Package 'carnation'

November 12, 2026

Title Interactive Exploration & Management of RNA-Seq Analyses

Version 1.1.3

Description Highly interactive & modular shiny app to explore three facets of RNA-Seq analysis: differential expression (DE), functional enrichment and pattern analysis. Several visualizations are implemented to provide a wide-ranging view of data sets. For DE analysis, we provide PCA plot, MA plot, Upset plot & heatmaps, in addition to a highly customizable gene plot. Seven different visualizations are available for functional enrichment analysis, and we also support gene pattern analysis. Genes of interest can be tracked across all modules using the gene scratchpad. In addition, carnation provides an integrated platform to manage multiple projects and user access that can be run on a central server to share with collaborators.

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Encoding UTF-8

Roxygen list(markdown = TRUE)

Imports BiocParallel, colorspace, ComplexUpset, DEGreport, dendextend, DESeq2, dplyr, DT, enrichplot, GeneTonic, ggplot2, ggrepel, heatmaply, htmltools, igraph, methods, MatrixGenerics, plotly, reticulate, RColorBrewer, rintrojs, scales, shiny, shinyBS, shinycssloaders, shinymanager, shinythemes, shinyWidgets, sortable, SummarizedExperiment, tools, utils, viridisLite, visNetwork, yaml

Suggests airway, BiocStyle, GenomicFeatures, goseq, knitr, org.Hs.eg.db, rmarkdown, testthat

VignetteBuilder knitr

URL https://nichd-bspc.github.io/carnation/

BugReports https://github.com/NICHD-BSPC/carnation/issues

biocViews GUI, GeneExpression, Software, ShinyApps, GO, Transcription, Transcriptomics, Visualization, DifferentialExpression, Pathways, GeneSetEnrichment

Config/roxygen2/version 8.0.0

git_url https://git.bioconductor.org/packages/carnation

git_branch devel

git_last_commit 1f53b15

git_last_commit_date 2026-10-28

Repository Bioconductor 3.24

Date/Publication 2026-11-12

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`carnation-package` *carnation*

Description

`carnation` is an interactive Shiny dashboard that makes complex bulk RNA-Seq data more accessible and intuitive, integrating all facets of bulk RNA-Seq analysis using three modules - differential expression analysis, functional enrichment and pattern analysis.

Details

- Deeply explore analysis results from complex experiments using interactive plots.
- Easily keep track of genes of interest using the 'Gene scratchpad'.
- Use fuzzy search to filter and search functional enrichment results.
- Visualize complex patterns using highly customizable gene plot.
- Manage local data in single-user mode or deploy on a server to share with collaborators using in-built user management system.

Main function to run the app: `run_carnation()`

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See Also

Useful links:

- https://nichd-bspc.github.io/carnation/
- Report bugs at https://github.com/NICHD-BSPC/carnation/issues

---

`add.set.column` *Add set column to UpSet plot matrix*

Description

This function adds a column denoting set number to a matrix generated for an upset plot with fromList.with.names()

Usage

```
add.set.column(df)
```

Arguments

`df`	binary matrix where row = genes & columns are gene sets, with 1 indicating that a gene is present is that gene set and vice-versa

Value

data.frame with added set column

Examples

```
# list of genes
lst <- list(group1 = c(a = "gene1", b = "gene2", c = "gene3", d = "gene4"),
            group2 = c(c = "gene3", d = "gene4"))

# binarized matrix with group membership
df <- fromList.with.names(lst)

# matrix with added set column
ldf <- add.set.column(df)
```

`add_metadata` *Add metadata to counts data frame*

Description

Add metadata to counts data frame

Usage

```
add_metadata(df, coldata, exclude.intgroups)
```

Arguments

`df`	data.frame with gene counts
`coldata`	data.frame with metadata
`exclude.intgroups`	metadata columns to ignore

Value

counts data frame with added metadata

Examples

```
library(DESeq2)

# make example DESeq data set
dds <- makeExampleDESeqDataSet()

# extract counts and metadata
df <- assay(dds)
coldata <- colData(dds)

# get gene counts df
counts_df <- get_gene_counts(dds, paste0('gene', seq_len(10)))

# add metadata
counts_df <- add_metadata(counts_df, coldata, exclude.intgroups=NULL)
```

`alluvialmod` *Alluvial plot module*

Description

UI & module to generate alluvial plots.

Usage

```
alluvialUI(id, panel)

alluvialServer(id, obj, res_obj, config)
```

Arguments

`id`	Module id
`panel`	string, can be 'sidebar' or 'main'
`obj`	reactiveValues object containing GeneTonic object
`res_obj`	reactive, dataframe containing enrichment results
`config`	reactive list with config settings

Value

UI returns tagList with plot UI server invisibly returns NULL (used for side effects)

Examples

```
library(shiny)

# get DESeqResults object
data(res_dex, package='carnation')

# get enrichResult object
data(eres_dex, package='carnation')

# convert to GeneTonic object

gt <- GeneTonic::shake_enrichResult(eres_dex)

obj <- reactive({
  list(l_gs = gt$l_gs,
       anno_df = gt$anno_df,
       label = 'comp1')
})

res_obj <- reactive({ res })

config <- reactiveVal(get_config())

# run simple shiny app with plot
if(interactive()){
  shinyApp(
```

```
    ui = fluidPage(
           sidebarPanel(alluvialUI('p', 'sidebar')),
           mainPanel(alluvialUI('p', 'main'))
         ),
    server = function(input, output, session){
               alluvialServer('p', obj, res_obj, config)
             }
  )
}
```

---

check_user_access *Get data areas a user has access to*

Description

This function takes a username and returns a list with two elements:

Usage

```
check_user_access(al, u, admin = "admin")
```

Arguments

al	list with access settings; should have two elements - user_group & data_area
u	user name
admin	Admin user group

Details

user_group: one element vector data_area: vector of data areas

Value

list of user groups and data areas

Examples

```
# save access details to file
home <- Sys.getenv('HOME')

# create carnation data area if it doesn't exist
carnation_home <- file.path(home, 'carnation/data')
if(!dir.exists(carnation_home)) dir.create(carnation_home)

create_access_yaml(user = 'admin',
                   user_group = 'admin',
                   data_area = carnation_home)

# get current user access details
al <- read_access_yaml()

lst <- check_user_access(al, u='admin')
```

cnetmod *Cnetplot module*

Description

UI & module to generate Cnetplots.

Usage

```
cnetPlotUI(id, panel)

cnetPlotServer(id, obj, config)
```

Arguments

id	Module id
panel	string, can be 'sidebar' or 'main'
obj	reactive, dataframe containing enrichment results
config	reactive list with config settings

Value

UI returns tagList with plot UI server invisibly returns NULL (used for side effects)

Examples

```
library(shiny)

# get DESeqResults object
data(res_dex, package='carnation')

obj <- reactive({ res })

config <- reactiveVal(get_config())

# run simple shiny app with plot
if(interactive()){
  shinyApp(
    ui = fluidPage(
           sidebarPanel(cnetPlotUI('p', 'sidebar')),
           mainPanel(cnetPlotUI('p', 'main'))
         ),
    server = function(input, output, session){
               cnetPlotServer('p', obj, config)
             }
  )
}
```

create_access_yaml *Create access yaml*

Description

This function creates an access yaml file. This is primarily intended for the first run.

Usage

```
create_access_yaml(user, user_group, data_area)
```

Arguments

user	User name
user_group	User group
data_area	Path to data area containing RDS files

Value

Invisibly returns NULL. This function is primarily used for its side effect of saving a yaml file with access settings

Examples

```
# save access details to file
home <- Sys.getenv('HOME')

# create carnation data area if it doesn't exist
carnation_home <- file.path(home, 'carnation/data')
if(!dir.exists(carnation_home)) dir.create(carnation_home)

create_access_yaml(user = 'admin',
                   user_group = 'admin',
                   data_area = carnation_home)
```

degmod *Pattern plot module*

Description

Module UI & server to generate pattern plots.

Usage

```
patternPlotUI(id, panel, tab)

patternPlotServer(id, obj, coldata, gene_scratchpad, upset_data, config)
```

Arguments

id	Module id
panel	string, can be 'sidebar' or 'main'
tab	string, if 'plot' show plot settings, if 'table' show table settings; if 'both', show settings for both.
obj	reactiveValues object containing carnation object
coldata	reactiveValues object containing object metadata
gene_scratchpad	reactive containing genes selected in scratchpad
upset_data	reactive containing list with data from upset plot module
config	reactive list with config settings

Value

UI returns tagList with module UI server returns reactive with selected genes for scratchpad updates

Examples

```
library(shiny)
library(DESeq2)

# Create reactive values to simulate app state
oobj <- make_example_carnation_object()

obj <- reactiveValues(
   dds = oobj$dds,
   rld = oobj$rld,
   res = oobj$res,
   all_dds = oobj$all_dds,
   all_rld = oobj$all_rld,
   dds_mapping = oobj$dds_mapping
)

cdata <- lapply(oobj$rld, function(x) colData(x))

coldata <- reactiveValues( all=cdata, curr=cdata )

gene_scratchpad <- reactive({ c('gene1', 'gene2') })
upset_data <- reactive({ list(genes=NULL, labels=NULL) })

config <- reactiveVal(get_config())

shinyApp(
  ui = fluidPage(
         sidebarPanel(
           patternPlotUI('p', 'sidebar', 'both'),
           conditionalPanel(condition = "input.pattern_mode == 'Plot'",
             patternPlotUI('p', 'sidebar', 'plot')
           ),
           conditionalPanel(condition = "input.pattern_mode == 'Table'",
             patternPlotUI('p', 'sidebar', 'table')
           )
         ),
```

```
          mainPanel(
            tabsetPanel(id='pattern_mode',
              tabPanel('Plot',
                patternPlotUI('p', 'plot')
              ), # tabPanel plot

              tabPanel('Cluster membership',
                patternPlotUI('p', 'table')
              ) # tabPanel cluster_membership

            ) # tabsetPanel pattern_mode
          ) # tabPanel pattern_analysis
        ),
  server = function(input, output, session){
            patternPlotServer('deg_plot', obj, coldata,
                              gene_scratchpad, upset_data, config)
          }
)
```

`degpatterns_dex` *A* `degPatterns` *object for differentially expressed genes in the dexamethasone treatment comparison.*

Description

A `degPatterns` object for differentially expressed genes in the dexamethasone treatment comparison.

Format

A `degPatterns` object, generated with the `degPatterns` function from the `DEGreport` package.

Details

This `degPatterns` object was created to test for groups of coexpressed genes in the top 100 differentially expressed genes from the dexamethasone treatment comparison.

Details on how this object has been created are included in the `create_carnation_data.R` script, included in the `scripts` folder of the `Carnation` package.

References

Himes BE, Jiang X, Wagner P, Hu R, Wang Q, Klanderman B, Whitaker RM, Duan Q, Lasky-Su J, Nikolos C, Jester W, Johnson M, Panettieri R Jr, Tantisira KG, Weiss ST, Lu Q. "RNA-Seq Transcriptome Profiling Identifies CRISPLD2 as a Glucocorticoid Responsive Gene that Modulates Cytokine Function in Airway Smooth Muscle Cells." PLoS One. 2014 Jun 13;9(6):e99625. PMID: 24926665. GEO: GSE52778

dendromod *Dendrogram module*

Description

UI & module to generate dendrograms.

