

# Package ‘MLInterfaces’

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**Title** Uniform interfaces to R machine learning procedures for data in Bioconductor containers

**Version** 1.87.0

**Description** This package provides uniform interfaces to machine learning code for data in R and Bioconductor containers.

**License** LGPL

**Depends** R (>= 3.5), Rcpp, methods, BiocGenerics (>= 0.13.11), Biobase, annotate, cluster

**Imports** gdata, pls, sfsmisc, MASS, rpart, genefilter, fpc, ggvis, shiny, gbm, RColorBrewer, hwriter, threejs (>= 0.2.2), mlbench, stats4, tools, grDevices, graphics, stats, magrittr, SummarizedExperiment

**Suggests** class, e1071, ipred, randomForest, gpls, pamr, nnet, ALL, hgu95av2.db, som, hu6800.db, lattice, caret (>= 5.07), golubEsets, ada, keggorthology, kernlab, mboost, party, klaR, BiocStyle, knitr, testthat

**Enhances** parallel

**LazyLoad** yes

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---

balKfold.xvspec	<i>generate a partition function for cross-validation, where the partitions are approximately balanced with respect to the distribution of a response variable</i>
-----------------	--

---

### Description

generate a partition function for cross-validation, where the partitions are approximately balanced with respect to the distribution of a response variable

### Usage

balKfold.xvspec(K)

**Arguments**

K                    number of partitions to be computed

**Details**

This function returns a closure. The symbol K is bound in the environment of the returned function.

**Value**

A closure consisting of a function that can be used as a `partitionFunc` for passage in `xvalSpec`.

**Author(s)**

VJ Carey <stvjc@channing.harvard.edu>

**Examples**

```
## The function is currently defined as
function(K)
function(data, clab, iternum) {
  clabs <- data[[clab]]
  narr <- nrow(data)
  cnames <- unique(clabs)
  ilyst <- list()
  for (i in 1:length(cnames)) ilyst[[cnames[i]]] <- which(clabs ==
    cnames[i])
  clen <- lapply(ilyst, length)
  nrep <- lapply(clen, function(x) ceiling(x/K))
  grpinds <- list()
  for (i in 1:length(nrep)) grpinds[[i]] <- rep(1:K, nrep[[i]])[1:clen[[i]]]
  (1:narr)[-which(unlist(grpinds) == iternum)]
}
# try it out
library("MASS")
data(crabs)
p1c = balKfold.xvspec(5)
inds = p1c( crabs, "sp", 3 )
table(crabs$sp[inds] )
inds2 = p1c( crabs, "sp", 4 )
table(crabs$sp[inds2] )
allc = 1:200
# are test sets disjoint?
intersect(setdiff(allc,inds), setdiff(allc,inds2))
```

---

brennan\_2013\_tabS7exc *Clinical characterization of 158 GBM samples from*  
*<https://doi.org/10.1016/j.cell.2013.09.034> supp table S7*

---

### **Description**

The clinical characteristics table of <https://doi.org/10.1016/j.cell.2013.09.034> in supplemental table S7 was aligned with the GBM samples in curatedTCGAData (selecting GBM and version 2.0.1 with curatedTCGAData 1.17.0).

### **Usage**

```
data("brennan_2013_tabS7exc")
```

### **Format**

A data frame with 158 observations on the following 16 variables.

Case\_ID a character vector

Secondary\_or\_Recurrent a character vector

Age\_at\_Procedure a numeric vector

Gender a character vector

Path\_Dx a character vector

MGMT\_Status a character vector

Methylation\_Class\_2012 a character vector

G\_CIMP\_methylation a character vector

IDH1\_status a character vector

Expression\_Subclass a character vector

Therapy\_Class a character vector

Vital\_Status a character vector

OS\_days a numeric vector

Progression\_Status a character vector

PFS\_days a numeric vector

V16 a logical vector

### **Details**

Simple intersection on Case\_ID in Supp Tab 7 with patientID in the GBM from curatedTCGAData.

### **Source**

<https://doi.org/10.1016/j.cell.2013.09.034>

**References**

The Somatic Genomic Landscape of Glioblastoma by Cameron W. Brennan, Roel G.W. Verhaak, Aaron McKenna, and others, Cell Oct 10 2013.

**Examples**

```
data(brennan_2013_tabS7exc)
head(brennan_2013_tabS7exc)
```

---

```
classifierOutput-class
      Class "classifierOutput"
```

---

**Description**

This class summarizes the output values from different classifiers.

**Objects from the Class**

Objects are typically created during the application of a supervised machine learning algorithm to data and are the value returned. It is very unlikely that any user would create such an object by hand.

