Package 'altcdfenvs'

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Title alternative CDF environments (aka probeset mappings)

Author Laurent Gautier <lgautier@gmail.com>

Maintainer Laurent Gautier <lgautier@gmail.com>

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Description Convenience data structures and functions to handle cdfenvs

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Collate appendCdfEnvAffy.R buildCdfEnv.matchprobes.R buildCdfEnv.biostrings.R CdfEnv.R cdfenvs.R copyCdfEnvAffy.R readFASTA.R removeIndex.R unique.CdfEnvAffy.R

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Contents

AffyProbesMatch-class 2
appendCdfEnvAffy 3
buildCdfEnv.biostrings 4
CdfEnvAffy-class 4
cdfenvEx 6
cdfenvs 7
copyCdfEnvAffy 8

countduplicated 8
geneNames.CdfEnvAffy 9
getxy.probeseq 9
index2xy 10
indexProbes.CdfEnvAffy 11
matchAffyProbes 12
plot.CdfEnvAffy 13
read.FASTA.entry 14
removeIndex 15
toHypergraph 16
unique.CdfEnvAffy 17
validAffyBatch 18

Index **19**

`AffyProbesMatch-class` *Class "AffyProbesMatch"*

Description

Store the results of a call to `matchAffyProbes`.

Objects from the Class

Objects can be created by calls of the form `new("AffyProbesMatch", ...)`.

An object will store the result of matching probe sequences against target sequences.

Slots

`pm`: Object of class `"list"`: each element is vector of index values

`mm`: Object of class `"list"`: each element is vector of index values

`labels`: Object of class `"character"`

`chip_type`: Object of class `"character"` and of length 1.

`probes`: Object of class `"ANY"`: the `probetable` object used to perform the matches.

Methods

combine `signature(x = "AffyProbesMatch", y = "AffyProbesMatch")`: combine two instances. This is can be useful when splitting the list of target sequences to parallelized the job.

show `signature(x = "AffyProbesMatch")`:Show the instance.

toHypergraph `signature(object = "AffyProbesMatch")`: build an `Hypergraph` from the matches.

Examples

```
showClass("AffyProbesMatch")
```

appendCdfEnvAffy *append probe sets to a CdfEnvAffy*

Description

append probe sets to a CdfEnvAffy

Usage

```
appendCdfEnvAffy(acdfenv, id, i, nocopy = TRUE)
```

Arguments

acdfenv	instance of class CdfEnvAffy
id	identifier for the probe set to add
i	a matrix of indexes (see details)
nocopy	whether to make a copy of the environment or not (see details)

Details

The matrix i must have one column per probe type. For typical Affymetrix chip types, there are two probe types: "pm" and "mm".

nocopy set to TRUE means that the environment is added the probe set 'in-situ' (this can boost execution speed if you add a lot of probe sets).

Value

An CdfEnvAffy is returned

Examples

```
data(cdfenvEx)

## pm and mm probe set
m <- matrix(1:10, ncol = 2)
colnames(m) <- c("pm", "mm")

appendCdfEnvAffy(cdfenvEx, "blabla", m)

indexProbes(cdfenvEx, c("pm", "mm"), "blabla")

## pm only probe set
m <- matrix(6:9, ncol = 1)
colnames(m) <- c("pm")
appendCdfEnvAffy(cdfenvEx, "blabla2", m)
## note that the unspecified "mm" were set to NA
indexProbes(cdfenvEx, c("pm", "mm"), "blabla2")
```

buildCdfEnv.biostrings *Build CDF environments*

Description

Build CDF environment from Biostrings matchPDict results

Usage

```
buildCdfEnv.biostrings(apm, abatch = NULL,
                       nrow.chip = NULL, ncol.chip = NULL,
                       simplify = TRUE,
                       x.colname = "x", y.colname = "y",
                       verbose = FALSE)
```

Arguments

apm	AffyProbesMatch
abatch	AffyBatch
nrow.chip	number of rows for the chip type (see details)
ncol.chip	number of columns for the chip type (see details)
simplify	simplify the environment built (removing target names when there is no matching probe)
x.colname	column name
y.colname	column name
verbose	verbose TRUE/FALSE

Details

Whenever an abatch is specified, nrow.chip and ncol.chip are not needed. Specifying the an AffyBatch in abatch is the easiest way to specify information about the geometry of a chip type.

Value

An instance of class CdfEnvAffy.

