Package 'dupRadar'

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Type Package

Title Assessment of duplication rates in RNA-Seq datasets

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URL https://www.bioconductor.org/packages/dupRadar, https://ssayols.github.io/dupRadar/index.html

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`analyzeDuprates` *Read in a BAM file and count the tags falling on the features described in the GTF file*

Description

`analyzeDuprates` returns a data.frame with tag counts

Usage

```
analyzeDuprates(
  bam,
  gtf,
  stranded = 0,
  paired = FALSE,
  threads = 1,
  verbose = FALSE,
  tmpDir = ".",
  ...
)
```

Arguments

bam	The bam file containing the duplicate-marked reads
gtf	The gtf file describing the features
stranded	Whether the reads are strand specific
paired	Paired end experiment?
threads	The number of threads to be used for counting
verbose	Whether to output Rsubread messages into the console
tmpDir	Directory for temporary files created by featureCounts (default: ".")
...	Other params sent to featureCounts

Details

This function makes use of the Rsubread package to count tags on the GTF features in different scenarios. The scenarios are the 4 possible combinations of allowing multimappers (yes/no) and duplicate reads (yes/no).

Value

A data.frame with counts on features, with and without taking into account multimappers/duplicated reads

Examples

```
bam <- system.file("extdata",
    "wgEncodeCaltechRnaSeqGm12878R1x75dAlignsRep2V2_duprm.bam",
    package = "dupRadar"
)
gtf <- system.file("extdata", "genes.gtf", package = "dupRadar")
stranded <- 2 # '0' (unstranded), '1' (stranded) and '2' (reverse)
paired <- FALSE
threads <- 4

# call the duplicate marker and analyze the reads
dm <- analyzeDuprates(bam, gtf, stranded, paired, threads)

# use a different directory for temporary files (e.g., for FUSE filesystem issues)
dm <- analyzeDuprates(bam, gtf, stranded, paired, threads, tmpDir = "/tmp")
```

---

bamutilMarkDuplicates *Mark duplicates using bamutil*

Description

bamutilMarkDuplicates Mark duplicated reads from a BAM file by calling bamutil

Usage

```
bamutilMarkDuplicates(bam, out, path, verbose)
```

Arguments

bam	The bam file to mark duplicates from
out	Regular expression describing the transformation on the original filename to get the output filename. By default, a "_duprm" suffix is added before the bam extension
path	Path to the duplicate marker binaries
verbose	Redirect all the program output to the R console

Details

This function is supposed to be called through the `markDuplicates` wrapper

Value

The return code of the system call

`cumulativeDuprateBarplot` *Barplot showing the cumulative read counts fraction*

Description

`cumulativeDuprateBarplot` Barplot showing the cumulative read counts fraction

Usage

```
cumulativeDuprateBarplot(DupMat, stepSize = 0.05, ...)
```

Arguments

DupMat	The duplication matrix calculated by `analyzeDuprates`
stepSize	The size of the windows used for plotting
...	Other params sent to barplot

Details

This function makes a barplot showing the cumulative read counts fraction from the duplication matrix calculated by `analyzeDuprates`.

Value

nothing

Examples

```
# dm is a duplication matrix calculated by analyzeDuprates:
# R> dm <- analyzeDuprates(bamDuprm,gtf,stranded,paired,threads)
attach(dupRadar_examples)

# call the plot
cumulativeDuprateBarplot(DupMat=dm)
```

dm *Duplication matrix of a good RNASeq experiment*

Description

A dataset containing the duplication matrix of a good RNASeq experiment, in terms of duplicates. Comes from the GM12878 SR1x75 replicate 2 from Caltech (UCSC's table Browser name: wgEncodeCaltechRnaSeqGm12878R1x75dAlignsRep2V2)

Usage

```
data(dupRadar_examples)
```

Format

A data frame with 23228 rows and 14 variables

dm.bad *Duplication matrix of a failed RNASeq experiment*

Description

A dataset containing the duplication matrix of a failed RNASeq experiment, containing unusual duplication rate. Comes from the HCT116 PE2x75 replicate 1 from Caltech (UCSC's table Browser name: wgEncodeCaltechRnaSeqHct116R2x75Il200AlignsRep1V2)

Usage

```
data(dupRadar_examples)
```

Format

A data frame with 23228 rows and 14 variables

dupRadar *dupRadar.*

Description

Duplication rate quality control for RNA-Seq datasets.