**Slots**

**testOutcomes:** Object of class "factor" that lists the actual outcomes in the records on the test set

**testPredictions:** Object of class "factor" that lists the predictions of outcomes in the test set

**testScores:** Object of class "ANY" – this element will include matrices or vectors or arrays that include information that is typically related to the posterior probability of occupancy of the predicted class or of all classes. The actual contents of this slot can be determined by inspecting the converter element of the learnerSchema used to select the model.

**trainOutcomes:** Object of class "factor" that lists the actual outcomes in records on the training set

**trainPredictions:** Object of class "factor" that lists the predicted outcomes in the training set

**trainScores:** Object of class "ANY" see the description of testScores above; the same information is returned, but applicable to the training set records.

**trainInd:** Object of class "numeric" with of indices of data to be used for training.

**RObject:** Object of class "ANY" – when the trainInd parameter of the MLearn call is numeric, this slot holds the return value of the underlying R function that carried out the predictive modeling. For example, if rpartI was used as MLearn method, RObject holds an instance of the rpart S3 class, and plot and text methods can be applied to this. When the trainInd parameter of the MLearn call is an instance of xvalSpec, this slot holds a list of results of cross-validatory iterations. Each element of this list has two elements: test.idx, giving the numeric indices of the test cases for the associated cross-validation iteration, and mlans,

which is the `classifierOutput` for the associated iteration. See the example for an illustration of 'digging out' the predicted probabilities associated with each cross-validation iteration executed through an `xvalSpec` specification.

`embeddedCV`: logical value that is TRUE if the procedure in use performs its own cross-validation

`fsHistory`: list of features selected through cross-validation process

`learnerSchema`: propagation of the learner schema object used in the call

`call`: Object of class "call" – records the call used to generate the `classifierOutput` RObject

## Methods

**confuMat** signature(`obj = "classifierOutput"`): Compute the confusion matrix for test records.

**confuMatTrain** signature(`obj = "classifierOutput"`): Compute the confusion matrix for training set. Typically yields optimistically biased information on misclassification rate.

**RObject** signature(`obj = "classifierOutput"`): The R object returned by the underlying classifier. This can then be passed on to specific methods for those objects, when they exist.

**trainInd** signature(`obj = "classifierOutput"`): Returns the indices of data used for training.

**show** signature(`object = "classifierOutput"`): A print method that provides a summary of the output of the classifier.

**predictions** signature(`object = "classifierOutput"`): Print the predicted classes for each sample/individual. The predictions for the training set are the training outcomes.

**predictions** signature(`object = "classifierOutput"`, `t = "numeric"`): Print the predicted classes for each sample/individual that have a `testScore` greater or equal than `t`. The predictions for the training set are the training outcomes. Non-predicted cases and cases that match multiple classes are returned as NAs.

**predScore** signature(`object = "classifierOutput"`): Returns the scores for predicted class for each sample/individual. The scores for the training set are set to 1.

**predScores** signature(`object = "classifierOutput"`): Returns the prediction scores for all classes for each sample/individual. The scores for the training set are set to 1 for the appropriate class, 0 otherwise.

**testScores** signature(`object = "classifierOutput"`): ...

**testPredictions** signature(`object = "classifierOutput"`): Print the predicted classes for each sample/individual in the test set.

**testPredictions** signature(`object = "classifierOutput"`, `t = "numeric"`): Print the predicted classes for each sample/individual in the test set that have a `testScore` greater or equal than `t`. Non-predicted cases and cases that match multiple classes are returned as NAs.

**trainScores** signature(`object = "classifierOutput"`): ...

**trainPredictions** signature(`object = "classifierOutput"`): Print the predicted classes for each sample/individual in the train set.

**trainPredictions** signature(`object = "classifierOutput"`, `t = "numeric"`): Print the predicted classes for each sample/individual in the train set that have a `testScore` greater or equal than `t`. Non-predicted cases and cases that match multiple classes are returned as NAs.

**fsHistory** signature(`object = "classifierOutput"`): ...

**Author(s)**

V. Carey

**Examples**

```

showClass("classifierOutput")
library(golubEsets)
data(Golub_Train) # now cross-validate a neural net
set.seed(1234)
xv5 = xvalSpec("LOG", 5, balkfold.xvspec(5))
m2 = MLearn(ALL.AML~, Golub_Train[1000:1050,], nnetI, xv5,
  size=5, decay=.01, maxit=1900 )
testScores(RObject(m2)[[1]]$mlans)
alls = lapply(RObject(m2), function(x) testScores(x$mlans))

```

---

clusteringOutput-class

*container for clustering outputs in uniform structure*


---

**Description**

container for clustering outputs in uniform structure

**Objects from the Class**

Objects can be created by calls of the form `new("clusteringOutput", ...)`.