Usage

```
dendrogramUI(id, panel)

dendrogramServer(id, obj, config)
```

Arguments

id	Module id
panel	string, can be 'sidebar' or 'main'
obj	reactiveValues object containing GeneTonic object
config	reactive list with config settings

Value

UI returns tagList with plot UI server invisibly returns NULL (used for side effects)

Examples

```
library(shiny)

# get enrichResult object
data(eres_dex, package='carnation')

# convert to GeneTonic object
gt <- GeneTonic::shake_enrichResult(eres_dex)

obj <- reactive({
  list(l_gs = gt$l_gs,
       anno_df = gt$anno_df,
       label = 'comp1')
})

config <- reactiveVal(get_config())

# run simple shiny app with plot
if(interactive()){
  shinyApp(
    ui = fluidPage(
           sidebarPanel(dendrogramUI('p', 'sidebar')),
           mainPanel(dendrogramUI('p', 'main'))
         ),
    server = function(input, output, session){
               dendrogramServer('p', obj, config)
             }
  )
```

```
}
```

`distillmod` *Distilled enrichment map module*

Description

UI & module to generate distill enrichment map plots.

Usage

```
distillPlotUI(id, panel)

distillPlotServer(id, obj, args, config)
```

Arguments

`id`	Module id
`panel`	string, can be 'sidebar' or 'main'
`obj`	reactive containing 'distilled' enrichment results
`args`	reactive, list with plot arguments, 'numcat' (number of categories to plot)
`config`	reactive list with config settings

Value

UI returns tagList with plot UI server returns reactive with number of plotted terms

Examples

```
library(GeneTonic)
library(shiny)

# get DESeqResults object
data(res_dex, package='carnation')

# get enrichResult object
data(eres_dex, package='carnation')

# preprocess & convert to GeneTonic object
eres2 <- GeneTonic::shake_enrichResult(eres_dex)
gt <- enrich_to_genetonic(eres_dex, res_dex)

# get distilled results
df <- distill_enrichment(
        eres2,
        res_dex,
        gt$anno_df,
        n_gs = 10,
        cluster_fun = "cluster_markov"
      )
```

```
# number of plotted terms
args <- reactive({ list(numcat=10) })

config <- reactiveVal(get_config())

# run simple shiny app with plot
if(interactive()){
  shinyApp(
    ui = fluidPage(
           sidebarPanel(distillPlotUI('p', 'sidebar')),
           mainPanel(distillPlotUI('p', 'main'))
         ),
    server = function(input, output, session){
               numcat <- observe({
                 distillPlotServer('p',
                                   reactive({ df }),
                                   args,
                                   config)
               })
             }
  )
}
```

---

dlmod *Download button module*

Description

Module UI & server for download buttons.

Usage

```
downloadButtonUI(id)

downloadButtonServer(id, outplot, plot_type)
```

Arguments

id	Module id
outplot	reactive plot handle
plot_type	reactive/static value used for output filename

Value

UI returns tagList with download button UI. Server invisibly returns NULL (used for side effects).

Examples

```
library(shiny)
library(ggplot2)

# get example object
```

```
obj <- make_example_carnation_object()
res <- as.data.frame(obj$res[[1]])

# make MA plot
p <- ggplot(res, aes(x=baseMean, y=log2foldChange)) +
       geom_point(color='black', alpha=0.5)

outplot <- reactive({ p })

# app with a single button to download a plot
if(interactive()){
  shinyApp(
    ui = fluidPage(
           downloadButtonUI('p')
         ),
    server = function(input, output, session){
               downloadButtonServer('p', outplot, 'maplot')
             }
  )
}
```

---

dummy_genetonic *Make dummy GeneTonic object*

Description

Make dummy GeneTonic object

Usage

```
dummy_genetonic(eres)
```

Arguments

eres	enrichResult object

Value

GeneTonic object

---

emapmod *Enrichment map plot module*

Description

UI & module to generate enrichment map plots.

Usage

```
enrichmapUI(id, panel)

enrichmapServer(id, obj, res_obj, config)
```

Arguments

id	Module id
panel	string, can be 'sidebar' or 'main'
obj	reactiveValues object containing GeneTonic object
res_obj	reactive, dataframe containing enrichment results
config	reactive list with config settings

Value

UI returns tagList with plot UI server invisibly returns NULL (used for side effects)

Examples

```
library(shiny)

# get DESeqResults object
data(res_dex, package='carnation')

# get enrichResult object
data(eres_dex, package='carnation')

# convert to GeneTonic object
gt <- GeneTonic::shake_enrichResult(eres_dex)

obj <- reactive({
  list(l_gs = gt$l_gs,
       anno_df = gt$anno_df,
       label = 'comp1')
})

res_obj <- reactive({ res })

config <- reactiveVal(get_config())

# run simple shiny app with plot
if(interactive()){
  shinyApp(
    ui = fluidPage(
           sidebarPanel(enrichmapUI('p', 'sidebar')),
           mainPanel(enrichmapUI('p', 'main'))
         ),
    server = function(input, output, session){
               enrichmapServer('p', obj, res_obj, config)
             }
  )
}
```

enrich_to_genetonic *Convert enrichResult to GeneTonic object*

Description

This function takes an enrichResult object and DE analysis results and creates a GeneTonic object.

Usage

```
enrich_to_genetonic(enrich, res)
```

Arguments

enrich	enrichResult object
res	data frame with DE analysis results

Value

GeneTonic object

Examples

```
# get enrich & res objects
data(res_dex, package="carnation")
data(eres_dex, package="carnation")

# convert to GeneTonic object
gt <- enrich_to_genetonic(eres_dex, res_dex)
```

eres_cell *An* enrichResult *object for differentially expressed genes in the cell line comparison.*

Description

An enrichResult object for differentially expressed genes in the cell line comparison.

Format

An enrichResult object, generated with the enrichGO function from the clusterProfiler package.

Details

This enrichResult object was created to test for functional enrichment using the GO Biological Process (BP) ontology on the top 100 differentially expressed genes from the cell line comparison.

Details on how this object has been created are included in the create_carnation_data.R script, included in the scripts folder of the Carnation package.

References

Himes BE, Jiang X, Wagner P, Hu R, Wang Q, Klanderman B, Whitaker RM, Duan Q, Lasky-Su J, Nikolos C, Jester W, Johnson M, Panettieri R Jr, Tantisira KG, Weiss ST, Lu Q. "RNA-Seq Transcriptome Profiling Identifies CRISPLD2 as a Glucocorticoid Responsive Gene that Modulates Cytokine Function in Airway Smooth Muscle Cells." PLoS One. 2014 Jun 13;9(6):e99625. PMID: 24926665. GEO: GSE52778

`eres_dex` *An* `enrichResult` *object for differentially expressed genes in the dexamethasone treatment comparison.*

Description

An `enrichResult` object for differentially expressed genes in the dexamethasone treatment comparison.

Format

An `enrichResult` object, generated with the `enrichGO` function from the `clusterProfiler` package.

Details

This `enrichResult` object was created to test for functional enrichment using the GO Biological Process (BP) ontology on the top 100 differentially expressed genes from the dexamethasone treatment comparison.

Details on how this object has been created are included in the `create_carnation_data.R` script, included in the `scripts` folder of the `Carnation` package.

References

Himes BE, Jiang X, Wagner P, Hu R, Wang Q, Klanderman B, Whitaker RM, Duan Q, Lasky-Su J, Nikolos C, Jester W, Johnson M, Panettieri R Jr, Tantisira KG, Weiss ST, Lu Q. "RNA-Seq Transcriptome Profiling Identifies CRISPLD2 as a Glucocorticoid Responsive Gene that Modulates Cytokine Function in Airway Smooth Muscle Cells." PLoS One. 2014 Jun 13;9(6):e99625. PMID: 24926665. GEO: GSE52778

`format_genes` *format gene names to look pretty in table output*

Description

This function works by grouping long lists of genes into groups of a specified size. Each group is collapsed using commas, while groups are separated by spaces so that datatable formatting is tricked into separating space-separated groups and not comma-separated groups

Usage

```
format_genes(g, sep = "\\/", genes.per.line = 6)
```

Arguments

g	vector of gene names
sep	gene name separator
genes.per.line	number of genes to show in a line

Value

vector of gene names prettified for data.table output

Examples

```
# string with genes separated by '/'
g <- "gene1/gene2/gene3/gene4/gene5/gene6/gene7"

gg <- format_genes(g, genes.per.line=3)
```

---

fromList.with.names *Prepare list for UpSet plots, but include rownames*

Description

Prepare list for UpSet plots, but include rownames

Usage

```
fromList.with.names(lst)
```

Arguments

lst	List of sets to compare (same input as to UpSetR::fromList)

Value

data.frame of 1 and 0 showing which genes are in which sets

Examples

```
# list of genes
lst <- list(group1 = c(a = "gene1", b = "gene2", c = "gene3", d = "gene4"),
            group2 = c(c = "gene3", d = "gene4"))

# binarized matrix with group membership
df <- fromList.with.names(lst)
```

funenrichmod *Functional enrichment module*

Description

UI & module to show functional enrichment tables & plots.

Usage

```
enrichUI(id, panel, tab = "none")

enrichServer(id, obj, upset_table, gene_scratchpad, reset_genes, config)
```

Arguments

id	ID string used to match the ID used to call the module UI function
panel	string, can be 'sidebar' or 'main'
tab	string, if 'table' show table settings, if 'plots' show plot settings; if 'compare_results', show comparison settings.
obj	reactiveValues object containing carnation object
upset_table	reactive, data from upset plot module
gene_scratchpad	reactive, genes selected in gene scratchpad
reset_genes	reactive to reset genes in scratchpad
config	reactive list with config settings

Value

UI returns tagList with plot UI server returns reactive with gene selected from functional enrichment tables.

Examples

```
library(shiny)
library(DESeq2)

# Create reactive values to simulate app state
oobj <- make_example_carnation_object()

obj <- reactiveValues(
   dds = oobj$dds,
   rld = oobj$rld,
   res = oobj$res,
   all_dds = oobj$all_dds,
   all_rld = oobj$all_rld,
   dds_mapping = oobj$dds_mapping
)

upset_table <- reactiveValues(tbl=NULL, intersections=NULL, set_labels=NULL)

gene_scratchpad <- reactive({ c('gene1', 'gene2') })
```

```
config <- reactiveVal(get_config())

shinyApp(
  ui = fluidPage(
         sidebarPanel(
           conditionalPanel(condition = "input.func == 'Table'",
             enrichUI('p', 'sidebar', 'table')
           ),
           conditionalPanel(condition = "input.func == 'Plot'",
             enrichUI('p', 'sidebar', 'plot')
           ),
           conditionalPanel(condition = "input.func == 'Compare results'",
             enrichUI('p', 'sidebar', 'compare_results')
           )
         ),
         mainPanel(
             tabsetPanel(id='func',
               tabPanel('Table',
                 enrichUI('p', 'main', 'table')
               ), # tabPanel table

               tabPanel('Plot',
                 enrichUI('p', 'main', 'plot')
               ), # tabPanel plot

               tabPanel('Compare results',
                 enrichUI('p', 'main', 'compare_results')
               ) # tabPanel compare_results

             ) # tabsetPanel func
           ) # tabPanel
         ),
  server = function(input, output, session){
             enrich_data <- enrichServer('p', obj,
                                         upset_table,
                                         gene_scratchpad,
                                         reactive({ FALSE }),
                                         config)
           }
)
```

`fuzzymod` *Fuzzy enrichment map module*

Description

UI & module to generate fuzzy enrichment map plots.

