

Introduction to RBM package

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1 Overview

This document provides an introduction to the RBM package. The RBM package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the RBM package computes the moderated t-statistics based on the observed data set for each feature using the `lmFit` and `eBayes` function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).

2 Getting started

The RBM package can be installed and loaded through the following R code. Install the RBM package with:

```
> if (!requireNamespace("BiocManager", quietly=TRUE))
+   install.packages("BiocManager")
> BiocManager::install("RBM")
```

Load the RBM package with:

```
> library(RBM)
```

3 RBM_T and RBM_F functions

There are two functions in the RBM package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The p -values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1),1000,6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata,mydesign,100,0.05)
> summary(myresult)
```

	Length	Class	Mode
<code>ordfit_t</code>	1000	-none-	numeric
<code>ordfit_pvalue</code>	1000	-none-	numeric
<code>ordfit_beta0</code>	1000	-none-	numeric
<code>ordfit_beta1</code>	1000	-none-	numeric
<code>permutation_p</code>	1000	-none-	numeric
<code>bootstrap_p</code>	1000	-none-	numeric

```
> sum(myresult$permutation_p<=0.05)
```

```

[1] 57

> which(myresult$permutation_p<=0.05)

[1] 18 20 21 60 70 75 91 107 193 202 222 230 236 254 269 271 305 309 317
[20] 340 343 404 410 423 424 428 434 445 453 503 506 530 532 552 569 582 585 590
[39] 594 642 655 699 749 757 761 775 807 819 835 845 871 913 915 930 945 964 996

> sum(myresult$bootstrap_p<=0.05)

[1] 10

> which(myresult$bootstrap_p<=0.05)

[1] 91 147 271 357 478 613 642 930 945 980

> permutation_adj_p <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adj_p<=0.05)

[1] 6

> bootstrap_adj_p <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adj_p<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7,0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutatioin_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 6

> which(myresult2$bootstrap_p<=0.05)

[1] 257 353 455 592 827 967

> bootstrap2_adj_p <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adj_p<=0.05)

[1] 0

```

- Examples using the RBM_F function: normdata_F simulates a standardized gene expression data and unifdata_F simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.

```

> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)

              Length Class  Mode
ordfit_t      3000  -none-  numeric
ordfit_pvalue 3000  -none-  numeric
ordfit_beta1  3000  -none-  numeric
permutation_p 3000  -none-  numeric
bootstrap_p   3000  -none-  numeric

> sum(myresult_F$permutation_p[, 1]<=0.05)

[1] 48

> sum(myresult_F$permutation_p[, 2]<=0.05)

[1] 43

> sum(myresult_F$permutation_p[, 3]<=0.05)

[1] 52

> which(myresult_F$permutation_p[, 1]<=0.05)

[1]  20  31  84  94 103 152 166 173 184 188 193 196 236 239 251 257 302 324 338
[20] 342 376 412 415 460 482 494 520 531 539 543 544 560 571 714 723 776 799 807
[39] 829 845 855 868 876 878 885 890 932 947

> which(myresult_F$permutation_p[, 2]<=0.05)

[1]  20  31  84  94 103 152 166 173 192 196 236 239 251 257 289 302 324 342 376
[20] 415 460 482 494 520 531 539 543 544 560 571 723 767 799 829 836 845 855 868
[39] 876 878 912 932 965

> which(myresult_F$permutation_p[, 3]<=0.05)

[1]  31  34  77  84  94 103 152 166 173 196 236 239 251 289 302 312 324 338 342
[20] 376 412 415 460 480 482 494 520 531 539 543 544 560 571 646 714 723 767 776
[39] 799 807 826 829 845 855 857 868 876 878 890 912 932 965

> con1_adjp <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adjp<=0.05/3)

[1] 12

```

```

> con2_adjp <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adjp<=0.05/3)

[1] 2

> con3_adjp <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adjp<=0.05/3)

[1] 8

> which(con2_adjp<=0.05/3)

[1] 31 520

> which(con3_adjp<=0.05/3)

[1] 84 103 342 520 531 723 855 878

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

      Length Class  Mode
ordfit_t      3000  -none- numeric
ordfit_pvalue 3000  -none- numeric
ordfit_beta1  3000  -none- numeric
permutation_p 3000  -none- numeric
bootstrap_p   3000  -none- numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 55

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 53

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 63

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 7 10 49 102 108 117 133 155 161 196 237 253 259 264 279 295 299 316 363
[20] 388 402 441 442 453 456 464 469 479 530 539 570 577 578 597 675 689 725 727
[39] 736 743 751 761 799 807 810 833 849 901 910 933 951 960 975 979 998

```

```

> which(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 49 102 108 117 133 139 155 161 196 237 247 253 264 279 295 299 303 363 388
[20] 391 402 414 453 456 461 464 469 479 529 539 577 578 675 689 719 725 727 743
[39] 761 799 807 810 849 853 884 910 951 957 960 975 979 994 998

> which(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 7 49 90 102 108 117 133 139 155 161 196 237 259 264 295 299 303 316 363
[20] 388 391 393 402 441 442 443 451 453 456 462 464 469 479 529 530 539 570 577
[39] 578 597 606 675 689 725 727 736 743 751 761 799 807 817 849 884 910 943 951
[58] 957 960 975 979 994 998

> con21_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
> sum(con21_adj_p<=0.05/3)

[1] 9

> con22_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
> sum(con22_adj_p<=0.05/3)

[1] 9

> con23_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
> sum(con23_adj_p<=0.05/3)

[1] 14

```

4 Ovarian cancer methylation example using the RBM_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of RBM_T in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the genome-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following codes show the process of generating significant differential DNA methylation loci using the RBM_T function and presenting the results for further validation and investigations.