**Slots**

**partition:** Object of class "integer", labels for observations as clustered

**silhouette:** Object of class "silhouette", structure from Rousseeuw cluster package measuring cluster membership strength per observation

**prcomp:** Object of class "prcompObj" a wrapped instance of stats package prcomp output

**call:** Object of class "call" for auditing

**learnerSchema:** Object of class "learnerSchema", a formal object indicating the package, function, and other attributes of the clustering algorithm employed to generate this object

**RObject:** Object of class "ANY", the unaltered output of the function called according to learnerSchema

**converter:** converter propagated from call

**distFun:** distfun propagated from call

## Methods

**RObject** signature(x = "clusteringOutput"): extract the unaltered output of the R function or method called according to learnerSchema

**plot** signature(x = "clusteringOutput", y = "ANY"): a 4-panel plot showing features of the clustering, including the scree plot for a principal components transformation and a display of the partition in PC1xPC2 plane. For a clustering method that does not have a native plot procedure, such as kmeans, the parameter y should be bound to a data frame or matrix with feature data for all records; an image plot of robust feature z-scores ( $z=(x-\text{median}(x))/\text{mad}(x)$ ) and the cluster indices is produced in the northwest panel.

**show** signature(object = "clusteringOutput"): concise report

## Author(s)

VJ Carey <stvjc@channing.harvard.edu>

## Examples

```
showClass("clusteringOutput")
```

---

confuMat-methods

*Compute the confusion matrix for a classifier.*

---

## Description

This function will compute the confusion matrix for a classifier's output

## Methods

**obj = "classifOutput", ...** Typically, an instance of class "[classifierOutput](#)" is built on a training subset of the input data. The model is then used to predict the class of samples in the test set. When the true class labels for the test set are available the confusion matrix is the cross-tabulation of the true labels of the test set against the predictions from the classifier. An optional `t` score threshold can also be specified.

**obj = "classifierOutput", type="character", ...** For instances of [classifierOutput](#), it is possible to specify the type of confusion matrix desired. The default is `test`, which tabulates classes from the test set against the associated predictions. If `type` is `train`, the training class vector is tabulated against the predictions on the training set. An optional `t` score threshold can also be specified.

**obj = "classifierOutput", type="numeric"** For instances of [classifierOutput](#), it is possible to specify the minimum score feature classification threshold. Features with a score less than the threshold are classified as NA in the confusion `train` or `test` confusion matrix.



## Examples

```
library(golubEsets)
data(Golub_Merge)
smallG <- Golub_Merge[101:150,]
k1 <- MLearn(ALL.AML~., smallG, knnI(k=1), 1:30)
confuMat(k1)
confuMat(k1, "train")
```

---

confuTab

*Compute confusion tables for a confusion matrix.*

---

## Description

Given a  $n$  by  $n$  confusion matrix, the function returns a list of  $n$  2 by 2 tables with false positives, false negatives, false positives and true negative for each initial variables.

## Usage

```
confuTab(obj, naAs0. = FALSE)
```

## Arguments

`obj` An instance of class `table`. Must be square.  
`naAs0.` A logical, defining if NAs are to be replaced by 0s.

## Value

A list of length `nrow(obj)` and names `rownames(obj)`.

## Author(s)

Laurent Gatto <lg390@cam.ac.uk>

## See Also

The `tp`, `tn`, `fp`, `fn`, methods to extract the respective classification outcomes from a contingency matrix.

## Examples

```
## the confusion matrix
cm <- table(iris$Species, sample(iris$Species))
## the 3 confusion tables
(ct <- confuTab(cm))
```

---

`fs.absT`*support for feature selection in cross-validation*

---

**Description**

support for feature selection in cross-validation

**Usage**

```
fs.absT(N)
fs.probT(p)
fs.topVariance(p)
```

**Arguments**

N	number of features to retain; features are ordered by descending value of abs(two-sample t stat.), and the top N are used.
p	cumulative probability (in (0,1)) in the distribution of absolute t statistics above which we retain features

**Details**

This function returns a function that will be used as a parameter to [xvalSpec](#) in applications of [MLearn](#).

**Value**

a function is returned, that will itself return a formula consisting of the selected features for application of [MLearn](#).

**Note**

The functions `fs.absT` and `fs.probT` are two examples of approaches to embedded feature selection that make sense for two-sample prediction problems. For selection based on linear models or other discrimination measures, you will need to create your own selection helper, following the code in these functions as examples.

`fs.topVariance` performs non-specific feature selection based on the variance. Argument `p` is the variance percentile beneath which features are discarded.