Usage

```
fuzzyPlotUI(id, panel)

fuzzyPlotServer(id, obj, args, config)
```

Arguments

id	Module id
panel	string, can be 'sidebar' or 'main'
obj	reactive containing 'distilled' enrichment results
args	reactive, list with plot arguments, 'numcat' (number of categories to plot)
config	reactive list with config settings

Value

UI returns tagList with plot UI server returns reactive with number of plotted terms

Examples

```
library(shiny)

# get enrichResult object
data(eres_dex, package='carnation')

# preprocess & convert to GeneTonic object
gt <- GeneTonic::shake_enrichResult(eres_dex)

# get distilled results
df <- GeneTonic::gs_fuzzyclustering(gt[seq_len(10),],
        similarity_threshold = 0.35,
        fuzzy_seeding_initial_neighbors = 3,
        fuzzy_multilinkage_rule = 0.5)

# number of plotted terms
args <- reactive({ list(numcat=10) })

config <- reactiveVal(get_config())

# run simple shiny app with plot
if(interactive()){
  shinyApp(
    ui = fluidPage(
           sidebarPanel(fuzzyPlotUI('p', 'sidebar')),
           mainPanel(fuzzyPlotUI('p', 'main'))
         ),
    server = function(input, output, session){
               numcat <- observe({
                 fuzzyPlotServer('p',
                                 reactive({ df }),
                                 args,
                                 config)
               })
             }
  )
}
```

geneplotmod *Gene plot module*

Description

UI & server for module to create gene plot

Usage

```
genePlotUI(id, panel)

genePlotServer(id, obj, coldata, plot_args, config)
```

Arguments

id	Module id
panel	string, can be 'sidebar' or 'main'
obj	reactiveValues object containing carnation object
coldata	reactiveValues object containing object metadata
plot_args	reactive list with 3 elements: 'gene.id' (all gene IDs) & 'gene_scratchpad' (genes selected in scratchpad) & 'comp_all' (selected comparison)
config	reactive list with config settings

Value

UI returns tagList with gene plot UI. Server invisibly returns NULL (used for side effects).

Examples

```
library(shiny)
library(DESeq2)

# Create reactive values to simulate app state
oobj <- make_example_carnation_object()

obj <- reactiveValues(
   dds = oobj$dds,
   rld = oobj$rld,
   res = oobj$res,
   all_dds = oobj$all_dds,
   all_rld = oobj$all_rld,
   dds_mapping = oobj$dds_mapping
)

# Set up coldata structure that the module expects
coldata <- reactiveValues(
  curr = list(
    all_samples = colData(oobj$dds$main),
    main = colData(oobj$dds$main)
  )
)
```

```
plot_args <- reactive({
  list(
    gene.to.plot = c("gene1", "gene2"),
    gene.id = rownames(oobj$dds$main),
    comp_all = "comp1"
  )
})

config <- reactiveVal(get_config())

shinyApp(
  ui = fluidPage(
         sidebarPanel(genePlotUI('p', 'sidebar')),
         mainPanel(genePlotUI('p', 'main'))
       ),
  server = function(input, output, session){
             genePlotServer('p', obj, coldata, plot_args, config)
           }
)
```

`getcountplot` *Create gene plot*

Description

This function creates the gene plot.

Usage

```
getcountplot(
  df,
  intgroup = "group",
  factor.levels,
  title = NULL,
  ylab = "Normalized counts",
  color = "gene",
  nrow = 2,
  ymin = NULL,
  ymax = NULL,
  log = TRUE,
  freey = FALSE,
  trendline = "smooth",
  facet = NULL,
  legend = TRUE,
  boxes = TRUE,
  rotate_x_labels = 30,
  box_dodge = "identity"
)
```

Arguments

df	data.frame with gene counts
intgroup	metadata variable to plot on x-axis
factor.levels	levels of intgroup to show on x-axis
title	title of plot
ylab	y-axis label
color	metadata variable to color by
nrow	number of rows to plot if faceting
ymin	y-axis lower limit
ymax	y-axis upper limit
log	should y-axis be log10-transformed?
freey	should y-axes of faceted plots have independent scales?
trendline	type of trendline to draw
facet	metadata variable to facet by
legend	show legend?
boxes	show boxes?
rotate_x_labels	angle to rotate x-axis labels (default=30)
box_dodge	box position if enabled, can be 'identity' (default) or 'dodge'

Value

ggplot handle

Examples

```
# make example DESeq dataset
dds <- DESeq2::makeExampleDESeqDataSet()

# get gene counts
df <- get_gene_counts(dds, gene = c('gene1', 'gene2'))

# standard gene plot
p <- getcountplot(df, intgroup = "condition", factor.levels = c("A", "B"))

# with genes faceted
p1 <- getcountplot(df, intgroup = "condition", factor.levels = c("A", "B"), facet = "gene")
```

get_access_path *Get path to access yaml file*

Description

This function checks for an environment variable 'CARNATION_ACCESS_YAML' to specify directory to save access yaml. If env variable does not exist uses home directory as save location.

Usage

```
get_access_path()
```

Value

path to access yaml

Examples

```
p <- get_access_path()
```

get_config *Get config*

Description

This function reads the bundled package config and returns it. If a local config yaml exists, only supported user-editable settings are merged into the returned config.

Usage

```
get_config(config_path = NULL)
```

Arguments

config_path	optional path to a local config yaml. If NULL, uses the path returned by get_config_path().

Value

list containing config items

Examples

```
cfg <- get_config()
```

get_config_path *Get path to local config yaml file*

Description

This function checks for an environment variable CARNATION_CONFIG_YAML to specify the local config yaml path. If the variable is not set, a default path in the home directory is used.

Usage

```
get_config_path()
```

Value

path to local config yaml

Examples

```
p <- get_config_path()
```

get_degplot *Plot a degPatterns object*

Description

This function plots a degPatterns object.

Usage

```
get_degplot(
  obj,
  time,
  color = NULL,
  cluster_column = "cluster",
  cluster_to_show,
  x_order,
  points = TRUE,
  boxes = TRUE,
  smooth = "smooth",
  lines = TRUE,
  facet = TRUE,
  prefix_title = "Cluster ",
  genes_to_label = NULL
)
```

Arguments

obj	degPatterns object
time	metadata variable to plot on x-axis
color	variable to color plot
cluster_column	column to use for grouping genes
cluster_to_show	which clusters to show in plot
x_order	order of x-axis values
points	boolean, show samples on plot? Default: TRUE
boxes	boolean, show boxes on plot? Default: TRUE
smooth	what type of trendline to use? can be 'smooth' (default) or 'line'.
lines	show lines joining samples? Default: TRUE
facet	boolean, should plot be faceted? Default: TRUE
prefix_title	string, prefix for facet titles
genes_to_label	genes to label on plot

Value

ggplot handle

Examples

```
# get degpatterns object
data(degpatterns_dex, package = 'carnation')

# get pattern plot
all_clusters <- unique(degpatterns_dex$normalized$cluster)

dp <- get_degplot(degpatterns_dex, time='dex',
                  cluster_to_show=all_clusters,
                  x_order=c('untrt','trt'))
```

---

get_gene_counts *Get read counts for gene*

Description

This is a simple function to obtain read counts for a specified gene, based on the DESeq2::plotCounts function.

Usage

```
get_gene_counts(dds, gene, intgroup = "condition", norm_method = "libsize")
```

Arguments

dds	DESeqDataSet object
gene	gene name vector
intgroup	metadata variable to attach to counts
norm_method	normalization method, can be 'libsize' (default) or 'vst'

Value

data.frame with gene counts

Examples

```
# make example DESeq data set
dds <- DESeq2::makeExampleDESeqDataSet()

# get counts for gene1
gg <- get_gene_counts(dds, 'gene1')
```

---

get_project_name_from_path

Get project name from path

Description

This function takes in a path to an RDS file and returns a string to be used as project name

Usage

```
get_project_name_from_path(
  x,
  depth = 2,
  end_offset = 0,
  staging_dir = "dev",
  fsep = .Platform$file.sep
)
```

Arguments

x	character path to RDS file
depth	integer how many levels below path to look?
end_offset	integer how far from the end of path to end?
staging_dir	name of staging directory
fsep	file separator to split path with

Value

project name parsed from path to object

Examples

```
# path to carnation object
obj_path <- "/path/to/project/test/main.rnaseq.rds"

# parsed project name
get_project_name_from_path(obj_path, depth = 2, end_offset = 0)
```

`get_upset_table` *Generate upset plot table*

Description

Generate upset plot table

Usage

```
get_upset_table(gene.lists, comp_split_pattern = ";")
```

Arguments

`gene.lists`	list with character vectors of gene names
`comp_split_pattern`	character used to separate gene set names

Value

list with upset table elements

Examples

```
lst <- list(group1 = c(a = "gene1", b = "gene2", c = "gene3", d = "gene4"),
            group2 = c(b = "gene2", d = "gene4", e = "gene5"),
            group3 = c(d = "gene4", e = "gene5", f = "gene6"))

df <- get_upset_table(lst)
str(df)
```

`get_y_init` *Get initial y-axis limits*

Description

Get initial y-axis limits

Usage

```
get_y_init(df, y_delta, pseudocount)
```

Arguments

df	data.frame with counts. Must have column 'count'
y_delta	y-axis padding for visualization, must be between 0 and 1
pseudocount	pseudo-count to add to the data.frame

Value

min and max limits for count column, padded for visualization

Examples

```
# make example DESeq dataset
dds <- DESeq2::makeExampleDESeqDataSet()

# get gene counts
df <- get_gene_counts(dds, gene = c('gene1', 'gene2'))

# get y axis limits
get_y_init(df, y_delta = 0.01, pseudocount = 1)
```

---

gs_radar *Radar plot*

Description

This is a copy of gs_radar from GeneTonic where the labels of gene sets are converted to parameters

Usage

```
gs_radar(
  res_enrich,
  res_enrich2 = NULL,
  label1 = "scenario 1",
  label2 = "scenario 2",
  n_gs = 20,
  p_value_column = "gs_pvalue"
)
```

Arguments

res_enrich	GeneTonic object for comparison 1
res_enrich2	GeneTonic object for comparison 2 (default = NULL)
label1	label for comparison 1
label2	label for comparison 2
n_gs	number of gene sets (default = 20)
p_value_column	column to use as p-value (default = 'gs_pvalue')

Value

ggplot handle

Examples

```
library(GeneTonic)

# get DESeqResults object
data(res_dex, package='carnation')

# get enrichResult object
data(eres_dex, package='carnation')

# convert to GeneTonic object
gt <- shake_enrichResult(eres_dex)

# get annotation df
idx <- match(c('gene','symbol'), tolower(colnames(res_dex)))
anno_df <- res_dex[,idx]
colnames(anno_df) <- c('gene_id', 'gene_name')

# add aggregate score columns
gt <- get_aggrscores(gt, res_dex, anno_df)

# make radar plot
p <- gs_radar(gt)
```

---

heatmapmod *Heatmap module*

Description

Module UI & server to generate heatmap.

