' , '3UTR' , '5UTR', 'UTR'. These features can be obtained by functions from txdb object.

(2) if users use self-made granges object, by can be everything. Because this by will not pass to functions to get features, which is different from the case of using txdb object. This by is only used to made labels showed in picture.

type means the property of the region. one of the "start site", "end site" and "body".

upstream and downstream parameter have different usages:

(1) if type == 'body', upstream and downstream can use to extend the flank of body region.

(2) if type == 'start_site'/'end_site', upstream and downstream refer to the upstream and downstream of the start_site or the end_site.

weightCol refers to column in peak file. This column acts as a weight value. Details see https://github.com/YuLab-SMU/ChIPseeker/issues/15

nbin refers to the number of bins. getTagMatrix() provide a binning method to get the tag matrix.

There are two ways input a list of window.

(1) Users can input a list of self-made granges objects

(2) Users can input a list of by and only one type. In this way, plotPeakProf_MultiWindows() can made a list of window from txdb object based on by and type.

Warning:

(1) All of these window should be the same type. It means users can only compare a list of "start site"/"end site"/"body region" with the same upstream and downstream.

(2) So it will be only one type and several by.

(3) Users can make window by txdb object or self-made granges object. Users can only choose one of 'gene', 'transcript', 'exon', 'intron' , '3UTR' , '5UTR' or 'UTR' in the way of using txdb object. User can input any by in the way of using self-made granges object.

(4) Users can mingle the by designed for the two ways. plotPeakProf_MultiWindows can accpet the hybrid by. But the above rules should be followed.

https://github.com/YuLab-SMU/ChIPseeker/issues/189

Value

ggplot object

plotPeakProf2   *plotPeakProf2*

Description

plot the profile of peaks automatically

Usage

```
plotPeakProf2(
  peak,
  upstream,
  downstream,
  conf,
  by,
  type,
  weightCol = NULL,
  TxDb = NULL,
  xlab = "Genomic Region (5'->3')",
  ylab = "Peak Count Frequency",
  facet = "none",
  free_y = TRUE,
  verbose = TRUE,
  nbin = NULL,
  ignore_strand = FALSE,
  ...
)
```

Arguments

peak	peak file or GRanges object
upstream	upstream position
downstream	downstream position
conf	confidence interval
by	e.g. 'gene', 'transcript', 'exon' or features of interest(e.g. "enhancer")
type	one of "start_site", "end_site", "body"
weightCol	column name of weight
TxDb	TxDb object, or self-made granges object
xlab	xlab
ylab	ylab
facet	one of 'none', 'row' and 'column'
free_y	if TRUE, y will be scaled by AvgProf
verbose	print message or not

`nbin`	the amount of nbines
`ignore_strand`	ignore the strand information or not
`...`	additional parameter

Details

`peak` stands for the peak file.

`by` the features of interest.

(1) if users use `txdb`, `by` can be one of 'gene', 'transcript', 'exon', 'intron' , '3UTR' , '5UTR', 'UTR'. These features can be obtained by functions from txdb object.

(2) if users use self-made granges object, `by` can be everything. Because this `by` will not pass to functions to get features, which is different from the case of using txdb object. This `by` is only used to made labels showed in picture.

`type` means the property of the region. one of the "start site", "end site" and "body".

`upstream` and `downstream` parameter have different usages:

(1) if `type == 'body'`, `upstream` and `downstream` can use to extend the flank of body region.

(2) if `type == 'start_site'/'end_site'`, `upstream` and `downstream` refer to the upstream and downstream of the start_site or the end_site.

`weightCol` refers to column in peak file. This column acts as a weight vaule. Details see https://github.com/YuLab-SMU/ChIPseeker/issues/15

`nbin` refers to the number of bins, providing a binning method to get the tag matrix.

`TxDb` parameter can accept txdb object. But many regions can not be obtained by txdb object. In this case, Users can provide self-made granges served the same role as txdb object and pass to `TxDb` object.

`plotPeakProf2()` is different from the `plotPeakProf()`. `plotPeakProf2()` do not need to provide `window` parameter, which means `plotPeakProf2()` will call relevent functions to make `window` automatically.