**Author(s)**

VJ Carey <stvjc@channing.harvard.edu>

**See Also**

[MLearn](#)

**Examples**

```

library("MASS")
data(crabs)
# we will demonstrate this procedure with the crabs data.
# first, create the closure to pick 3 features
demFS = fs.absT(3)
# run it on the entire dataset with features excluding sex
demFS(sp~.-sex, crabs)
# emulate cross-validation by excluding last 50 records
demFS(sp~.-sex, crabs[1:150,])
# emulate cross-validation by excluding first 50 records -- different features retained
demFS(sp~.-sex, crabs[51:200,])

```

---

fsHistory	<i>extract history of feature selection for a cross-validated machine learner</i>
-----------	---

---

**Description**

extract history of feature selection for a cross-validated machine learner

**Usage**

```
fsHistory(x)
```

**Arguments**

x                   instance of `classifierOutput`

**Details**

returns a list of names of selected features

**Value**

a list; the names of variables are made 'syntactic'

**Author(s)**

Vince Carey <stvjc@channing.harvard.edu>

**Examples**

```

data(iris)
iris2 = iris[ iris$Species %in% levels(iris$Species)[1:2], ]
iris2$Species = factor(iris2$Species) # drop unused levels
x1 = MLearn(Species~., iris2, ldaI, xvalSpec("LOG", 3,
      balkfold.xvspec(3), fs.absT(3)))
fsHistory(x1)

```

---

hclustWidget	<i>shiny-oriented GUI for cluster or classifier exploration</i>
--------------	---

---

### Description

shiny-oriented GUI for cluster or classifier exploration

### Usage

```
hclustWidget(mat, featureName = "feature", title =  
             paste0("hclustWidget for ", deparse(substitute(mat))),  
             minfeats = 2, auxdf = NULL)
```

```
mlearnWidget(eset, infmla)
```

### Arguments

mat	matrix with feature vectors in rows
featureName	name to be used for control that asks for number of features to use
title	widget title
minfeats	lower bound on number of features to use
auxdf	data.frame with number of rows equal to nrow(mat), with metadata to be displayed in hovering tooltip
eset	instance of <a href="#">ExpressionSet-class</a>
infmla	instance of formula, with dependent variable values obtained in pData(eset), and independent variable specified as "." or as properly munged elements of featureNames(eset).

### Details

Experimental tool to illustrate impacts of choice of distance, agglomeration method, etc.

### Value

a shinyApp result that will display in active browser

### Note

mlearnWidget will attempt to nicely produce a variable importance plot using [randomForestI](#). This means that the annotation package for probe identifiers should be loaded or an error will be thrown.

### Author(s)

VJ Carey <stvjc@channing.harvard.edu>

**Examples**

```
# should run with example(hclustWidget, ask=FALSE)
if (interactive()) {
  library(shiny)
  library(MASS)
  data(crabs)
  cr = data.matrix(crabs[, -c(1:3)])
  au = crabs[, 1:3]
  show(hclustWidget(cr, auxdf=au))
## must use stop widget button to proceed
  library(ALL)
  library(hgu95av2.db)
  data(ALL)
  show(mlearnWidget(ALL[1:500,], mol.biol~.))
}
```

---

learnerSchema-class    *Class "learnerSchema" – convey information on a machine learning function to the MLearn wrapper*

---

**Description**

conveys information about machine learning functions in CRAN packages, for example, to MLearn wrapper

**Objects from the Class**

Objects can be created by calls of the form `new("learnerSchema", ...)`.

**Slots**

**packageName:** Object of class "character" string naming the package in which the function to be used is defined.

**mLFunName:** Object of class "character" string naming the function to be used

**converter:** Object of class "function" function with parameters `obj`, `data`, `trainInd`, that will produce a `classifierOutput` instance

**Methods**

**MLearn** signature(`formula = "formula"`, `data = "ExpressionSet"`, `method = "learnerSchema"`, `trainInd = "numeric"`): execute desired learner passing a formula and `ExpressionSet`

**MLearn** signature(`formula = "formula"`, `data = "data.frame"`, `method = "learnerSchema"`, `trainInd = "numeric"`): execute desired learner passing a formula

**show** signature(`object = "learnerSchema"`): concise display

**Author(s)**

Vince Carey <stvjc@channing.harvard.edu>

**Examples**

```
showClass("learnerSchema")
```

---

MLearn

*revised MLearn interface for machine learning*


---

**Description**

revised MLearn interface for machine learning, emphasizing a schematic description of external learning functions like knn, lda, nnet, etc.

**Usage**

```
MLearn( formula, data, .method, trainInd, ... )
makeLearnerSchema(packname, mlfunname, converter, predictor)
```

**Arguments**

formula	standard model formula
data	data.frame or ExpressionSet instance
.method	instance of learnerSchema
trainInd	obligatory numeric vector of indices of data to be used for training; all other data are used for testing, or instance of the xvalSpec class
...	additional named arguments passed to external learning function
packname	character – name of package harboring a learner function
mlfunname	character – name of function to use
converter	function – with parameters (obj, data, trainInd) that tells how to convert the material in obj [produced by [packname::mlfunname] ] into a classifierOutput instance.
predicter	function – with parameters (obj, newdata, ...) that tells how to use the material in obj to predict newdata.