Usage

```
heatmapUI(id, panel)

heatmapServer(id, obj, coldata, plot_args, gene_scratchpad, config)
```

Arguments

id	Module id
panel	string, can be 'sidebar' or 'main'
obj	reactiveValues object containing carnation object
coldata	reactiveValues object containing object metadata
plot_args	reactive containing 'fdr.thres' (padj threshold), 'fc.thres' (log2FC) & 'upset_data' (list containing data from upset plot module)
gene_scratchpad	reactiveValues object containing genes selected in scratchpad which will be labeled
config	reactive list with config settings

Value

UI returns tagList with heatmap UI. Server invisibly returns NULL (used for side effects).

Examples

```
library(shiny)
library(DESeq2)

# Create reactive values to simulate app state
oobj <- make_example_carnation_object()

obj <- reactiveValues(
   dds = oobj$dds,
   rld = oobj$rld,
   res = oobj$res,
   all_dds = oobj$all_dds,
   all_rld = oobj$all_rld,
   dds_mapping = oobj$dds_mapping
)

cdata <- lapply(oobj$rld, function(x) colData(x))

coldata <- reactiveValues( all=cdata, curr=cdata )

plot_args <- reactive({
  list(
    fdr.thres=0.1,
    fc.thres=0,
    upset_data=list(genes=NULL, labels=NULL)
  )
})

gene_scratchpad <- reactive({ c('gene1', 'gene2') })

config <- reactiveVal(get_config())

shinyApp(
  ui = fluidPage(
         sidebarPanel(heatmapUI('p', 'sidebar')),
         mainPanel(heatmapUI('p', 'sidebar'))
       ),
  server = function(input, output, session){
             heatmapServer('p', obj, coldata,
                           plot_args, gene_scratchpad, config)
           }
)
```

`helpmod` *Help button module*

Description

Module UI & server for help buttons.

Usage

```
helpButtonUI(id)

helpButtonServer(id, ...)
```

Arguments

id	Module id. This also doubles as prefixes for help text files.
...	other params passed to helpModal()

Value

UI returns tagList with help button UI. Server invisibly returns NULL (used for side effects).

Examples

```
library(shiny)

# app with a single help button to show DE summary table details
if(interactive()){
  shinyApp(
    ui = fluidPage(
           helpButtonUI('de_summary_help')
         ),
    server = function(input, output, session){
               helpButtonServer('de_summary_help')
             }
  )
}
```

---

helpModal *Help modal*

Description

This generates a modal dialog that includes text from a markdown file.

Usage

```
helpModal(mdfile, title = NULL, ...)
```

Arguments

mdfile	path to markdown file
title	Title of modal dialog
...	other params passed to modalDialog()

Value

Modal dialog with help documentation.

horizonmod *Horizon plot module*

Description

UI & module to generate horizon plots.

Usage

```
horizonUI(id, panel)

horizonServer(id, obj, config)
```

Arguments

id	Module id
panel	string, can be 'sidebar' or 'main'
obj	reactiveValues object containing two GeneTonic objects
config	reactive list with config settings

Value

UI returns tagList with plot UI server invisibly returns NULL (used for side effects)

Examples

```
library(shiny)

# get enrichResult object
data(eres_dex, package='carnation')

# convert to GeneTonic object
gt <- GeneTonic::shake_enrichResult(eres_dex)

# get second enrichResult object
data(eres_cell, package='carnation')

# convert to GeneTonic object
gt1 <- GeneTonic::shake_enrichResult(eres_cell)

obj <- reactive({
  list(
    obj1 = list(l_gs = gt$l_gs,
             anno_df = gt$anno_df,
             label = 'comp1'),
    obj2 = list(l_gs = gt1$l_gs,
             anno_df = gt1$anno_df,
             label = 'comp2')
  )
})

config <- reactiveVal(get_config())
```

```
# run simple shiny app with plot
if(interactive()){
  shinyApp(
    ui = fluidPage(
           sidebarPanel(horizonUI('p', 'sidebar')),
           mainPanel(horizonUI('p', 'main'))
         ),
    server = function(input, output, session){
               horizonServer('p', obj, config)
             }
  )
}
```

`init_local_config` *Initialize local config*

Description

This function copies the bundled package config to a user-writable local config yaml. This is intended for users who want to customize the supported config settings without editing the installed package.

Usage

```
init_local_config(config_path = get_config_path(), overwrite = FALSE)
```

Arguments

`config_path`	path to the local config yaml to create. Defaults to `get_config_path()`.
`overwrite`	logical indicating whether to overwrite an existing file.

Value

Path to the local config yaml, invisibly.

Examples

```
cfg_out <- tempfile(fileext = ".yaml")
init_local_config(cfg_out)
```

install_carnation *Create carnation python environment*

Description

This function installs 'plotly' and 'kaleido' python packages in an environment to allow PDF downloads from plotly plots.

Usage

```
install_carnation(envname, ...)
```

Arguments

envname	name of the python environment
...	parameters passed to reticulate::py_install

Value

NULL, invisibly. The function is called for its side effects.

Examples

```
if(interactive()){
    install_carnation()
}
```

in_admin_group *is user is in admin group?*

Description

is user is in admin group?

Usage

```
in_admin_group(u)
```

Arguments

u	username

Value

TRUE/FALSE to indicate if the user is part of the admin group

Examples

```
# save access details to file
home <- Sys.getenv('HOME')

# create carnation data area if it doesn't exist
carnation_home <- file.path(home, 'carnation/data')
if(!dir.exists(carnation_home)) dir.create(carnation_home)

create_access_yaml(user = 'admin',
                   user_group = 'admin',
                   data_area = carnation_home)

check <- in_admin_group('user')
```

---

is_site_admin *is user an admin?*

Description

is user an admin?

Usage

```
is_site_admin(u)
```

Arguments

u	username

Value

boolean to indicate is user is in admin group

Examples

```
# check if default user is admin
yy <- is_site_admin(u='admin')
```

---

is_valid_pattern_obj *Validate Pattern Analysis Object Schema*

Description

Validate the schema for a single `degpatterns` analysis element used by the pattern analysis module.

Usage

```
is_valid_pattern_obj(pattern_obj, require_symbol = FALSE)
```

Arguments

pattern_obj	A single pattern analysis element. Must be either a data.frame or a list containing a normalized data.frame.
require_symbol	Logical, if TRUE require a symbol column in the analysis table.

Value

Returns TRUE when validation succeeds, otherwise returns FALSE after emitting a message describing the issue.

Examples

```
data(degpatterns_dex, package = "carnation")

is_valid_pattern_obj(degpatterns_dex)
```

---

loadmod *Load data module*

Description

Module UI & server to load new data

Usage

```
loadDataUI(id)

loadDataServer(id, username, config, rds = NULL)
```

Arguments

id	Module id
username	user name
config	reactive list with config settings
rds	Object to be edited

Value

UI returns tagList with module UI Server returns reactive with app reload trigger

Examples

```
library(shiny)

username <- 'admin'

config <- reactiveVal(get_config())

obj <- make_example_carnation_object()
```

```
rds <- reactive({ obj=obj })

shinyApp(
  ui = fluidPage(
         loadDataUI('p')
       ),
  server = function(input, output, session){
             loadDataServer('p', username=username, config, rds)
           }
)
```

`makeEnrichResult` *Make an enrichResult obj from a data frame*

Description

Most of the parameters are just placeholders and the dataframe must contain the columns 'ID' and 'geneID'

Usage

```
makeEnrichResult(
  df,
  split = "/",
  keytype = "UNKNOWN",
  ontology = "UNKNOWN",
  type = "enrichResult"
)
```

Arguments

`df`	data frame with functional enrichment results
`split`	string, character used to split gene IDs
`keytype`	type of gene ID
`ontology`	ontology database being used
`type`	string, can be 'enrichResult' or 'gseaResult'

Value

enrichResult object

Examples

```
# get enrichResult object
data(eres_dex, package='carnation')

# extract the results
df <- as.data.frame(eres_dex)

# convert to a stripped down enrichResult object
eres2 <- makeEnrichResult(df)
```

make_example_carnation_object *Make example carnation object*

Description

Returns example carnation object used in examples & testing

Usage

```
make_example_carnation_object()
```

Value

reactiveValues object containing carnation object

Examples

```
obj <- make_example_carnation_object()
```

make_final_object *Make final object for internal use by the app*

Description

This function takes an uploaded object and sanitizes it to make sure it is suitable for internal use along with other additions:

- adds a 'dds_mapping' element that maps dds_list keys to res_list objects.
- if there are multiple dds_list objects, it adds a 'all_dds' element combining all samples.

Usage

```
make_final_object(obj)
```

Arguments

obj	list object containing lists of DE analysis results, functional enrichment objects, pattern analysis objects & raw and normalized counts objects.

Value

final carnation object with additional pre-processing

Examples

```
library(DESeq2)

# make example DESeq dataset
dds <- makeExampleDESeqDataSet()

# run DE analysis
dds <- DESeq(dds)

# extract comparison of interest
res <- results(dds, contrast = c("condition", "A", "B"))

# perform VST normalization
rld <- varianceStabilizingTransformation(dds, blind = TRUE)

# build minimal object
obj <- list(
           res_list = list(
                          comp = list(
                              res = res,
                              dds = "main",
                              label = "A vs B"
                          )
                      ),
           dds_list = list(main = dds),
           rld_list = list(main = rld)
       )

# final object
final_obj <- make_final_object(obj)
```

`maplotmod` *MA plot module*

Description

UI & server for module to create MA plot

Usage

```
maPlotUI(id, panel)

maPlotServer(id, obj, plot_args, config)
```

Arguments

`id`	Module id
`panel`	string, can be 'sidebar' or 'main'
`obj`	reactiveValues object containing carnation object
`plot_args`	reactive containing 'fdr.thres' (padj threshold), 'fc.thres' (log2FC threshold) & 'gene.to.plot' (genes selected in scratchpad)
`config`	reactive list with config settings

Value

UI returns tagList with MA plot UI. Server invisibly returns NULL (used for side effects).

Examples

```
library(shiny)
library(DESeq2)

# Create reactive values to simulate app state
oobj <- make_example_carnation_object()

obj <- reactiveValues(
   dds = oobj$dds,
   rld = oobj$rld,
   res = oobj$res,
   all_dds = oobj$all_dds,
   all_rld = oobj$all_rld,
   dds_mapping = oobj$dds_mapping
)

# Set up coldata structure that the module expects
coldata <- reactiveValues(
  curr = list(
    all_samples = colData(oobj$dds$main),
    main = colData(oobj$dds$main)
  )
)

plot_args <- reactive({
  list(
    fdr.thres=0.1,
    fc.thres=0,
    gene.to.plot=c('gene1', 'gene2')
  )
})

config <- reactiveVal(get_config())

shinyApp(
  ui = fluidPage(
         sidebarPanel(maPlotUI('p', 'sidebar')),
         mainPanel(maPlotUI('p', 'main'))
       ),
  server = function(input, output, session){
             maPlotServer('p', obj, plot_args, config)
           }
)
```

`materialize_carnation_object` *Materialize expensive carnation object components*

Description

This function materializes expensive derived pieces for a validated carnation object, including DESeqDataSet creation from raw count matrices, variance-stabilized counts, and GeneTonic conversions.