CdfEnvAffy-class *Class "CdfEnvAffy"*

Description

A class to hold the information necessary to handle the grouping of probes in set of probes, and to find XY coordinates of probes on a chip

Objects from the Class

Objects can be created by calls of the form new("CdfEnvAffy", ...). Typically, there is an instance of the class for each type of chip (e.g. Hu6800, HG-U95A, etc...).

Slots

`envir`: Object of class `"environment"`. It has to be thought of as a hashtable: the keys are probe set identifiers, or gene names, and the values are indexes.

`envName`: Object of class `"character"`. A name for the environment.

`index2xy`: Object of class `"function"`. The function used to resolve index into xy coordinates. Unless you are an advanced user, you probably want to ignore this (and rely on the default provided with the package).

`xy2index`: Object of class `"function"`. The function used to resolve xy coordinates into index. Unless you are an advanced user, you probably want to ignore this (and rely on the default provided with the package).

`nrow`: Object of class `"integer"`. The number of rows of probes for the chip type.

`ncol`: Object of class `"integer"`. The number of columns of probes for the chip type.

`probeTypes`: Object of class `"character"`. The different types of probes stored for each probe set. In the case of Affymetrix chips, the probes are typically perfect match (pm) probes or mismatch probes (mm).

`chipType`: Object of class `"character"`. The name of the chip type the instance is associated with. This is useful when one starts to create alternative mappings of the probes on a chip (see associated vignette).

Methods

[`signature(object = "CdfEnvAffy",i = "character", j = "missing", drop = "boolean")`: subset a cdf, that is return a new cdf containing only a subset of the probe sets. The subset of probe sets to take is identified as a vector of identifiers (mode "character").

coerce `signature(object = "CdfEnvAffy", "environment")`: coerce an instance of the class to an `environment`.

coerce `signature(object = "CdfEnvAffy", "Cdf")`: coerce an instance of the class to a `Cdf`.

geneNames `signature(object="CdfEnvAffy")`: Return the names of the known probe sets (of course, it depends on the associated CDF).

index2xy `signature(object = "CdfEnvAffy", i="integer")`: convert index values into XY coordinates.

indexProbes `signature(object = "CdfEnvAffy", which = "character", probeSetNames = NULL)`: obtain the indexes for the probes associated wit the probe set name `probeSetNames`. When `probeSetNames` is set to `NULL` (default), the indexes are returned for the probe sets defined on the chip. See `indexProbes.CdfEnvAffy`

plot `signature(x = "CdfEnvAffy", y = "missing")`: Plot the chip. It mainly sets coordinates for further plotting (see examples). See `plot.CdfEnvAffy`

show `signature(object = "CdfEnvAffy")`: Print method.

xy2index `signature(object = "CdfEnvAffy", x="integer", y="integer")`: convert XY coordinates into index values.

toHypergraph `signature(object = "CdfEnvAffy")`: convert XY coordinates into index values.

Author(s)

Laurent Gautier

See Also

`indexProbes.CdfEnvAffy`, `plot.CdfEnvAffy`

Examples

```
## build an instance
library(hgu95acdf)
cdfenv.hgu95a <- wrapCdfEnvAffy(hgu95acdf, 640, 640, "HG-U95A")

show(cdfenv.hgu95a)

## find the indexes for a probe set (pm only)
ip <- indexProbes(cdfenv.hgu95a, "pm", "1000_at")[[1]]
## get the XY coordinates for the probe set
xy <- index2xy(cdfenv.hgu95a, ip)

## plot the chip
plot(cdfenv.hgu95a)

## plot the coordinates
plotLocation(xy)

## subset the environment

cdfenv.hgu95a.mini <- cdfenv.hgu95a["1000_at"]
```

---

cdfenvEx *CdfEnvAffy*

Description

An example of CdfEnvAffy

Usage

```
data(cdfenvEx)
```

Format

The format is: Formal class 'CdfEnvAffy' [package "altcdfenvs"] with 8 slots ..@ index2xy :function (object, i) ..@ xy2index :function (object, x, y) ..@ envir :length 2 <environment> ..@ envName : chr "ZG-DU33" ..@ nrow : int 100 ..@ ncol : int 100 ..@ probeTypes: chr [1:2] "pm" "mm" ..@ chipType : chr "ZG-DU33"