**Details**

The purpose of the MLearn methods is to provide a uniform calling sequence to diverse machine learning algorithms. In R package, machine learning functions can have parameters (x, y, ...) or (formula, data, ...) or some other sequence, and these functions can return lists or vectors or other sorts of things. With MLearn, we always have calling sequence MLearn(formula, data, .method, trainInd, ...), and data can be a data.frame or ExpressionSet. MLearn will always return an S4 instance of classifierObject or clusteringObject.

At this time (1.13.x), NA values in predictors trigger an error.

To obtain documentation on the older (pre bioc 2.1) version of the MLearn method, please use help(MLearn-OLD).

**randomForestI** [randomForest](#). Note, that to obtain the default performance of `randomForestB`, you need to set `mtry` and `sampsiz` parameters to `sqrt(number of features)` and `table([training set response factor])` respectively, as these were not taken to be the function's defaults. Note you can use `xvalSpec("NOTEST")` as `trainInd`, to use all the samples; the `RObject()` result will print the misclassification matrix estimate along with OOB error rate estimate.

**knnI(k=1,l=0)** [knn](#); special support bridge required, defined in MLint

**knn.cvI(k=1,l=0)** [knn.cv](#); special support bridge required, defined in MLint. This option uses the embedded leave-one-out cross-validation of `knn.cv`, and thereby achieves high performance. You can have more general cross-validation using `knnI` with an `xvalSpec`, but it will be slower. When using this learner schema, you should use the numerical `trainInd` setting with `1:N` where `N` is the number of samples.

**dldaI** [diagDA](#); special support bridge required, defined in MLint

**nnetI** [nnet](#)

**rpartI** [rpart](#)

**ldaI** [lda](#)

**svmI** [svm](#)

**qdaI** [qda](#)

**logisticI(threshold)** [glm](#) – with binomial family, expecting a dichotomous factor as response variable, not bulletproofed against other responses yet. If response probability estimate exceeds threshold, predict 1, else 0

**adaI** [ada](#)

**BgbmI** [gbm](#), forcing the Bernoulli loss function.

**blackboostI** [blackboost](#) – you MUST supply a family parameter relevant for `mboost` package procedures

**lvqI** [lvqtest](#) after building codebook with `lvqinit` and updating with `olvq1`. You will need to write your own detailed schema if you want to tweak tuning parameters.

**naiveBayesI** [naiveBayes](#)

**baggingI** [bagging](#)

**sldaI** [slda](#)

**ksvmI** [ksvm](#)

**hclustI(distMethod, agglomMethod)** [hclust](#) – you must explicitly specify distance and agglomeration procedure.

**kmeansI(centers, algorithm)** [kmeans](#) – you must explicitly specify centers and algorithm name.

If the `parallel` package is attached, cross-validation will be distributed to cores using [mclapply](#).

## Value

Instances of `classifierOutput` or `clusteringOutput`

## Author(s)

Vince Carey <[stvj@channing.harvard.edu](mailto:stvj@channing.harvard.edu)>

**See Also**

Try `example(hclustWidget, ask=FALSE)` for an interactive approach to cluster analysis tuning.

**Examples**

```

library("MASS")
data(crabs)
set.seed(1234)
kp = sample(1:200, size=120)
rf1 = MLearn(sp~CW+RW, data=crabs, randomForestI, kp, ntree=600 )
rf1
nn1 = MLearn(sp~CW+RW, data=crabs, nnetI, kp, size=3, decay=.01,
  trace=FALSE )
nn1
RObject(nn1)
knn1 = MLearn(sp~CW+RW, data=crabs, knnI(k=3,l=2), kp)
knn1
names(RObject(knn1))
dlda1 = MLearn(sp~CW+RW, data=crabs, dldaI, kp )
dlda1
names(RObject(dlda1))
lda1 = MLearn(sp~CW+RW, data=crabs, ldaI, kp )
lda1
names(RObject(lda1))
slda1 = MLearn(sp~CW+RW, data=crabs, sldaI, kp )
slda1
names(RObject(slda1))
svm1 = MLearn(sp~CW+RW, data=crabs, svmI, kp )
svm1
names(RObject(svm1))
ldapp1 = MLearn(sp~CW+RW, data=crabs, ldaI.predParms(method="debiased"), kp )
ldapp1
names(RObject(ldapp1))
qda1 = MLearn(sp~CW+RW, data=crabs, qdaI, kp )
qda1
names(RObject(qda1))
logi = MLearn(sp~CW+RW, data=crabs, glmI.logistic(threshold=0.5), kp, family=binomial ) # need family
logi
names(RObject(logi))
rp2 = MLearn(sp~CW+RW, data=crabs, rpartI, kp)
rp2
## recode data for RAB
#nsp = ifelse(crabs$sp=="0", -1, 1)
#nsp = factor(nsp)
#ncrabs = cbind(nsp,crabs)
#rab1 = MLearn(nsp~CW+RW, data=ncrabs, RAB1, kp, maxiter=10)
#rab1
#
# new approach to adaboost
#
ada1 = MLearn(sp ~ CW+RW, data = crabs, .method = adaI,
  trainInd = kp, type = "discrete", iter = 200)