See Also

Useful links:

- https://www.bioconductor.org/packages/dupRadar
- https://ssayols.github.io/dupRadar/index.html

dupRadar_examples *Example data containing precomputed matrices for two RNASeq experiments*

Description

Precomputed duplication matrices for two RNASeq experiments used as examples of a good and a failed (in terms of high redundancy of reads) experiments. The experiments come from the ENCODE project, as a source of a broad variety of protocols, library types and sequencing facilities.

Usage

```
data(dupRadar_examples)
```

Format

A list with two example duplication matrices

duprateExpBoxplot *Duplication rate ~ total reads per kilobase (RPK) boxplot*

Description

`duprateExpBoxplot` Duplication rate ~ total reads per kilobase (RPK) boxplot

Usage

```
duprateExpBoxplot(DupMat, stepSize = 0.05, ...)
```

Arguments

`DupMat`	The duplication matrix calculated by `analyzeDuprates`
`stepSize`	Expression bin seze for the boxplot
`...`	Other params sent to boxplot

Details

This function makes a boxplot showing the distribution of per gene duplication rate versus the reads per kilobase (RPK) inside gene expression bins.

Value

nothing

Examples

```
# dm is a duplication matrix calculated by analyzeDuprates:
# R> dm <- analyzeDuprates(bamDuprm,gtf,stranded,paired,threads)
attach(dupRadar_examples)

# duprate boxplot
duprateExpBoxplot(DupMat=dm)
```

duprateExpDensPlot *Duplication rate ~ total read count plot*

Description

duprateExpDensPlot Duplication rate ~ total read count plot

Usage

```
duprateExpDensPlot(
  DupMat,
  pal = c("cyan", "blue", "green", "yellow", "red"),
  tNoAlternative = TRUE,
  tRPKM = TRUE,
  tRPKMval = 0.5,
  tFit = TRUE,
  addLegend = TRUE,
  ...
)
```

Arguments

DupMat	The duplication matrix calculated by analyzeDuprates
pal	The color palette to use to display the density
tNoAlternative	Display threshold of 1000 reads per kilobase
tRPKM	Display threshold at a given RPKM level
tRPKMval	The given RPKM level
tFit	Whether to fit the model
addLegend	Whether to add a legend to the plot
...	Other parameters sent to plot()

Details

This function makes a scatter plot showing the per gene duplication rate versus the total read count.

Value

nothing

Examples

```
# dm is a duplication matrix calculated by analyzeDuprates:
# R> dm <- analyzeDuprates(bamDuprm,gtf,stranded,paired,threads)
attach(dupRadar_examples)

# duprate plot
duprateExpDensPlot(DupMat=dm)
```

duprateExpFit *Duplication rate ~ total read count fit model*

Description

duprateExpDensPlot Duplication rate ~ total read count fit model

Usage

```
duprateExpFit(DupMat)
```

Arguments

DupMat	The duplication matrix calculated by analyzeDuprates

Details

Fit a Generalized Linear Model using a logit function between thegene duplication rate and the total read count.

Value

The GLM and the coefficients of the fitted logit function

Examples

```
# dm is a duplication matrix calculated by analyzeDuprates:
# R> dm <- analyzeDuprates(bamDuprm,gtf,stranded,paired,threads)
attach(dupRadar_examples)

# duprate plot
duprateExpFit(DupMat=dm)
```

duprateExpIdentify *Identify genes plotted by* duprateExpPlot

Description

duprateExpIdentify Identify genes plotted by duprateExpPlot

Usage

```
duprateExpIdentify(DupMat, idCol = "ID")
```

Arguments

DupMat	The duplication matrix calculated by analyzeDuprates
idCol	The column from the duplication matrix containing the labels

Details

This function makes a barplot showing the cumulative read counts fraction from the duplication matrix calculated by `analyzeDuprates`.