Examples

```
data(cdfenvEx)

print(cdfenvEx)
```

cdfenvs *functions related to cdfenvs*

Description

A set of functions to handle cdfenvs

Usage

```
wrapCdfEnvAffy(cdfenv, nrow.chip, ncol.chip, chiptype, check = TRUE,
               verbose = FALSE)
getCdfEnvAffy(abatch)
buildCdfEnv.matchprobes(matches, ids, probes.pack, abatch=NULL,
                        nrow.chip=NULL, ncol.chip=NULL, chiptype=NULL,
                        mm=NA, simplify = TRUE,
                        x.colname = "x", y.colname = "y", verbose=FALSE)
```

Arguments

abatch	an AffyBatch
cdfenv	A cdfenv environment
check	perform consistency check or not
chiptype	A name for the chip type
ids	a vector of probe set identifiers for the matches
matches	a list as returned by the function combineAffyBatch
mm	The value to store for MMs
ncol.chip	The number of columns for the chip type
nrow.chip	The number of rows for the chip type
probes.pack	The name of the probe package
simplify	Simplify the environment created by removing the ids without any matching probe
x.colname, y.colname	see the getxy.probeseq
verbose	verbosity (TRUE or FALSE)

Value

An instance of class CdfEnvAffy.

Examples

```
## See the main vignette
```

copyCdfEnvAffy *make a copy of a CdfEnvAffy*

Description

make a copy of a CdfEnvAffy

Usage

```
copyCdfEnvAffy(acdfenv)
```

Arguments

acdfenv	instance of class CdfEnvAffy

Details

Make a copy can be needed since a CdfEnvAffy contains an environment

Value

A CdfEnvAffy

See Also

CdfEnvAffy-class, copyEnv

countduplicated *Count the number of times probes are used*

Description

This function counts the number of times the probes in a CdfEnvAffy are found in this object.

Usage

```
countduplicated(x, incomparables = FALSE, verbose = FALSE)
```

Arguments

x	An instance of CdfEnvAffy-class
incomparables	(not implemented yet, keep away)
verbose	verbose or not

Value

An environment is returned. Each element in this environment has the same identifier than its corresponding probe set in the CdfEnvAffy-class and contains the number of times a probe is in use in the environment (instead of an index number in the CdfEnvAffy-class).

Author(s)

Laurent

See Also

CdfEnvAffy-class

---

geneNames.CdfEnvAffy *get the names of the known probe sets*

Description

get the names of the probe sets known to the CdfEnv

Usage

```
geneNames.CdfEnvAffy(object)
```

Arguments

object	CdfEnvAffy-class

Value

a vector of mode character

---

getxy.probeseq *A function to get the XY coordinates from a probes sequences data frame*

Description

A function to get the XY coordinates from a probes sequences data.frame

Usage

```
getxy.probeseq(ppset.id = NULL, probeseq = NULL, i.row = NULL,
xy.offset = NULL, x.colname = "x", y.colname = "y")
```

Arguments

ppset.id	The probe sets of interest (a vector of mode character.
probeseq	The probe sequence data.frame (see details).
i.row	Row indexes in the data.frame (see details).
xy.offset	Offset for the xy coordinates. if NULL, uses the default offset stored as an option for the affy package.
x.colname, y.colname	The probe sequence packages have seen the names for the columns in their data.frame. This parameters exists to let us follow these changes.

Details

The data.frame passed as argument probeseq is expected to have (at least) the following columns: Probe.X, Probe.Y and Probe.Set.Name. When the argument ppset.id is not null, the probe sets

Value

A matrix of two columns. The first column contains x coordinates, while the second column contains y coordinates.

Warning

The parameter xy.offset.one is here for historical reasons. This should not be touched, the option in the affy package should be modified if one wishes to modify this.

This function should not be confused with the methods index2xy and similar. Here the the XY coordinate come from a data.frame that stores information about an arbitrary number probes on the chip. (See the 'probe sequence' data packages on Bioconductor, and the package Biostrings).

The methods index2xy are meant to interact with instances of class AffyBatch.

Author(s)

Laurent

Examples

```
##---- Should be DIRECTLY executable !! ----
```

---

index2xy *Functions to shuttle from indexes to XY coordinates*

Description

Functions to shuttle from indexes to XY coordinates.

Usage

```
index2xy(object, ...)
xy2index(object, ...)
index2xy.CdfEnvAffy(object, i)
xy2index.CdfEnvAffy(object, x, y)
```

Arguments

object	An object of class CdfEnvAffy.
i	A vector of indexes.
x, y	Vectors of X and Y coordinates.
...	Optional parameters (not used).