```



```

ada1
confuMat(ada1)
#
lvq.1 = MLearn(sp~CW+RW, data=crabs, lvqI, kp )
lvq.1
nb.1 = MLearn(sp~CW+RW, data=crabs, naiveBayesI, kp )
confuMat(nb.1)
bb.1 = MLearn(sp~CW+RW, data=crabs, baggingI, kp )
confuMat(bb.1)
#
# new mboost interface -- you MUST supply family for nonGaussian response
#
require(party) # trafo ... killing cmd check
blb.1 = MLearn(sp~CW+RW+FL, data=crabs, blackboostI, kp, family=mboost::Binomial() )
confuMat(blb.1)
#
# ExpressionSet illustration
#
data(sample.ExpressionSet)
# needed to increase training set size to avoid a new randomForest condition
# on empty class
set.seed(1234)
X = MLearn(type~., sample.ExpressionSet[100:250,], randomForestI, 1:19, importance=TRUE )
library(randomForest)
library(hgu95av2.db)
opar = par(no.readonly=TRUE)
par(las=2)
plot(getVarImp(X), n=10, plat="hgu95av2", toktype="SYMBOL")
par(opar)
#
# demonstrate cross validation
#
nn1cv = MLearn(sp~CW+RW, data=crabs[c(1:20,101:120),],
  nnetI, xvalSpec("L00"), size=3, decay=.01, trace=FALSE )
confuMat(nn1cv)
nn2cv = MLearn(sp~CW+RW, data=crabs[c(1:20,101:120),], nnetI,
  xvalSpec("LOG",5, balKfold.xvspec(5)), size=3, decay=.01,
  trace=FALSE )
confuMat(nn2cv)
nn3cv = MLearn(sp~CW+RW+CL+BD+FL, data=crabs[c(1:20,101:120),], nnetI,
  xvalSpec("LOG",5, balKfold.xvspec(5), fsFun=fs.absT(2)), size=3, decay=.01,
  trace=FALSE )
confuMat(nn3cv)
nn4cv = MLearn(sp~.-index-sex, data=crabs[c(1:20,101:120),], nnetI,
  xvalSpec("LOG",5, balKfold.xvspec(5), fsFun=fs.absT(2)), size=3, decay=.01,
  trace=FALSE )
confuMat(nn4cv)
#
# try with expression data
#
library(golubEsets)
data(Golub_Train)
litg = Golub_Train[ 100:150, ]

```

```

g1 = MLearn(ALL.AML~. , litg, nnetI,
  xvalSpec("LOG",5, balkfold.xvspec(5),
  fsFun=fs.probt(.75)), size=3, decay=.01, trace=FALSE )
confuMat(g1)
#
# computations related to ALL that were used for rda and may be used elsewhere
#
library(ALL)
data(ALL)
#
# restrict to BCR/ABL or NEG
#
bio <- which( ALL$mol.biol %in% c("BCR/ABL", "NEG"))
#
# restrict to B-cell
#
isb <- grep("^B", as.character(ALL$BT))
kp <- intersect(bio,isb)
all2 <- ALL[,kp]
mads = apply(exprs(all2),1,mad)
kp = which(mads>1) # get around 250 genes
vall2 = all2[kp, ]
vall2$mol.biol = factor(vall2$mol.biol) # drop unused levels

# illustrate clustering support

c11 = MLearn(~CW+RW+CL+FL+BD, data=crabs, hclustI(distFun=dist, cutParm=list(k=4)))
plot(c11)

c11a = MLearn(~CW+RW+CL+FL+BD, data=crabs, hclustI(distFun=dist, cutParm=list(k=4)),
  method="complete")
plot(c11a)

c12 = MLearn(~CW+RW+CL+FL+BD, data=crabs, kmeansI, centers=5, algorithm="Hartigan-Wong")
plot(c12, crabs[, -c(1:3)])

c3 = MLearn(~CL+CW+RW, crabs, pamI(dist), k=5)
c3
plot(c3, data=crabs[,c("CL", "CW", "RW")])

# new interfaces to PLS thanks to Laurent Gatto

set.seed(1234)
kp = sample(1:200, size=120)

#plsda.1 = MLearn(sp~CW+RW, data=crabs, plsdaI, kp, probMethod="Bayes")
#plsda.1
#confuMat(plsda.1)
#confuMat(plsda.1,t=.65) ## requires at least 0.65 post error prob to assign species
#
#plsda.2 = MLearn(type~., data=sample.ExpressionSet[100:250,], plsdaI, 1:16)
#plsda.2

```

```
#confuMat(plsda.2)
#confuMat(plsda.2,t=.65) ## requires at least 0.65 post error prob to assign outcome

## examples for predict
#clout <- MLearn(type~., sample.ExpressionSet[100:250,], svmI , 1:16)
#predict(clout, sample.ExpressionSet[100:250,17:26])
```

---

MLIntInternals

*MLInterfaces infrastructure*

---

## Description

These functions are internal tools for MLInterfaces. Users will generally not call these functions directly.