Value

The identified points. `x` and `y` values match the ones from `duprateExpPlot`

Examples

```
# dm is a duplication matrix calculated by analyzeDuprates:
# R> dm <- analyzeDuprates(bamDuprm,gtf,stranded,paired,threads)
attach(dupRadar_examples)

# call the plot and identify genes
duprateExpPlot(DupMat=dm)
duprateExpIdentify(DupMat=dm)
```

---

duprateExpPlot *Duplication rate ~ total read count plot*

Description

`duprateExpPlot` Duplication rate ~ total read count plot

Usage

```
duprateExpPlot(
  DupMat,
  tNoAlternative = TRUE,
  tRPKM = TRUE,
  tRPKMval = 0.5,
  addLegend = TRUE,
  ...
)
```

Arguments

`DupMat`	The duplication matrix calculated by `analyzeDuprates`
`tNoAlternative`	Display threshold of 1000 reads per kilobase
`tRPKM`	Display threshold at a given RPKM level
`tRPKMval`	The given RPKM level
`addLegend`	Whether to add a legend to the plot
`...`	Other parameters sent to smoothScatter()

Details

This function makes a smooth scatter plot showing the per gene duplication rate versus the total read count.

Value

nothing

Examples

```
# dm is a duplication matrix calculated by analyzeDuprates:
# R> dm <- analyzeDuprates(bamDuprm,gtf,stranded,paired,threads)
attach(dupRadar_examples)

# duprate plot
duprateExpPlot(DupMat=dm)
```

`expressionHist` *Draw histogram with the expression values*

Description

`expressionHist` Draw histogram with the expression values

Usage

```
expressionHist(DupMat, value = "RPK", ...)
```

Arguments

`DupMat`	The duplication matrix calculated by `analyzeDuprates`
`value`	The column from the duplication matrix containing the expression values
`...`	Other parameters sent to hist()

Details

This function draws a histogram of the expression values from the duplication matrix calculated by `analyzeDuprates`.

Value

nothing

Examples

```
# dm is a duplication matrix calculated by analyzeDuprates:
# R> dm <- analyzeDuprates(bamDuprm,gtf,stranded,paired,threads)
attach(dupRadar_examples)

# histogram of expression values for annotation
expressionHist(DupMat=dm)
```

getBinDuplication *Helper function used in* duprateExpBoxplot

Description

getBinDuplication get duplication rate for a subset of the duplication matrix

Usage

```
getBinDuplication(p, stepSize, DupMat)
```

Arguments

p	The vector of bins
stepSize	The window size
DupMat	The duplication matrix calculated by analyzeDuprates

Value

The duplication rate per bin

getBinRpkMean *Helper function used in* duprateExpBoxplot

Description

getBinRpkMean get mean duplication rate per bin

Usage

```
getBinRpkMean(p, stepSize, DupMat)
```

Arguments

p	The vector of bins
stepSize	The window size
DupMat	The duplication matrix calculated by analyzeDuprates

Value

The averaged RPK per bin

getDupMatBin *Helper function used in* getBinDuplication *and* getBinRpkMean

Description

getDupMatBin get a subset of the matrix for values in a specific bin defined by the upper bound p and stepSize

Usage

```
getDupMatBin(p, stepSize = 0.05, value = "allCounts", DupMat)
```

Arguments

p	The vector of bins
stepSize	The window size
value	The column to be subset
DupMat	The duplication matrix calculated by analyzeDuprates

Value

The subseted matrix

getDupMatStats *Report duplication stats on regions*

Description

getDupMatStats Report duplication stats based on the data calculated in the duplication matrix

Usage

```
getDupMatStats(DupMat)
```

Arguments

DupMat	The duplication matrix calculated by analyzeDuprates

Value

A data.frame containing the stats about the number of genes covered (1+ tags) and the number of genes containing duplicates (1+)

Examples

```
# dm is a duplication matrix calculated by analyzeDuprates:
# R> dm <- analyzeDuprates(bamDuprm,gtf,stranded,paired,threads)
attach(dupRadar_examples)

# call the plot and identify genes
getDupMatStats(DupMat=dm)
```

getDynamicRange *Dynamic range*

Description

getDynamicRange Calculate the dynamic range of the RNAseq experiment

Usage

```
getDynamicRange(dm)
```

Arguments

dm	The duplication matrix calculated by analyzeDuprates

Details

This function calculates the dynamic range of the RNAseq eperiment

Value

A list with 2 elements, containing the dynamic range counting all reads and the dynamic range after removing duplicates.