Value

A vector of integers (for xy2index methods), or a matrix of two columns (for index2xy methods).

See Also

CdfEnvAffy-class

Examples

```
## To be done...
```

`indexProbes.CdfEnvAffy` *indexes for probes*

Description

A function to get the index for probes

Usage

```
indexProbes.CdfEnvAffy(object, which, probeSetNames = NULL)
```

Arguments

`object`	`CdfEnvAffy`
`which`	which kind of probe are of interest (see details).
`probeSetNames`	names of the probe sets of interest. If `NULL`, all the probe sets are considered.

Details

The parameter `which` let one specify which category of probes are of interest. In the case of Affymetrix chips, probes can be `"pm"` probes or `"mm"` probes. It the parameter is set to `c("pm", "mm")`, both are returned. Should other categories be defined, they can be handled as well.

Value

A `list` of indexes.

See Also

CdfEnvAffy-class, AffyBatch-class

matchAffyProbes *Match the probes on an Affymetrix array*

Description

Match the individual probes on an Affymetrix array to arbitrary targets.

Usage

```
mmProbes(probes)

matchAffyProbes(probes, targets, chip_type,
                matchmm = TRUE,
                selectMatches = function(x) which(elementNROWS(x) > 0),
                ...)
```

Arguments

probes	a probetable object
targets	a vector of references
chip_type	a name for the chip type.
matchmm	whether to match MM probes or not
selectMatches	a function to select matches (see Details).
...	further arguments to be passed to matchPDict.

Details

The matching is performed by the function matchPDict. The man page for that function will indicate what are the options it accepts.

In the case where a large number targets are given, like when each target represents a possible mRNA, is it expected to have a largely sparse incidence matrix, that is a low number of probes matching every target. For that reason, only the index of matching probes are associated with each given target, with the function selectMatches giving the definition of what are matching probes. The default function just count anything matching, but the user can specify a more stringent definition if wanted.

Value

mmProbes returns a vector of MM probe sequences.

matchAffyProbes returns an instance of AffyProbesMatch-class.

Author(s)

Laurent Gautier

See Also

matchPDict for details on how the matching is performed, AffyProbesMatch-class and buildCdfEnv.biostrings

Examples

```
library(hgu133aprobe)

filename <- system.file("exampleData", "sample.fasta",
                        package="altcdfenvs")

fasta.seq <- readDNAStringSet(filename)

targets <- as.character(fasta.seq)
names(targets) <- sub("^>.+\\|(NM[^ \\|]+|Hs[^ \\|]+)\\| ? .+$", "",
                      names(targets))

m <- matchAffyProbes(hgu133aprobe,
                     targets,
                     "HG-U133A")
```

`plot.CdfEnvAffy` *A function to 'plot' a CdfEnvAffy*

Description

A function to set the axis and plot the outline for a CdfEnvAffy

Usage

```
## S3 method for class 'CdfEnvAffy'
plot(x, xlab = "", ylab = "", main = x@chipType, ...)
```

Arguments

`x`	a `CdfEnvAffy`
`xlab`	label for the rows
`ylab`	label for the columns
`main`	label for the plot. The chip-type by default.
`...`	optional parameters to be passed to the underlying function `plot`

Details

This function does not 'plot' much, but sets the coordinates for further plotting (see the examples).

Author(s)

Laurent

See Also

`CdfEnvAffy-class`

Examples

```
## See "CdfEnvAffy-class"
```

`read.FASTA.entry` *Functions to work with FASTA files / connections*

Description

Set of function to work with biological sequences stored in FASTA format.

Usage

```
countskip.FASTA.entries(con, linebreaks = 3000)
grep.FASTA.entry(pattern, con, ...)
## S3 method for class 'FASTA'
print(x, ...)
read.FASTA.entry(con, linebreaks = 3000)
read.n.FASTA.entries(con, n, linebreaks = 3000)
read.n.FASTA.entries.split(con, n, linebreaks = 3000)
read.n.FASTA.headers(con, n, linebreaks = 3000)
read.n.FASTA.sequences(con, n, linebreaks = 3000)
skip.FASTA.entry(con, skip, linebreaks = 3000)
write.FASTA(x, file="data.fasta", append = FALSE)
```