Value

ggplot object

Author(s)

G Yu, Ming Li

plotPeakProf_MultiWindows

plotPeakProf_MultiWindows

Description

plot the profile of peaks in two or more windows

Usage

```
plotPeakProf_MultiWindows(
  peak,
  upstream,
  downstream,
  conf,
  by,
  type,
  windows_name = NULL,
  weightCol = NULL,
  TxDb = NULL,
  xlab = "Genomic Region (5'->3')",
  ylab = "Peak Count Frequency",
  facet = "row",
  free_y = TRUE,
  verbose = TRUE,
  nbin = NULL,
  ignore_strand = FALSE,
  ...
)
```

Arguments

peak	peak file or GRanges object
upstream	upstream position
downstream	downstream position
conf	confidence interval
by	feature of interest
type	one of "start_site", "end_site", "body"
windows_name	the name for each window, which will also be showed in the picture as labels
weightCol	column name of weight
TxDb	TxDb object or self-made granges objects
xlab	xlab
ylab	ylab
facet	one of 'none', 'row' and 'column'

`free_y`	if TRUE, y will be scaled by AvgProf
`verbose`	print message or not
`nbin`	the amount of bines
`ignore_strand`	ignore the strand information or not
`...`	additional parameter

Details

This function comes from <https://github.com/YuLab-SMU/ChIPseeker/issues/189> 'plotPeakProf_MultiWindows()` is almost the same as plotPeakProf2(), having the main difference of accepting two or more granges objects. Accepting more granges objects can help compare the same peaks in different windows.

`TxDb` parameter can accept txdb object. But many regions can not be obtained by txdb object. In this case, Users can provide self-made granges served the same role as txdb object and pass to `TxDb` object.

by the features of interest.

(1) if users use `txdb`, by can be one of 'gene', 'transcript', 'exon', 'intron' , '3UTR' , '5UTR', 'UTR'. These features can be obtained by functions from txdb object.

(2) if users use self-made granges object, by can be everything. Because this by will not pass to functions to get features, which is different from the case of using txdb object. This by is only used to made labels showed in picture.

`type` means the property of the region. one of the "start site", "end site" and "body".

`upstream` and `downstream` parameter have different usages:

(1) if `type == 'body'`, upstream and downstream can use to extend the flank of body region.

(2) if `type == 'start_site'/'end_site'`, upstream and downstream refer to the upstream and downstream of the start_site or the end_site.

`weightCol` refers to column in peak file. This column acts as a weight value. Details see <https://github.com/YuLab-SMU/ChIPseeker/issues/15>

`nbin` refers to the number of bins. getTagMatrix() provide a binning method to get the tag matrix.

There are two ways input a list of window.

(1) Users can input a list of self-made granges objects

(2) Users can input a list of by and only one `type`. In this way, plotPeakProf_MultiWindows() can made a list of window from txdb object based on by and `type`.

Warning:

(1) All of these window should be the same type. It means users can only compare a list of "start site"/"end site"/"body region" with the same upstream and downstream.

(2) So it will be only one `type` and several by.

(3) Users can make window by txdb object or self-made granges object. Users can only choose one of 'gene', 'transcript', 'exon', 'intron' , '3UTR' , '5UTR' or 'UTR' in the way of using txdb object. User can input any by in the way of using self-made granges object.

(4) Users can mingle the by designed for the two ways. `plotPeakProf_MultiWindows` can accpet the hybrid by. But the above rules should be followed.

Value

ggplot object

---

readPeakFile   *readPeakFile*

Description

read peak file and store in data.frame or GRanges object

Usage

```
readPeakFile(peakfile, as = "GRanges", ...)
```

Arguments

peakfile   peak file

as   output format, one of GRanges or data.frame

...   additional parameter (pass to 'utils::read.delim()')

Value

peak information, in GRanges or data.frame object

Author(s)

G Yu

Examples

```
peakfile <- system.file("extdata", "sample_peaks.txt", package="ChIPseeker")
peak.gr <- readPeakFile(peakfile, as="GRanges")
peak.gr
```

---

reexports   *Objects exported from other packages*

Description

These objects are imported from other packages. Follow the links below to see their documentation.