Usage

```
materialize_carnation_object(obj, config = NULL, cores = NULL)
```

Arguments

obj	A validated object returned by validate_carnation_object() or validate_loaded_carnation_o
config	Optional config list. If NULL, will use get_config().
cores	Optional number of worker processes. If NULL, uses config$server$cores.

Value

The input object with materialized dds_list, rld_list, and optional genetonic slots.

Examples

```
# Minimal example with DE results and counts
library(DESeq2)

# Create example data
dds <- makeExampleDESeqDataSet()
dds <- DESeq(dds)
res <- results(dds, contrast = c("condition", "A", "B"))
rld <- varianceStabilizingTransformation(dds, blind = TRUE)

# Validate object inputs
obj <- validate_carnation_object(
  res_list = list(
    comp1 = list(
      res = as.data.frame(res),
      dds = "main",
      label = "A vs B"
    )
  ),
  dds_list = list(main = dds),
  rld_list = list(main = rld)
)

materialized <- materialize_carnation_object(obj, cores = 1)
```

---

metamod *Metadata module*

Description

This module generates the metadata tab that allows users to view the metadata associated with the loaded carnation object.

Usage

```
metadataUI(id, panel)

metadataServer(id, obj, cols.to.drop)
```

Arguments

id	Module id
panel	context for generating ui elements ('sidebar' or 'main')
obj	reactiveValues object containing carnation object
cols.to.drop	columns to hide from table

Value

UI returns tagList with metadata UI. Server returns reactive object with metadata.

Examples

```
library(shiny)

# Create reactive values to simulate app state
oobj <- make_example_carnation_object()

obj <- reactiveValues(
   dds = oobj$dds,
   rld = oobj$rld,
   res = oobj$res,
   all_dds = oobj$all_dds,
   all_rld = oobj$all_rld,
   dds_mapping = oobj$dds_mapping
)

config <- get_config()
cols.to.drop <- config$server$cols.to.drop

shinyApp(
  ui = fluidPage(
         sidebarPanel(metadataUI('p', 'sidebar')),
         mainPanel(metadataUI('p', 'main'))
       ),
  server = function(input, output, session){
             # reactiveVal to save updates
             saved_data <- reactiveVal()

             cdata <- metadataServer('p', obj, cols.to.drop)

             observeEvent(cdata(), {
               saved_data(cdata())
             })
           }
)
```

`my.summary` *Summarize DESeq2 results into a dataframe*

Description

summary(res) prints out info; this function captures it into a dataframe

Usage

```
my.summary(res, dds, alpha, lfc.thresh = 0)
```

Arguments

`res`	DESeq2 results object
`dds`	DEseq2 object
`alpha`	Alpha level at which to call significantly changing genes
`lfc.thresh`	log2FoldChange threshold

Value

Dataframe of summarized results

Examples

```
n_genes <- 100

#  make mock dds list
dds <- DESeq2::makeExampleDESeqDataSet(n=n_genes)

# make mock results df
res <- data.frame(
         baseMean = runif(n_genes, 10, 1000),
         log2FoldChange = rnorm(n_genes, 0, 2),
         lfcSE = runif(n_genes, 0.1, 0.5),
         stat = rnorm(n_genes, 0, 3),
         pvalue = runif(n_genes, 0, 1),
         padj = runif(n_genes, 0, 1),
         symbol = paste0("GENE", 1:n_genes),
         row.names = paste0("gene", 1:n_genes)
       )

# get summary
df <- my.summary(res, dds, alpha=0.1)
```

pcamod *PCA plot module*

Description

Module UI + server to generate a pca plot.

Usage

```
pcaPlotUI(id, panel)

pcaPlotServer(id, obj, coldata, config)
```

Arguments

id	Module id
panel	string, can be 'sidebar' or 'main'
obj	reactiveValues object containing carnation object
coldata	reactiveValues object containing object metadata
config	reactive list with config settings

Value

UI returns tagList with PCA plot UI. Server invisibly returns NULL (used for side effects).

Examples

```
library(shiny)
library(DESeq2)

# Create reactive values to simulate app state
oobj <- make_example_carnation_object()

obj <- reactiveValues(
   dds = oobj$dds,
   rld = oobj$rld,
   res = oobj$res,
   all_dds = oobj$all_dds,
   all_rld = oobj$all_rld,
   dds_mapping = oobj$dds_mapping
)

# Set up coldata structure that the module expects
coldata <- reactiveValues(
  curr = list(
    all_samples = colData(oobj$dds$main),
    main = colData(oobj$dds$main)
  )
)

config <- reactiveVal(get_config())
```

```
shinyApp(
  ui = fluidPage(
         sidebarPanel(pcaPlotUI('p', 'sidebar')),
         mainPanel(pcaPlotUI('p', 'main'))
       ),
  server = function(input, output, session){
             pcaPlotServer('p', obj, coldata, config)
           }
)
```

`plotMA.label` *Create a labeled MA plot*

Description

This function creates an MA plot from a data.frame containing DE analysis results.

Usage

```
plotMA.label(
  res,
  fdr.thres = 0.01,
  fc.thres = 0,
  fc.lim = NULL,
  lab.genes = NULL,
  opacity = 0.8,
  tolower.cols = c("SYMBOL", "ALIAS")
)
```

Arguments

`res`	data.frame with DE analysis results. Must contain "padj" & "log2FoldChange" columns
`fdr.thres`	False discovery rate (FDR) threshold
`fc.thres`	log2FoldChange threshold
`fc.lim`	y-axis limits
`lab.genes`	genes to label on MA plot
`opacity`	point opacity between 0 and 1
`tolower.cols`	column names that will be converted to lower case

Value

ggplot handle

Examples

```
# make mock results df
n_genes <- 100
res <- data.frame(
         baseMean = runif(n_genes, 10, 1000),
         log2FoldChange = rnorm(n_genes, 0, 2),
         lfcSE = runif(n_genes, 0.1, 0.5),
         stat = rnorm(n_genes, 0, 3),
         pvalue = runif(n_genes, 0, 1),
         padj = runif(n_genes, 0, 1),
         symbol = paste0("GENE", 1:n_genes),
         row.names = paste0("gene", 1:n_genes)
       )

plotMA.label(res, lab.genes = c("gene1", "gene2"))
```

---

plotMA.label_ly *Create an interactive labeled MA plot*

Description

This function creates an MA plot from a data.frame containing DE analysis results using plot_ly

Usage

```
plotMA.label_ly(
  res,
  fdr.thres = 0.01,
  fc.thres = 0,
  fc.lim = NULL,
  lab.genes = NULL,
  opacity = 0.3,
  tolower.cols = c("SYMBOL", "ALIAS")
)
```

Arguments

res	data.frame with DE analysis results. Must contain "padj" & "log2FoldChange" columns
fdr.thres	False discovery rate (FDR) threshold
fc.thres	log2FoldChange threshold
fc.lim	y-axis limits
lab.genes	genes to label on MA plot
opacity	point opacity between 0 and 1
tolower.cols	column names that will be converted to lower case

Value

plotly handle

Examples

```
# make mock results df
n_genes <- 100
res <- data.frame(
         baseMean = runif(n_genes, 10, 1000),
         log2FoldChange = rnorm(n_genes, 0, 2),
         lfcSE = runif(n_genes, 0.1, 0.5),
         stat = rnorm(n_genes, 0, 3),
         pvalue = runif(n_genes, 0, 1),
         padj = runif(n_genes, 0, 1),
         symbol = paste0("GENE", 1:n_genes),
         row.names = paste0("gene", 1:n_genes)
       )

plotMA.label_ly(res, lab.genes = c("gene1", "gene2"))
```

---

plotPCA.ly *Plot an interactive PCA plot*

Description

Plot an interactive PCA plot

Usage

```
plotPCA.ly(rld, intgroup)
```

Arguments

rld	DESeqTransform object output by varianceStabilizingTransformation() or rlog()
intgroup	character vector of names in colData(x) to use for grouping

Value

Handle to ggplot with added label field in aes_string() for plotting with ggplotly()

Examples

```
# make example dds object
dds <- DESeq2::makeExampleDESeqDataSet()

# normalize
rld <- DESeq2::varianceStabilizingTransformation(dds, blind=TRUE)

# make pca plot
p <- plotPCA.ly(rld, intgroup='condition')
```

`plotPCA.san` *Adjustable PCA plot*

Description

Create a PCA plot with specified PCs on x- and y-axis

Usage

```
plotPCA.san(
  object,
  intgroup = "group",
  pcx,
  pcy,
  pcz = NULL,
  ntop = 500,
  samples = NULL,
  loadings = FALSE,
  loadings_ngenes = 10
)
```

Arguments

`object`	normalized DESeqDataSet object
`intgroup`	metadata variable to use for grouping samples
`pcx`	principal component to plot on x-axis
`pcy`	principal component to plot on y-axis
`pcz`	principal component to plot on z-axis. If not NULL, function returns a 3-D PCA plot.
`ntop`	number of most-variable genes to use
`samples`	vector of sample names to show on plot
`loadings`	boolean, show gene loadings? Default is FALSE.
`loadings_ngenes`	integer, # genes to show loadings for (default=10)

Value

ggplot handle

Examples

```
# make example dds object
dds <- DESeq2::makeExampleDESeqDataSet()

# normalize
rld <- DESeq2::varianceStabilizingTransformation(dds, blind=TRUE)

# make pca plot
p <- plotPCA.san(rld, intgroup='condition', pcx='PC1', pcy='PC2')
```

`plotScatter.label` *Plot a scatterplot to compare two contrasts*

Description

Plot a scatterplot to compare two contrasts

Usage

```
plotScatter.label(
  compare,
  df,
  label_x,
  label_y,
  lim.x,
  lim.y,
  color.palette,
  lab.genes = NULL,
  plot_all = "no",
  name.col = "geneid",
  lines = c("yes", "yes", "yes"),
  alpha = 1,
  size = 4,
  show.grid = "yes"
)
```

Arguments

`compare`	string, what values to plot? can be 'log2FoldChange' or 'P-adj'
`df`	data frame with log2FoldChange & padj values to plot from 2 contrasts
`label_x`	string, label for x-axis
`label_y`	string, label for y-axis
`lim.x`	x-axis limits
`lim.y`	y-axis limits
`color.palette`	character vector of colors to use for significance categories 'Both - same LFC sign', 'Both - opposite LFC sign', 'None', label_x, label_y
`lab.genes`	genes to label (default=NULL)
`plot_all`	string, can be 'yes' or 'no'. if 'yes', points outside axis limits are plotted along x/y axis lines (default='no').
`name.col`	gene name column to merge the 2 results, also used for labeling points
`lines`	3-element character vector to plot gridlines in the order (x=0, y=0, x=y), with 'yes' or 'no' values. E.g. ('yes', 'yes', 'no') will plot dotted lines for x = 0 & y = 0, but not the x = y diagonal.
`alpha`	float, marker opacity (default=1).
`size`	float, marker size (default=4).
`show.grid`	string, can be 'yes' (default) or 'no'.