## Usage

```
getGrid(x)
```

## Arguments

x                    a vector or matrix or ExpressionSet

## Details

Forthcoming.

## Value

Functions with ‘new’ as prefix are constructor helpers.

## Author(s)

VJ Carey <stvjc@channing.harvard.edu>

## Description

Methods to calculate the number of true positives (tp), true negatives (tn), false negatives (fn), false positive (fp), accuracy (acc), precision, recall (same as sensitivity), specificity, F1 and macroF1 scores.

Each method also accepts an `naAs0` argument defining if NAs should be replaced by 0 (default is FALSE).

## Methods

Methods `tp`, `tn`, `fp`, `fn`, `F1`, `acc` and `specificity`:

```
signature(obj = "table")
```

Methods `recall` (sensitivity), `precision` and `macroF1`:

```
signature(obj = "classifierOutput", type = "character")
```

```
signature(obj = "classifierOutput", type = "missing")
```

```
signature(obj = "classifierOutput", type = "numeric")
```

```
signature(obj = "table")
```

## Examples

```
## the confusion matrix
cm <- table(iris$Species, sample(iris$Species))
tp(cm)
tn(cm)
fp(cm)
fn(cm)
acc(cm)
precision(cm)
recall(cm)
F1(cm)
macroF1(cm)
```

---

planarPlot-methods      *Methods for Function planarPlot in Package 'MLInterfaces'*

---

## Description

show the classification boundaries on the plane dictated by two genes in an ExpressionSet

## Methods

**clo = "classifierOutput", eset = "ExpressionSet", classifLab = "character"** uses two genes in the ExpressionSet to exhibit the decision boundaries in the plane

**clo = "classifierOutput", eset = "data.frame", classifLab = "character"** uses two columns in the data.frame to exhibit the decision boundaries in the plane

## Examples

```
library(ALL)
library(hgu95av2.db)
data(ALL)
#
# restrict to BCR/ABL or NEG
#
bio <- which( ALL$mol.biol %in% c("BCR/ABL", "NEG"))
#
# restrict to B-cell
#
isb <- grep("^B", as.character(ALL$BT))
kp <- intersect(bio,isb)
all2 <- ALL[,kp]
#
# sample 2 genes at random
#
set.seed(1234)
ng <- nrow(exprs(all2)) # pick 5 in case any NAs come back
pick <- sample(1:ng, size=5, replace=FALSE)
gg <- all2[pick,]
sym <- unlist(mget(featureNames(gg), hgu95av2SYMBOL))
bad = which(is.na(sym))
if (length(bad)>0) {
  gg = gg[-bad,]
  sym = sym[-bad]
}
gg = gg[1:2,]
sym = sym[1:2]
featureNames(gg) <- sym
gg$class = factor(ifelse(all2$mol.biol=="NEG", "NEG", "POS"))

c11 <- which( gg$class == "NEG" )
c12 <- which( gg$class != "NEG" )
```

```

#
# create balanced training sample
#
trainInds <- c( sample(c11, size=floor(length(c11)/2) ),
               sample(c12, size=floor(length(c12)/2)) )
#
# run rpart
#
tgg <- MLearn(class~., gg, rpartI, trainInds, minsplit=4 )
opar <- par(no.readonly=TRUE)
par(mfrow=c(2,2))
planarPlot( tgg, gg, "class" )
title("rpart")
points(exprs(gg)[1,trainInds], exprs(gg)[2,trainInds], col=ifelse(gg$class[trainInds]=="NEG", "yellow", "black"))
#
# run nnet
#
ngg <- MLearn( class~., gg, nnetI, trainInds, size=8 )
planarPlot( ngg, gg, "class" )
points(exprs(gg)[1,trainInds], exprs(gg)[2,trainInds], col=ifelse(gg$class[trainInds]=="NEG", "yellow", "black"))
title("nnet")
#
# run knn
#
kgg <- MLearn( class~., gg, knnI(k=3,l=1), trainInds)
planarPlot( kgg, gg, "class" )
points(exprs(gg)[1,trainInds], exprs(gg)[2,trainInds], col=ifelse(gg$class[trainInds]=="NEG", "yellow", "black"))
title("3-nn")
#
# run svm
#
sgg <- MLearn( class~., gg, svmI, trainInds )
planarPlot( sgg, gg, "class" )
points(exprs(gg)[1,trainInds], exprs(gg)[2,trainInds], col=ifelse(gg$class[trainInds]=="NEG", "yellow", "black"))
title("svm")
par(opar)