Examples

```
# dm is a duplication matrix calculated by analyzeDuprates:
# R> dm <- analyzeDuprates(bamDuprm,gtf,stranded,paired,threads)
attach(dupRadar_examples)

# calculate the dynamic range
getDynamicRange(dm)
```

getRpkBinReadCountFraction *Helper function used in* readcountExpressionBoxplot

Description

readcountExpressionBoxplot Calculates the fraction of total reads in a vector of bins

Usage

```
getRpkBinReadCountFraction(p, stepSize = stepSize, DupMat = DupMat)
```

Arguments

p	The vector of bins
stepSize	The window size
DupMat	The duplication matrix calculated by analyzeDuprates

Value

The fraction of total reads in a vector of bins

`getRpkCumulativeReadCountFraction` *Helper function used in* `readcountExpressionBoxplot`

Description

`getRpkCumulativeReadCountFraction` get the cumulative read count fraction

Usage

```
getRpkCumulativeReadCountFraction(p, DupMat = DupMat)
```

Arguments

`p`	The vector of bins
`DupMat`	The duplication matrix calculated by `analyzeDuprates`

Value

The cumulative read count fraction

`markDuplicates` *Program dispatchers to mark duplicated reads from a BAM file*

Description

`markDuplicates` Mark duplicated reads from a BAM file by calling widely used tools.

Usage

```
markDuplicates(
  dupremover = c("bamutil", "picard"),
  bam = NULL,
  out = gsub("\\.bam$", "_duprm.bam", bam),
  rminput = TRUE,
  path = ".",
  verbose = TRUE,
  ...
)
```

Arguments

dupremover	The tool to be called. Currently, "picard" and "bamutils" are supported
bam	The bam file to mark duplicates from
out	Regular expression describing the transformation on the original filename to get the output filename. By default, a "_duprm" suffix is added before the bam extension
rminput	Whether to keep the original, non duplicate-marked, bam file
path	Path to the duplicate marker binaries
verbose	Redirect all the program output to the R console
...	Other parameters sent to the caller function

Details

This function works as a wrapper for several tools widely adopted tr mark duplicated reads in a BAM file. Currently, it supports PICARD and BamUtils.

Value

The output filename

Examples

```
## Not run:
bam <- system.file("extdata","sample1Aligned.out.bam",package="dupRadar")
gtf <- "genes.gtf"
stranded <- 2    # '0' (unstranded), '1' (stranded) and '2' (reverse)
paired   <- FALSE
threads  <- 4

# call the duplicate marker and analyze the reads
bamDuprm <- markDuplicates(dupremover="bamutil",bam,
                           path="/opt/bamUtil-master/bin",rminput=FALSE)
dm <- analyzeDuprates(bamDuprm,gtf,stranded,paired,threads)

## End(Not run)
```

---

picardMarkDuplicates *Mark duplicates using Picard tools*

Description

picardMarkDuplicates Mark duplicated reads from a BAM file by calling picard tools

Usage

```
picardMarkDuplicates(bam, out, path, verbose, threads = 1, maxmem = "4g")
```

Arguments

bam	The bam file to mark duplicates from
out	Regular expression describing the transformation on the original filename to get the output filename. By default, a "_duprm" suffix is added before the bam extension
path	Path to the duplicate marker binaries
verbose	Redirect all the program output to the R console
threads	Number of threads to use
maxmem	Max memory assigned to the jvm

Details

This function is supposed to be called through the `markDuplicates` wrapper

Value

The return code of the system call

---

readcountExpBoxplot *Barplot of percentage of reads falling into expression bins*

Description

`readcountExpBoxplot` Barplot of percentage of reads falling into expression bins

Usage

```
readcountExpBoxplot(DupMat, stepSize = 0.05, ...)
```

Arguments

DupMat	The duplication matrix calculated by `analyzeDuprates`
stepSize	The number of bars to be shown
...	Other parameters sent to barplot()

Details

This function makes a barplot of percentage of reads falling into expression bins

Value

nothing Other parameters sent to barplot()

Examples

```
# dm is a duplication matrix calculated by analyzeDuprates:
# R> dm <- analyzeDuprates(bamDuprm,gtf,stranded,paired,threads)
attach(dupRadar_examples)


# barplot of percentage of reads falling into expression bins
readcountExpBoxplot(DupMat=dm)
```

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