Value

ggplot handle

Examples

```
# make mock results df
n_genes <- 100
res1 <- data.frame(
          baseMean = runif(n_genes, 10, 1000),
          log2FoldChange = rnorm(n_genes, 0, 2),
          lfcSE = runif(n_genes, 0.1, 0.5),
          stat = rnorm(n_genes, 0, 3),
          pvalue = runif(n_genes, 0, 1),
          padj = runif(n_genes, 0, 1),
          symbol = paste0("GENE", 1:n_genes),
          row.names = paste0("gene", 1:n_genes)
        )

res2 <- data.frame(
          baseMean = runif(n_genes, 10, 1000),
          log2FoldChange = rnorm(n_genes, 0, 2),
          lfcSE = runif(n_genes, 0.1, 0.5),
          stat = rnorm(n_genes, 0, 3),
          pvalue = runif(n_genes, 0, 1),
          padj = runif(n_genes, 0, 1),
          symbol = paste0("GENE", 1:n_genes),
          row.names = paste0("gene", 1:n_genes)
        )

# add geneid column
res1 <- cbind(geneid=rownames(res1), res1)
res2 <- cbind(geneid=rownames(res2), res2)

# make merged df from the two comparisons
cols.sub <- c('log2FoldChange', 'padj', 'geneid')
df_full <- dplyr::inner_join(
  dplyr::select(as.data.frame(res1), all_of(cols.sub)),
  dplyr::select(as.data.frame(res2), all_of(cols.sub)),
  by = 'geneid',
  suffix = c('.x', '.y')
)

# calculate x & y limits for log2FoldChange
xlim <- range(df_full[[ 'log2FoldChange.x' ]])
ylim <- range(df_full[[ 'log2FoldChange.y' ]])

# get color palette
color.palette <- RColorBrewer::brewer.pal(n=5, name='Set2')

# add significance column
sig.x <- df_full$padj.x < 0.1 & !is.na(df_full$padj.x)
sig.y <- df_full$padj.y < 0.1 & !is.na(df_full$padj.y)
up.x <- df_full$log2FoldChange.x >= 0
up.y <- df_full$log2FoldChange.y >= 0
significance <- rep('None', nrow(df_full))
significance[ sig.x & sig.y & ((up.x & up.y) | (!up.x & !up.y)) ] <- 'Both - same LFC sign'
```

```
significance[ sig.x & sig.y & ((up.x & !up.y) | (!up.x & up.y)) ] <- 'Both - opposite LFC sign'
significance[ sig.x & !sig.y ] <- 'A vs B'
significance[ !sig.x & sig.y ] <- 'B vs A'
df_full$significance <- significance

# generate scatter plot
p <- plotScatter.label(compare = 'log2FoldChange',
                       df = df_full,
                       label_x = 'A vs B',
                       label_y = 'B vs A',
                       lim.x = xlim,
                       lim.y = ylim,
                       color.palette = color.palette)
```

`plotScatter.label_ly` *Plot an interactive scatterplot to compare two contrasts*

Description

Plot an interactive scatterplot to compare two contrasts

Usage

```
plotScatter.label_ly(
  compare,
  df,
  label_x,
  label_y,
  lim.x,
  lim.y,
  color.palette,
  lab.genes = NULL,
  name.col = "geneid",
  lines = c("yes", "yes", "yes"),
  alpha = 1,
  size = 4,
  show.grid = "yes",
  source = "A"
)
```

Arguments

`compare`	string, what values to plot? can be 'log2FoldChange' or 'P-adj'
`df`	data frame with log2FoldChange & padj values to plot from 2 contrasts
`label_x`	string, label for x-axis
`label_y`	string, label for y-axis
`lim.x`	x-axis limits
`lim.y`	y-axis limits
`color.palette`	character vector of colors to use for significance categories 'Both - same LFC sign', 'Both - opposite LFC sign', 'None', label_x, label_y

lab.genes	genes to label (default=NULL)
name.col	gene name column to merge the 2 results, also used for labeling points
lines	3-element character vector to plot gridlines in the order (x=0, y=0, x=y), with 'yes' or 'no' values. E.g. ('yes', 'yes', 'no') will plot dotted lines for x = 0 & y = 0, but not the x = y diagonal.
alpha	float, marker opacity (default=1).
size	float, marker size (default=4).
show.grid	string, can be 'yes' (default) or 'no'.
source	name of source to return event_data from

Value

plotly handle

Examples

```
# make mock results df
n_genes <- 100
res1 <- data.frame(
          baseMean = runif(n_genes, 10, 1000),
          log2FoldChange = rnorm(n_genes, 0, 2),
          lfcSE = runif(n_genes, 0.1, 0.5),
          stat = rnorm(n_genes, 0, 3),
          pvalue = runif(n_genes, 0, 1),
          padj = runif(n_genes, 0, 1),
          symbol = paste0("GENE", 1:n_genes),
          row.names = paste0("gene", 1:n_genes)
        )

res2 <- data.frame(
          baseMean = runif(n_genes, 10, 1000),
          log2FoldChange = rnorm(n_genes, 0, 2),
          lfcSE = runif(n_genes, 0.1, 0.5),
          stat = rnorm(n_genes, 0, 3),
          pvalue = runif(n_genes, 0, 1),
          padj = runif(n_genes, 0, 1),
          symbol = paste0("GENE", 1:n_genes),
          row.names = paste0("gene", 1:n_genes)
        )

# add geneid column
res1 <- cbind(geneid=rownames(res1), res1)
res2 <- cbind(geneid=rownames(res2), res2)

# make merged df from the two comparisons
cols.sub <- c('log2FoldChange', 'padj', 'geneid')
df_full <- dplyr::inner_join(
  dplyr::select(as.data.frame(res1), all_of(cols.sub)),
  dplyr::select(as.data.frame(res2), all_of(cols.sub)),
  by = 'geneid',
  suffix = c('.x', '.y')
)

# calculate x & y limits for log2FoldChange
```

```
xlim <- range(df_full[[ 'log2FoldChange.x' ]])
ylim <- range(df_full[[ 'log2FoldChange.y' ]])

# get color palette
color.palette <- RColorBrewer::brewer.pal(n=5, name='Set2')

# add significance column
sig.x <- df_full$padj.x < 0.1 & !is.na(df_full$padj.x)
sig.y <- df_full$padj.y < 0.1 & !is.na(df_full$padj.y)
up.x <- df_full$log2FoldChange.x >= 0
up.y <- df_full$log2FoldChange.y >= 0
significance <- rep('None', nrow(df_full))
significance[ sig.x & sig.y & ((up.x & up.y) | (!up.x & !up.y)) ] <- 'Both - same LFC sign'
significance[ sig.x & sig.y & ((up.x & !up.y) | (!up.x & up.y)) ] <- 'Both - opposite LFC sign'
significance[ sig.x & !sig.y ] <- 'A vs B'
significance[ !sig.x & sig.y ] <- 'B vs A'
df_full$significance <- significance

# generate scatter plot
p <- plotScatter.label_ly(compare = 'log2FoldChange',
                          df = df_full,
                          label_x = 'A vs B',
                          label_y = 'B vs A',
                          lim.x = xlim,
                          lim.y = ylim,
                          color.palette = color.palette)
```

`plotVolcano.label` *Create a static labeled Volcano plot*

Description

This function creates a static ggplot Volcano plot from a data.frame containing DE analysis results. Mirrors plotVolcano.label_ly but renders with ggplot2 for use in downloads.

Usage

```
plotVolcano.label(
  res,
  fdr.thres = 0.1,
  fc.thres = 0,
  y.axis.lim = NULL,
  fc.lim = c(-4, 4),
  lab.genes = NULL,
  tolower.cols = c("SYMBOL", "ALIAS"),
  color_by = c("baseMean", "significance"),
  colorscale = "viridis",
  alpha = 0.6,
  pvalue_padj_switch = c("pvalue", "padj")
)
```

Arguments

res	data.frame with DE analysis results. Must contain "pvalue", "padj", "baseMean" & "log2FoldChange" columns
fdr.thres	False discovery rate (FDR) threshold
fc.thres	log2FoldChange threshold
y.axis.lim	y-axis limits for negative log10 p-values
fc.lim	x-axis limits
lab.genes	genes to label on plot
tolower.cols	column names that will be converted to lower case
color_by	variable to color points by ('baseMean' or 'significance')
colorscale	ggplot-compatible continuous colorscale name (viridis option), only used when color_by='baseMean'
alpha	point opacity between 0 and 1
pvalue_padj_switch	Switches between having the y-axis display raw or adjusted p-values. Significance is always determined by adjusted values.

Value

ggplot handle

Examples

```
n_genes <- 100
res <- data.frame(
         baseMean = runif(n_genes, 10, 1000),
         log2FoldChange = rnorm(n_genes, 0, 2),
         lfcSE = runif(n_genes, 0.1, 0.5),
         stat = rnorm(n_genes, 0, 3),
         pvalue = runif(n_genes, 0, 1),
         padj = runif(n_genes, 0, 1),
         symbol = paste0("GENE", 1:n_genes),
         row.names = paste0("gene", 1:n_genes)
       )

plotVolcano.label(res, lab.genes = c("gene1", "gene2"))
```

---

plotVolcano.label_ly *Create an interactive labeled Volcano plot*

Description

This function creates an Volcano plot from a data.frame containing DE analysis results using plot_ly

Usage

```
plotVolcano.label_ly(
  res,
  fdr.thres = 0.1,
  fc.thres = 0,
  y.axis.lim = NULL,
  fc.lim = c(-4, 4),
  lab.genes = NULL,
  tolower.cols = c("SYMBOL", "ALIAS"),
  color_by = c("baseMean", "significance"),
  colorscale = NULL,
  alpha = 0.6,
  pvalue_padj_switch = c("pvalue", "padj")
)
```

Arguments

res	data.frame with DE analysis results. Must contain "pvalue", "padj", "log2FoldChange", & "baseMean" columns
fdr.thres	False discovery rate (FDR) threshold
fc.thres	log2FoldChange threshold
y.axis.lim	y-axis limits for negative log10 p-values
fc.lim	x-axis limits for log2FoldChange
lab.genes	genes to label on volcano plot
tolower.cols	column names that will be converted to lower case
color_by	variable to color points by ('baseMean' or 'significance')
colorscale	ggplot-compatible continuous colorscale name (viridis option), only used when color_by='baseMean'
alpha	point opacity between 0 and 1
pvalue_padj_switch	Switches between having the y-axis display raw or adjusted p-values. Significance is always determined by adjusted values.

Value

plotly handle

Examples

```
# make mock results df
n_genes <- 100
res <- data.frame(
         baseMean = runif(n_genes, 10, 1000),
         log2FoldChange = rnorm(n_genes, 0, 2),
         lfcSE = runif(n_genes, 0.1, 0.5),
         stat = rnorm(n_genes, 0, 3),
         pvalue = runif(n_genes, 0, 1),
         padj = runif(n_genes, 0, 1),
         symbol = paste0("GENE", 1:n_genes),
         row.names = paste0("gene", 1:n_genes)
       )
```

```
plotVolcano.label_ly(res, lab.genes = c("gene1", "gene2"))
```

---

radarmod *Radar plot module*

Description

UI & module to generate radar plots.