GenomicRanges [GRangesList](#)

ggplot2 [rel](#)

seq2gene   *seq2gene*

Description

annotate genomic regions to genes in many-to-many mapping

Usage

```
seq2gene(seq, tssRegion, flankDistance, TxDb, sameStrand = FALSE)
```

Arguments

seq	genomic regions in GRanges object
tssRegion	TSS region
flankDistance	flanking search radius
TxDb	TranscriptDb object
sameStrand	logical whether find nearest/overlap gene in the same strand

Details

This funciton associates genomic regions with coding genes in a many-to-many mapping. It first maps genomic regions to host genes (either located in exon or intron), proximal genes (located in promoter regions) and flanking genes (located in upstream and downstream within user specify distance).

Value

gene vector

Author(s)

Guangchuang Yu

Examples

```
## Not run:
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
TxDb <- TxDb.Hsapiens.UCSC.hg19.knownGene
file <- getSampleFiles()[[1]] # a bed file
gr <- readPeakFile(file)
genes <- seq2gene(gr, tssRegion=c(-1000, 1000), flankDistance = 3000, TxDb)

## End(Not run)
```

---

show *show method*

Description

show method for `csAnno` instance

Usage

```
show(object)
```

Arguments

object   A csAnno instance

Value

message

Author(s)

Guangchuang Yu <https://guangchuangyu.github.io>

---

shuffle *shuffle*

Description

shuffle the position of peak

Usage

```
shuffle(peak.gr, TxDb)
```

Arguments

peak.gr   GRanges object
TxDb   TxDb

Value

GRanges object

Author(s)

G Yu

tagHeatmap	*tagHeatmap*

Description

plot the heatmap of tagMatrix

Usage

```
tagHeatmap(
  tagMatrix,
  xlab = "",
  ylab = "",
  title = NULL,
  palette = "RdBu",
  nrow = NULL,
  ncol = NULL
)
```

Arguments

tagMatrix	tagMatrix or a list of tagMatrix
xlab	xlab
ylab	ylab
title	title
palette	palette to be filled in,details see scale_colour_brewer
nrow	the nrow of plotting a list of peak
ncol	the ncol of plotting a list of peak

Value

figure

Author(s)

G Yu

---

upsetplot   *upsetplot method*

Description

upsetplot method generics

Usage

```
upsetplot(x, ...)
```

Arguments

x	A csAnno instance
...	additional parameter

Value

plot

Author(s)

Guangchuang Yu <https://guangchuangyu.github.io>

---

vennpie   *vennpie method generics*

Description

vennpie method generics

Usage

```
vennpie(x, r = 0.2, cex = 1.2, ...)

vennpie(x, r = 0.2, cex=1.2, ...)
```

Arguments

x	A csAnno instance
r	initial radius
cex	value to adjust legend
...	additional parameter

Value

plot

Author(s)

Guangchuang Yu <https://guangchuangyu.github.io>

---

vennplot   *vennplot*

Description

plot the overlap of a list of object

Usage

```
vennplot(Sets, by = "gplots", ...)
```

Arguments

Sets   a list of object, can be vector or GRanges object

by   one of gplots, ggVennDiagram or Vennerable

...   extra parameters using ggVennDiagram. Details see ggVennDiagram

Details

There are two ways to plot, which users can specify through 'by'.

The first way is to use 'gplots' packages, by setting 'by = gplots'. This method is default method. The venn plot produced through this way has no color.

The second way is to use 'ggVennDiagram' packages, by setting 'by = ggVennDiagram'. The venn plot produced through this way has colors which can be defined by users using ggplot2 grammar e.g.(scale_fill_distiller()). And users can specify any details, like digital number, text size and showing percentage or not, by inputting '...' extra parameters.

Value

venn plot that summarize the overlap of peaks from different experiments or gene annotation from different peak files.

Author(s)

G Yu

Examples

```
## example not run
## require(TxDb.Hsapiens.UCSC.hg19.knownGene)
## txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
## peakfiles <- getSampleFiles()
## peakAnnoList <- lapply(peakfiles, annotatePeak)
## names(peakAnnoList) <- names(peakfiles)
## genes= lapply(peakAnnoList, function(i) as.data.frame(i)$geneId)
## vennplot(genes)
```

---

vennplot.peakfile   *vennplot.peakfile*

Description

vennplot for peak files

Usage

```
vennplot.peakfile(files, labels = NULL)
```

Arguments

files	peak files
labels	labels for peak files

Value

figure

Author(s)

G Yu

Index