Arguments

`append`	append to the file (or not)
`con`	a connection
`file`	a file name
`linebreaks`	(to optimize the parsing, probably safe to leave it as it is)
`n`	number of entries to read
`pattern`	a pattern (to be passed to the function `grep`)
`skip`	number of entries to skip
`x`	a FASTA sequence object
`...`	optional arguments to be forwarded to the function `print` or to the function `grep`

Details

`countskip.FASTA.entries` skips the remaining FASTA entries currently remaining in the connection and return the count. `grep.FASTA.entry` returns the next FASTA entry in the connection that matches a given regular expression. `print.FASTA` prints a FASTA object. `read.FASTA.entry` reads the next FASTA entry in the connection. `read.n.FASTA.entries` reads the `n` next FASTA entries and returns a `list` of FASTA objects. `read.n.FASTA.entries.split` reads the `n` next FASTA entries and returns a list of two elements: headers and sequences. `read.n.FASTA.headers` reads the `n` next FASTA headers. `read.n.FASTA.sequences` reads the `n` next FASTA sequences. `skip.FASTA.entry` skips a given number of FASTA entries. `write.FASTA` write a FASTA object into a connection.

Value

The value returned depends on the function. See above.

Author(s)

Laurent Gautier

Examples

```
filename <- system.file("exampleData", "sample.fasta",
                        package="altcdfenvs")
con <- file(filename, open="r")

fasta.seq <- grep.FASTA.entry("NM_001544\\.2", con)
close(con)


print(fasta.seq)
```

---

removeIndex *A function to remove probes in an environment*

Description

A function to remove probes in an environment, given their index.

Usage

```
removeIndex(x, i, simplify = TRUE, verbose = FALSE)
```

Arguments

x	An instance of CdfEnvAffy-class
i	A vector of indexes (integers !).
simplify	Simply the resulting CdfEnvAffy (see details).
verbose	verbose output or not.

Details

The probes to be removed are set to NA in the CdfEnvAffy. When simplify is set to TRUE the probe sets are simplified whenever possible. For example, if both pm and mm for the same probe pair are set to NA, then the probe pair is removed from the probe set.

Value

An instance of CdfEnvAffy-class is returned.

Author(s)

Laurent Gautier

See Also

CdfEnvAffy-class

Examples

```
## use plasmodiumanopheles chip as an example
if (require(plasmodiumanophelescdf)) {

  ## wrap in a (convenient) CdfEnvAffy object
  planocdf <- wrapCdfEnvAffy(plasmodiumanophelescdf, 712, 712, "plasmodiumanophelescdf")
  print(planocdf)

  ## ask for the probe indexed '10759' to be removed
  ## (note: if one wishes to remove from X/Y coordinates,
  ## the function xy2index can be of help).
  planocdfCustom <- removeIndex(planocdf, as.integer(10759))

  ## let see what happened (we made this example knowing in which
  ## probe set the probe indexed '10759' is found).
  indexProbes(planocdf, "pm", "200000_s_at")
  indexProbes(planocdfCustom, "pm", "200000_s_at")
  ## The 'second' pm probe (indexed '10579') in the probe set is now set
  ## to NA.
}
```

`toHypergraph` *Transform to an hypergraph*

Description

Transform to an hypergraph

Usage

```
toHypergraph(object, ...)
```

Arguments

`object`	Object derived from class `AffyProbesMatch`.
`...`	Unused.

Value

An `Hypergraph-class` object.

`unique.CdfEnvAffy` *Remove duplicated elements from a CdfEnvAffy*

Description

Remove duplicated elements from a CdfEnvAffy

Usage

```
## S3 method for class 'CdfEnvAffy'
unique(x, incomparables = FALSE, simplify = TRUE, verbose = FALSE, ...)
```

Arguments

`x`	An instance of `CdfEnvAffy-class`
`incomparables`	(not yet implemented)
`simplify`	simplify the result
`verbose`	verbose or not
`...`	(here for compatibility with the generic `unique`)

Details

The parameter `simplify` has the same function as the one with the same name in `countduplicated`.

Value

An instance of `CdfEnvAffy-class` in which probes used several times are removed.

Warning

The function differs slightly from the generic `unique`. Here the elements found in several place a merely removed.

Author(s)

Laurent

See Also

`countduplicated`

Examples

```
##not yet here...
```

validAffyBatch *Check validity of a CdfEnvAffy.*

Description

Tries to see if a CdfEnvAffy, or a pair of AffyBatch / CdfEnvAffy is valid.