Usage

```
radarUI(id, panel, type = "")

radarServer(id, obj, config, type = "")
```

Arguments

id	Module id
panel	string, can be 'sidebar' or 'main'
type	string, if 'comp' then show the comparison view
obj	reactiveValues object containing GeneTonic object
config	reactive list with config settings

Value

UI returns tagList with plot UI server invisibly returns NULL (used for side effects)

Examples

```
library(shiny)

# get enrichResult object
data(eres_dex, package='carnation')

# convert to GeneTonic object
gt <- GeneTonic::shake_enrichResult(eres_dex)

obj <- reactive({
  list(l_gs = gt$l_gs,
       anno_df = gt$anno_df,
       label = 'comp1')
})

config <- reactiveVal(get_config())

# run simple shiny app with plot
if(interactive()){
  shinyApp(
    ui = fluidPage(
           sidebarPanel(radarUI('p', 'sidebar')),
```

```
            mainPanel(radarUI('p', 'main'))
          ),
    server = function(input, output, session){
               radarServer('p', obj, config)
             }
  )
}
```

---

read_access_yaml *Read access yaml with user groups and data areas*

Description

This function reads the access yaml file and returns user groups and data areas as a list of data frames.

Usage

```
read_access_yaml()
```

Value

return carnation access settings from yaml file

Examples

```
# save access details to file
home <- Sys.getenv('HOME')

# create carnation data area if it doesn't exist
carnation_home <- file.path(home, 'carnation/data')
if(!dir.exists(carnation_home)) dir.create(carnation_home)

create_access_yaml(user = 'admin',
                   user_group = 'admin',
                   data_area = carnation_home)

al <- read_access_yaml()
```

---

res_cell *A* DESeqResults *object testing the difference between two cell lines of smooth muscle cells*

Description

A DESeqResults object testing the difference between two cell lines of smooth muscle cells

Format

A DESeqResults object, generated in the DESeq2 framework

Details

This DESeqResults object on the data from the airway package has been created comparing two smooth muscle cell lines, accounting for the effect of dexamethasone treatment.

Details on how this object has been created are included in the create_carnation_data.R script, included in the scripts folder of the Carnation package.

References

Himes BE, Jiang X, Wagner P, Hu R, Wang Q, Klanderman B, Whitaker RM, Duan Q, Lasky-Su J, Nikolos C, Jester W, Johnson M, Panettieri R Jr, Tantisira KG, Weiss ST, Lu Q. "RNA-Seq Transcriptome Profiling Identifies CRISPLD2 as a Glucocorticoid Responsive Gene that Modulates Cytokine Function in Airway Smooth Muscle Cells." PLoS One. 2014 Jun 13;9(6):e99625. PMID: 24926665. GEO: GSE52778

---

res_dex *A* DESeqResults *object testing the effect of dexamethasone on smooth muscle cells*

Description

A DESeqResults object testing the effect of dexamethasone on smooth muscle cells

Format

A DESeqResults object, generated in the DESeq2 framework

Details

This DESeqResults object on the data from the airway package has been created comparing dexamethasone treated vs untreated samples, accounting for the different cell lines included.

Details on how this object has been created are included in the create_carnation_data.R script, included in the scripts folder of the Carnation package.

References

Himes BE, Jiang X, Wagner P, Hu R, Wang Q, Klanderman B, Whitaker RM, Duan Q, Lasky-Su J, Nikolos C, Jester W, Johnson M, Panettieri R Jr, Tantisira KG, Weiss ST, Lu Q. "RNA-Seq Transcriptome Profiling Identifies CRISPLD2 as a Glucocorticoid Responsive Gene that Modulates Cytokine Function in Airway Smooth Muscle Cells." PLoS One. 2014 Jun 13;9(6):e99625. PMID: 24926665. GEO: GSE52778

run_carnation *Carnation*

Description

Interactive shiny dashboard for exploring RNA-Seq analysis.

Usage

```
run_carnation(
  credentials = NULL,
  passphrase = NULL,
  enable_admin = TRUE,
  config_path = NULL,
  ...
)
```

Arguments

credentials	path to encrypted sqlite db with user credentials.
passphrase	passphrase for credentials db.
enable_admin	if TRUE, admin view is shown. Note, this is only available if credentials have sqlite backend.
config_path	optional path to a local config yaml override.
...	parameters passed to shinyApp() call

Value

shinyApp object

Examples

```
if(interactive()){
    shiny::runApp(
        run_carnation()
    )
}
```

savemod *Save object module UI*

Description

Module UI & server to save carnation object.

Usage

```
saveUI(id)

saveServer(id, original, current, coldata, pattern, username, config)
```

Arguments

id	Module id
original	original carnation object
current	current carnation object
coldata	reactiveValues object containing object metadata
pattern	regex pattern for finding carnation data
username	user name
config	reactive list with config settings

Value

UI returns actionButton Server returns reactive with trigger to refresh the app

Examples

```
library(shiny)
library(DESeq2)

# default username
username <- reactive({ NULL })

# internal carnation config
config <- reactiveVal(get_config())

# regex to find carnation files
pattern <- reactive({ config()$server$pattern })

# get example object
obj <- make_example_carnation_object()

# make reactive with obj & path
original <- reactiveValues( obj = obj, path = "/path/to/carnation/obj.rds" )

# extract metadata
coldata <- reactive({ lapply(obj$dds, colData) })

# edit metadata
coldata_edit <- lapply(coldata, function(x){
                  x$type <- 'new'; x
                })

# add to object
edit_obj <- obj
for(name in names(edit_obj$dds)){
  colData(edit_obj$dds[[ name ]]) <- coldata_edit[[ name ]]
}

# run simple shiny app with plot
shinyApp(
  ui = fluidPage(
         saveUI('p')
       ),
  server = function(input, output, session){
             save_event <- saveServer('save_object',
```

```
                                    original=original,
                                    current=reactive({ edit_obj }),
                                    coldata=coldata,
                                    pattern=pattern(),
                                    username=username,
                                    config)
            }
)
```

---

save_access_yaml *Save access yaml to file*

Description

This function saves access details (user groups and data areas) to the designated access yaml file.

Usage

```
save_access_yaml(lst)
```

Arguments

lst	list of data frames with user_groups and data_areas

Value

save access settings to yaml file

Examples

```
# save access details to file
home <- Sys.getenv('HOME')

# create carnation data area if it doesn't exist
carnation_home <- file.path(home, 'carnation/data')
if(!dir.exists(carnation_home)) dir.create(carnation_home)

create_access_yaml(user = 'admin',
                   user_group = 'admin',
                   data_area = carnation_home)

# read access yaml
lst <- read_access_yaml()

# add new user
lst$user_group$admin <- c(lst$user_group$admin, 'user1')

# save to access settings
save_access_yaml(lst)
```

scattermod *Scatterplot module*

Description

Module UI + server for generating scatter plots.

Usage

```
scatterPlotUI(id, panel)

scatterPlotServer(id, obj, plot_args, gene_scratchpad, reset_genes, config)
```

Arguments

id	Module id
panel	string, can be 'sidebar' or 'main' passed to UI
obj	reactiveValues object containing carnation object passed to server
plot_args	reactive containing 'fdr.thres' (padj threshold), 'fc.thres' (log2FC)
gene_scratchpad	reactive containing gene scratchpad genes
reset_genes	reactive to reset gene scratchpad selection
config	reactive list with config settings passed to server

Value

UI returns tagList with scatter plot UI. Server invisibly returns NULL (used for side effects).

Examples

```
library(shiny)

# Create reactive values to simulate app state
oobj <- make_example_carnation_object()

obj <- reactiveValues(
   dds = oobj$dds,
   rld = oobj$rld,
   res = oobj$res,
   all_dds = oobj$all_dds,
   all_rld = oobj$all_rld,
   dds_mapping = oobj$dds_mapping
)

plot_args <- reactive({
  list(
    fdr.thres=0.1,
    fc.thres=0
  )
})
```

```
gene_scratchpad <- reactive({ c('gene1', 'gene2') })
reset_genes <- reactiveVal()

config <- reactiveVal(get_config())

shinyApp(
  ui = fluidPage(
         sidebarPanel(scatterPlotUI('p', 'sidebar')),
         mainPanel(scatterPlotUI('p', 'sidebar'))
       ),
  server = function(input, output, session){
             scatter_data <- scatterPlotServer('p', obj, plot_args,
                                gene_scratchpad, reset_genes, config)
           }
)
```

`settingsmod` *Settings module*

Description

Module UI & server for user access details interface.

Server code for settings module

Usage

```
settingsUI(id, panel, username)

settingsServer(id, details, depth, end_offset, assay_fun, config)
```

Arguments

`id`	Module id
`panel`	context for generating ui elements ('sidebar' or 'main')
`username`	user name
`details`	reactive list with user name & app location details
`depth`	project name depth
`end_offset`	project name end offset
`assay_fun`	function to parse assay names from file path
`config`	reactive list with config settings

Value

UI returns tagList with module UI Server returns reactive with list containing user access details

Examples

```
library(shiny)

# default username
username <- reactive({ NULL })

# internal carnation config
config <- reactiveVal(get_config())

# regex to find carnation files
pattern <- reactive({ config()$server$pattern })

# access permissions
assay.list <- reactiveValues(l=read_access_yaml())

if(interactive()){
  shinyApp(
    ui = fluidPage(
           sidebarPanel(uiOutput('settings_sidebar')),
           mainPanel(uiOutput('settings_main'))
         ),
    server = function(input, output, session){
               output$settings_main <- renderUI({
                 settingsUI('settings', panel='main', username=username)
               })

               output$settings_sidebar <- renderUI({
                 settingsUI('settings', panel='sidebar', username=username)
               })

               settings <- settingsServer('p',
                                          details=reactive({
                                                    list(username=username,
                                                         where=NULL)
                                                  }),
                                          depth=2,
                                          end_offset=0,
                                          assay_fun=function(x)
                                            sub(paste0(pattern(), '\\.rds$'), '',
                                                basename(x),
                                                ignore.case=TRUE),
                                          config
                                          )
             }
  )
}
```

`set_config` *Set config*

Description

This function updates a limited subset of the package config YAML. Only stable user-facing settings are writable; style settings and other internal options are intentionally left untouched.

Usage

```
set_config(
  config_path = get_config_path(),
  de_analysis = NULL,
  fdr_threshold = NULL,
  log2fc_threshold = NULL,
  max_upload_size = NULL,
  cores = NULL,
  pattern = NULL
)
```

Arguments

config_path	character path to the config YAML file to update. Defaults to the local config returned by `get_config_path()`. If the file does not exist yet, it is initialized from the bundled package config.
de_analysis	optional list with DE analysis config updates. Currently only `de_analysis$column_names` is supported, and the provided aliases are merged into the existing column-name mappings.
fdr_threshold	optional numeric FDR threshold between 0 and 1.
log2fc_threshold	optional numeric log2 fold-change threshold greater than or equal to 0.
max_upload_size	optional positive numeric upload limit in MB.
cores	optional positive integer number of cores to use.
pattern	optional character suffix pattern used to match dataset filenames before the trailing `.rds`. Use `""` to match all RDS files.

Value

Updated config list, invisibly.