```

> system.file("data", package = "RBM")

[1] "/tmp/RtmpP1sCTm/Rinst20c425498b8d7c/RBM/data"

> data(ovarian_cancer_methylation)
> summary(ovarian_cancer_methylation)

```

IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]
cg00000292: 1	Min. :0.01058	Min. :0.01187	Min. :0.009103
cg00002426: 1	1st Qu.:0.04111	1st Qu.:0.04407	1st Qu.:0.041543
cg00003994: 1	Median :0.08284	Median :0.09531	Median :0.087042
cg00005847: 1	Mean :0.27397	Mean :0.28872	Mean :0.283729
cg00006414: 1	3rd Qu.:0.52135	3rd Qu.:0.59031	3rd Qu.:0.558575
cg00007981: 1	Max. :0.97069	Max. :0.96937	Max. :0.970155
(Other) :994		NA's :4	

exmdata4[, 2]	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]
Min. :0.01019	Min. :0.01108	Min. :0.01937	Min. :0.01278
1st Qu.:0.04092	1st Qu.:0.04059	1st Qu.:0.05060	1st Qu.:0.04260
Median :0.09042	Median :0.08527	Median :0.09502	Median :0.09362
Mean :0.28508	Mean :0.28482	Mean :0.27348	Mean :0.27563
3rd Qu.:0.57502	3rd Qu.:0.57300	3rd Qu.:0.52099	3rd Qu.:0.52240
Max. :0.96658	Max. :0.97516	Max. :0.96681	Max. :0.95974
	NA's :1		

exmdata8[, 2]
Min. :0.01357
1st Qu.:0.04387
Median :0.09282
Mean :0.28679
3rd Qu.:0.57217
Max. :0.96268

```

> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
> summary(diff_results)

```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(diff_results$ordfit_pvalue<=0.05)
```

```
[1] 47
```

```
> sum(diff_results$permutation_p<=0.05)
```

```
[1] 62
```

```
> sum(diff_results$bootstrap_p<=0.05)
```

```
[1] 37
```

```
> ordfit_adjp <- p.adjust(diff_results$ordfit_pvalue, "BH")  
> sum(ordfit_adjp<=0.05)
```

```
[1] 0
```

```
> perm_adjp <- p.adjust(diff_results$permutation_p, "BH")  
> sum(perm_adjp<=0.05)
```

```
[1] 10
```

```
> boot_adjp <- p.adjust(diff_results$bootstrap_p, "BH")  
> sum(boot_adjp<=0.05)
```

```
[1] 2
```

```
> diff_list_perm <- which(perm_adjp<=0.05)  
> diff_list_boot <- which(boot_adjp<=0.05)  
> sig_results_perm <- cbind(ovarian_cancer_methylation[diff_list_perm, ], diff_results$ordfit_t[diff_list_perm, ])  
> print(sig_results_perm)
```

	IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]	exmdata4[, 2]
19	cg00016968	0.80628480	NA	0.81440820	0.83623180
95	cg00081975	0.03633894	0.04975194	0.06024723	0.05598723
131	cg00121904	0.15449580	0.17949750	0.23608110	0.24354150
237	cg00215066	0.94926640	0.95311870	0.94634910	0.94561120
245	cg00224508	0.04479948	0.04972043	0.04152814	0.04189373
280	cg00260778	0.64319890	0.60488960	0.56735060	0.53150910
764	cg00730260	0.90471270	0.90542290	0.91002680	0.91258610
851	cg00830029	0.58362500	0.59397870	0.64739610	0.67269640
887	cg00862290	0.43640520	0.54047160	0.60786800	0.56325950
928	cg00901493	0.03737166	0.03903724	0.04684618	0.04981432
	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]	exmdata8[, 2]	
19	0.80831380	0.73306440	0.82968340	0.84917800	
95	0.04561792	0.05115624	0.06068253	0.06168212	
131	0.17352980	0.12564280	0.18193170	0.20847670	
237	0.94837410	0.94665570	0.94089070	0.94600090	
245	0.04208405	0.05284988	0.03775905	0.03955271	
280	0.61920530	0.61925200	0.46753250	0.55632410	
764	0.90575890	0.88760470	0.90756300	0.90946790	
851	0.50820240	0.34657470	0.66276570	0.64634510	
887	0.50259740	0.40111730	0.56646700	0.54552980	
928	0.04490690	0.04204062	0.05050039	0.05268215	
	diff_results\$ordfit_t[diff_list_perm]				
19	-2.547097				
95	-2.654324				


```

131          -3.562745
237          1.021426
245          1.494678
280          4.337628
764         -1.560713
851         -2.986319
887         -3.368752
928         -1.982308
diff_results$permutation_p[diff_list_perm]
19          0
95          0
131         0
237         0
245         0
280         0
764         0
851         0
887         0
928         0

```

```

> sig_results_boot <- cbind(ovarian_cancer_methylation[diff_list_boot, ], diff_results$ordfit_t[diff_list_boot, ])
> print(sig_results_boot)

```

```

      IlmnID      Beta exmdata2[, 2] exmdata3[, 2] exmdata4[, 2]
146 cg00134539 0.6110132    0.5332178    0.4599934    0.4678742
979 cg00945507 0.1343225    0.2385460    0.3474976    0.2890334
      exmdata5[, 2] exmdata6[, 2] exmdata7[, 2] exmdata8[, 2]
146    0.6719151    0.6313738    0.4792961    0.4542830
979    0.1184851    0.1665385    0.3071842    0.2662474
diff_results$ordfit_t[diff_list_boot]
146          5.636263
979         -4.968792
diff_results$bootstrap_p[diff_list_boot]
146          0
979          0

```