Usage

```
validAffyBatch(abatch, cdfenv)
validCdfEnvAffy(cdfenv, verbose=TRUE)
printValidCdfEnvAffy(x)
```

Arguments

abatch	instance of AffyBatch-class
cdfenv	instance of CdfEnvAffy-class
verbose	verbose or not
x	object returned by validCdfEnvAffy

Details

The function validAffyBatch calls in turn validCdfEnvAffy.

See Also

AffyBatch-class, CdfEnvAffy-class

Examples

```
## To be done...
```

Index

∗ **IO**
read.FASTA.entry, 14
∗ **classes**
AffyProbesMatch-class, 2
CdfEnvAffy-class, 4
∗ **connection**
read.FASTA.entry, 14
∗ **datasets**
cdfenvEx, 6
∗ **hplot**
plot.CdfEnvAffy, 13
∗ **manip**
appendCdfEnvAffy, 3
buildCdfEnv.biostrings, 4
cdfenvs, 7
copyCdfEnvAffy, 8
countduplicated, 8
geneNames.CdfEnvAffy, 9
getxy.probeseq, 9
index2xy, 10
indexProbes.CdfEnvAffy, 11
matchAffyProbes, 12
read.FASTA.entry, 14
removeIndex, 15
toHypergraph, 16
unique.CdfEnvAffy, 17
validAffyBatch, 18
[,CdfEnvAffy,character,missing,missing-method *(CdfEnvAffy-class)*, 4

AffyProbesMatch-class, 2
appendCdfEnvAffy, 3

buildCdfEnv.biostrings, 4, *12*
buildCdfEnv.matchprobes *(cdfenvs)*, 7

CdfEnv *(cdfenvs)*, 7
CdfEnvAffy-class, 4
cdfenvEx, 6
cdfenvs, 7
coerce,CdfEnvAffy,Cdf-method *(CdfEnvAffy-class)*, 4
coerce,CdfEnvAffy,environment-method *(CdfEnvAffy-class)*, 4
combine,AffyProbesMatch,AffyProbesMatch-method *(AffyProbesMatch-class)*, 2
connection, *14*
copyCdfEnvAffy, 8
copyEnv, *8*
countduplicated, 8, *17*
countskip.FASTA.entries *(read.FASTA.entry)*, 14

geneNames,CdfEnvAffy-method *(CdfEnvAffy-class)*, 4
geneNames.CdfEnvAffy, 9
getCdfEnvAffy *(cdfenvs)*, 7
getxy.probeseq, 9
grep.FASTA.entry *(read.FASTA.entry)*, 14

Hypergraph, *2*

index2xy, 10
index2xy,CdfEnvAffy-method *(CdfEnvAffy-class)*, 4
indexProbes,CdfEnvAffy,character-method *(CdfEnvAffy-class)*, 4
indexProbes.CdfEnvAffy, *5*, 11

matchAffyProbes, 12
matchPDict, *12*
mmProbes *(matchAffyProbes)*, 12

plot,CdfEnvAffy,missing-method *(CdfEnvAffy-class)*, 4
plot.CdfEnvAffy, *5*, 13
print.FASTA *(read.FASTA.entry)*, 14
printValidCdfEnvAffy *(validAffyBatch)*, 18

read.FASTA.entry, 14
read.n.FASTA.entries *(read.FASTA.entry)*, 14
read.n.FASTA.headers *(read.FASTA.entry)*, 14
read.n.FASTA.sequences *(read.FASTA.entry)*, 14
removeIndex, 15

show,AffyProbesMatch-method
(AffyProbesMatch-class), 2
show,CdfEnvAffy-method
(CdfEnvAffy-class), 4
skip.FASTA.entry (read.FASTA.entry), 14

toHypergraph, 16
toHypergraph,AffyProbesMatch-method
(AffyProbesMatch-class), 2
toHypergraph,CdfEnvAffy-method
(CdfEnvAffy-class), 4

unique.CdfEnvAffy, 17

validAffyBatch, 18
validCdfEnvAffy (validAffyBatch), 18

wrapCdfEnvAffy (cdfenvs), 7
write.FASTA (read.FASTA.entry), 14

xy2index (index2xy), 10
xy2index,CdfEnvAffy-method
(CdfEnvAffy-class), 4