Examples

```
cfg_out <- tempfile(fileext = ".yaml")

set_config(
  config_path = cfg_out,
  de_analysis = list(
    column_names = list(
      padj = "qvalue",
      log2FoldChange = c("logFC", "avg_log2FC")
    )
  ),
  fdr_threshold = 0.05,
  log2fc_threshold = 1,
  max_upload_size = 50,
```

```
  cores = 2,
  pattern = "carnation"
)
```

`summarize_res_list` *Combine everything in the results list into a single table*

Description

Combine everything in the results list into a single table

Usage

```
summarize_res_list(
  res.list,
  dds.list,
  dds_mapping,
  alpha,
  lfc.thresh,
  labels = NULL
)
```

Arguments

`res.list`	Named list of lists, where each sublist contains the following names: c('res', 'dds', 'label'). "res" is a DESeqResults object, "dds" is either the indexing label for the dds.list object or the DESeq object, and "label" is a nicer-looking label to use. NOTE: backwards compatibility with older versions of lcdb-wf depends on no dds.list object being passed.
`dds.list`	List of DESeqDataSet objects whose names are expected to match 'dds' slots in the 'res.list' object
`dds_mapping`	List mapping names of dds.list to res.list elements
`alpha`	false-discovery rate threshold
`lfc.thresh`	log2FoldChange threshold
`labels`	list of descriptions for res.list elements

Value

Dataframe

Examples

```
n_genes <- 100

#  make mock dds list
dds_list <- list(main=DESeq2::makeExampleDESeqDataSet(n=n_genes))

# make mock results df
res1 <- data.frame(
          baseMean = runif(n_genes, 10, 1000),
          log2FoldChange = rnorm(n_genes, 0, 2),
```

```
            lfcSE = runif(n_genes, 0.1, 0.5),
            stat = rnorm(n_genes, 0, 3),
            pvalue = runif(n_genes, 0, 1),
            padj = runif(n_genes, 0, 1),
            symbol = paste0("GENE", 1:n_genes),
            row.names = paste0("gene", 1:n_genes)
          )

res2 <- data.frame(
            baseMean = runif(n_genes, 10, 1000),
            log2FoldChange = rnorm(n_genes, 0, 2),
            lfcSE = runif(n_genes, 0.1, 0.5),
            stat = rnorm(n_genes, 0, 3),
            pvalue = runif(n_genes, 0, 1),
            padj = runif(n_genes, 0, 1),
            symbol = paste0("GENE", 1:n_genes),
            row.names = paste0("gene", 1:n_genes)
          )

# make list of results
res_list <- list(
               comp1=res1,
               comp2=res2
             )

# make dds mapping
dds_mapping <- list(comp1='main', comp2='main')

# get summary
df <- summarize_res_list(res_list, dds_list, dds_mapping, alpha=0.1, lfc.thresh=0)
```

---

sumovmod *Summary overview plot module*

Description

UI & module to generate summary overview plots.

Usage

```
sumovPlotUI(id, panel, type = "")

sumovPlotServer(id, obj, config, type = "")
```

Arguments

id	Module id
panel	string, can be 'sidebar' or 'main'
type	string, if 'comp' then show the comparison view
obj	reactiveValues object containing GeneTonic object
config	reactive list with config settings

Value

UI returns tagList with plot UI server invisibly returns NULL (used for side effects)

Examples

```
library(shiny)

# get enrichResult object
data(eres_dex, package='carnation')

# convert to GeneTonic object
gt <- GeneTonic::shake_enrichResult(eres_dex)

obj <- reactive({
  list(l_gs = gt$l_gs,
       anno_df = gt$anno_df,
       label = 'comp1')
})

config <- reactiveVal(get_config())

# run simple shiny app with plot
if(interactive()){
  shinyApp(
    ui = fluidPage(
           sidebarPanel(sumovPlotUI('p', 'sidebar')),
           mainPanel(sumovPlotUI('p', 'main'))
         ),
    server = function(input, output, session){
               sumovPlotServer('p', obj, config)
             }
  )
}
```

`top.genes` *Get top DE genes by log2FoldChange or adjusted p-value*

Description

Get top DE genes by log2FoldChange or adjusted p-value

Usage

```
top.genes(res, fdr.thres = 0.01, fc.thres = 0, n = 10, by = "log2FoldChange")
```

Arguments

`res`	data.frame with DE analysis results
`fdr.thres`	FDR threshold
`fc.thres`	log2FoldChange threshold
`n`	number of genes to return
`by`	metric to determine top genes ('log2FoldChange' or 'padj')

Value

vector of gene symbols

Examples

```
# get DE results
data(res_dex, package='carnation')

g <- top.genes(res_dex)
```

---

upsetmod *Upset plot module*

Description

Module UI & server to generate upset plots.

Usage

```
upsetPlotUI(id, panel)

upsetPlotServer(id, obj, plot_args, gene_scratchpad, reset_genes, config)
```

Arguments

id	Module id
panel	string, can be 'sidebar' or 'main'
obj	reactiveValues object containing carnation object
plot_args	reactive containing 'fdr.thres' (padj threshold) & 'fc.thres' (log2FC)
gene_scratchpad	reactiveValues object containing genes selected in scratchpad
reset_genes	reactive to reset gene scratchpad selection
config	reactive list with config settings

Value

UI returns tagList with upset plot UI. Server returns reactive with list containing upset table, intersections & selected genes.

Examples

```
library(shiny)

oobj <- make_example_carnation_object()

obj <- reactiveValues(
   dds = oobj$dds,
   rld = oobj$rld,
   res = oobj$res,
   all_dds = oobj$all_dds,
   all_rld = oobj$all_rld,
```

```
   dds_mapping = oobj$dds_mapping
)

plot_args <- reactive({
  list(
    fdr.thres=0.1,
    fc.thres=0
  )
})

gene_scratchpad <- reactive({ c('gene1', 'gene2') })

reset_genes <- reactiveVal()

config <- reactiveVal(get_config())

shinyApp(
  ui = fluidPage(
         sidebarPanel(upsetPlotUI('p', 'sidebar')),
         mainPanel(upsetPlotUI('p', 'sidebar'))
       ),
  server = function(input, output, session){
             upset_data <- upsetPlotServer('p', obj, plot_args,
                                           gene_scratchpad,
                                           reset_genes, config)
           }
)
```

`validate_carnation_object` *Validate a carnation object*

Description

This function takes various input data types (DE results, counts, enrichment, pattern analysis) and validates them according to carnation's requirements, returning a normalized intermediate object. Expensive derived-object creation steps such as variance-stabilized counts and GeneTonic conversion are handled separately by `materialize_carnation_object()`.

Usage

```
validate_carnation_object(
  res_list,
  dds_list,
  rld_list = NULL,
  labels = NULL,
  enrich_list = NULL,
  degpatterns = NULL,
  metadata = NULL,
  dds_mapping = NULL,
  config = NULL
)
```

Arguments

res_list	Named list of DE results. Each element should be either: • A data frame with DE results containing gene, symbol, pvalue, padj, log2FoldChange, and baseMean columns (or tool-specific alternatives) • A list with slots: res (data frame), dds (name reference to dds_list element), label (comparison label)
dds_list	Named list of count data. Each element should be either: • A DESeqDataSet object • A data frame or matrix of raw counts (first column=gene IDs, remaining=samples)
rld_list	Optional named list of variance-stabilized count objects. If NULL, these can be generated later via materialize_carnation_object().
labels	Optional named list of comparison labels. If NULL and res_list contains nested structure with label slots, labels will be extracted.
enrich_list	Optional named list of functional enrichment results. Should be structured as: enrich_list[[func_id]][[effect]][[pathway]]. Each enrichment result must be a data frame in clusterProfiler format: • Over-representation: ID, Description, GeneRatio, BgRatio, pvalue, p.adjust, qvalue, geneID, Count • GSEA: ID, Description, core_enrichment, setSize, pvalue, p.adjust, qvalue, NES
degpatterns	Optional named list of pattern analysis results. Each element should be either a data frame or a list with $normalized slot containing a data frame with columns: genes, value, and either cluster or columns starting with "cutoff".
metadata	Optional data frame with sample metadata. Required if dds_list contains count matrices instead of DESeqDataSet objects. First column should be sample names matching column names in count matrices.
dds_mapping	Optional named list mapping res_list elements to dds_list objects. Required if res_list is a list of data frames.
config	Optional config list. If NULL, will use get_config(), including any supported local config overrides.

Details

This function performs comprehensive validation of all input data:

- DE results: Checks for required columns (with support for DESeq2, edgeR, limma), ensures gene and symbol columns exist
- Counts: Validates structure, checks sample name matching with metadata
- Enrichment: Validates clusterProfiler format (OR or GSEA)
- Pattern analysis: Checks for required columns (genes, value, cluster)

If validation fails, the function will stop with an informative error message.

Value

A validated list with canonical slots res_list, dds_list, optional rld_list, labels, dds_mapping, enrich_list, degpatterns, and metadata when supplied.

A list containing normalized inputs with elements res_list, dds_list, optional rld_list, labels, dds_mapping, and optional enrich_list, degpatterns, and metadata.

Examples

```
# Minimal example with DE results and counts
library(DESeq2)

# Create example data
dds <- makeExampleDESeqDataSet()
dds <- DESeq(dds)
res <- results(dds, contrast = c("condition", "A", "B"))
rld <- varianceStabilizingTransformation(dds, blind = TRUE)

# Validate object inputs
obj <- validate_carnation_object(
  res_list = list(
    comp1 = list(
      res = as.data.frame(res),
      dds = "main",
      label = "A vs B"
    )
  ),
  dds_list = list(main = dds),
  rld_list = list(main = rld)
)

materialized <- materialize_carnation_object(obj, cores = 1)
final_obj <- make_final_object(materialized)

# Save for use with carnation
saveRDS(final_obj, "my_analysis.rds")

# Alternative: start from count matrix and metadata
counts <- as.data.frame(counts(dds))
counts$gene <- rownames(counts)
counts <- counts[, c(ncol(counts), 1:(ncol(counts)-1))]
metadata <- as.data.frame(colData(dds))
metadata$sample <- rownames(metadata)
metadata <- metadata[, c(ncol(metadata), 1:(ncol(metadata)-1))]

obj <- validate_carnation_object(
  res_list = list(comp1 = as.data.frame(res)),
  dds_list = list(main = counts),
  metadata = metadata,
  dds_mapping = list(comp1 = "main")
)
```

`volcanoplotmod` *Volcano plot module*

Description

UI & server for module to create volcano plot

Usage

```
volcanoPlotUI(id, panel)

volcanoPlotServer(id, obj, plot_args, config)
```

Arguments

id	Module id
panel	string, can be 'sidebar' or 'main'
obj	reactiveValues object containing carnation object
plot_args	reactive containing 'fdr.thres' (FDR threshold), 'fc.thres' (log2FC threshold) & 'gene.to.plot' (genes selected in scratchpad)
config	reactive list with config settings

Value

UI returns tagList with volcano plot UI. Server invisibly returns NULL (used for side effects).

Examples

```
library(shiny)
library(DESeq2)

# Create reactive values to simulate app state
oobj <- make_example_carnation_object()

obj <- reactiveValues(
   dds = oobj$dds,
   rld = oobj$rld,
   res = oobj$res,
   all_dds = oobj$all_dds,
   all_rld = oobj$all_rld,
   dds_mapping = oobj$dds_mapping
)

# Set up coldata structure that the module expects
coldata <- reactiveValues(
  curr = list(
    all_samples = colData(oobj$dds$main),
    main = colData(oobj$dds$main)
  )
)

plot_args <- reactive({
  list(
    fdr.thres=0.1,
    fc.thres=0,
    gene.to.plot=c('gene1', 'gene2')
  )
})

config <- reactiveVal(get_config())

shinyApp(
```

```
  ui = fluidPage(
         sidebarPanel(volcanoPlotUI('p', 'sidebar')),
         mainPanel(volcanoPlotUI('p', 'main'))
       ),
  server = function(input, output, session){
             volcanoPlotServer('p', obj, plot_args, config)
           }
)
```

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