```

---

plspinHcube

*shiny app for interactive 3D visualization of mlbench hypercube*

---

### Description

shiny app for interactive 3D visualization of mlbench hypercube

### Usage

```
plspinHcube(insbwidth=4)
```

### Arguments

insbwidth      numeric, sidebar width

**Value**

Runs [shinyApp](#) on ui and server that render gaussian data at hypercube vertices.

**Author(s)**

VJ Carey <stvjc@channing.harvard.edu>

**See Also**

[mlbench.hypercube](#)

**Examples**

```
if (interactive()) plspinHcube()
```

---

predict.classifierOutput

*Predict method for classifierOutput objects*

---

**Description**

This function predicts values based on models trained with MLInterfaces' MLEarn interface to many machine learning algorithms.

**Usage**

```
## S3 method for class 'classifierOutput'
predict(object, newdata, ...)
```

**Arguments**

object	An instance of class <a href="#">classifierOutput</a> .
newdata	An object containing the new input data: either a matrix, a data.frame or an ExpressionSet.
...	Other arguments to be passed to the algorithm-specific predict methods.

**Details**

This S3 method will extract the ML model from the [classifierOutput](#) instance and call either a generic predict method or, if available, a specifically written wrapper to do classes prediction and class probabilities.

**Value**

Currently, a list with

testPredictions

A factor with class predictions.

testScores

A numeric or matrix with class probabilities.

**Note**

The function output will most likely be updated in a near future to a `classifierOutput` (or similar) object.

**Author(s)**

Laurent Gatto <lg390@cam.ac.uk>

**See Also**

[MLearn](#) and [classifierOutput](#).

**Examples**

```
## Not run:
set.seed(1234)
data(sample.ExpressionSet)
trainInd <- 1:16

cclout.svm <- MLearn(type~., sample.ExpressionSet[100:250,], svmI, trainInd)
predict(cclout.svm, sample.ExpressionSet[100:250,-trainInd])

cclout.ksvm <- MLearn(type~., sample.ExpressionSet[100:250,], ksvmI, trainInd)
predict(cclout.ksvm, sample.ExpressionSet[100:250,-trainInd])

cclout.nnet <- MLearn(type~., sample.ExpressionSet[100:250,], nnetI, trainInd, size=3, decay=.01)
predict(cclout.nnet, sample.ExpressionSet[100:250,-trainInd])

cclout.knn <- MLearn(type~., sample.ExpressionSet[100:250,], knnI(k=3), trainInd)
predict(cclout.knn, sample.ExpressionSet[100:250,-trainInd],k=1)
predict(cclout.knn, sample.ExpressionSet[100:250,-trainInd],k=3)

#cclout.plsda <- MLearn(type~., sample.ExpressionSet[100:250,], plsdaI, trainInd)
#predict(cclout.plsda, sample.ExpressionSet[100:250,-trainInd])

cclout.nb <- MLearn(type~., sample.ExpressionSet[100:250,], naiveBayesI, trainInd)
predict(cclout.nb, sample.ExpressionSet[100:250,-trainInd])

# this can fail if training set does not yield sufficient diversity in response vector;
# setting seed seems to help with this example, but other applications may have problems
#
cclout.rf <- MLearn(type~., sample.ExpressionSet[100:250,], randomForestI, trainInd)
predict(cclout.rf, sample.ExpressionSet[100:250,-trainInd])

## End(Not run) # end of dontrun
```



---

```
projectedLearner-class
      Class "projectedLearner"
```

---

### Description

helps depict prediction hyperregions from high-dimensional models

### Objects from the Class

Objects can be created by calls of the form `new("projectedLearner", ...)`.

### Slots

**fittedLearner:** Object of class "classifierOutput"  
**trainingSetPCA:** Object of class "prcomp"  
**trainingLabels:** Object of class "ANY" given labels for features used in training  
**testLabels:** Object of class "ANY" given labels for features used in testing  
**gridFeatsProjectedToTrainingPCs:** Object of class "matrix" rotated coordinates of training features  
**gridPredictions:** Object of class "ANY" predicted labels for all grid points  
**trainFeatsProjectedToTrainingPCs:** Object of class "matrix" rotated coordinates of training features  
**testFeatsProjectedToTrainingPCs:** Object of class "matrix" rotated coordinates of test features  
**trainPredictions:** Object of class "ANY" predicted labels for training features  
**testPredictions:** Object of class "ANY" predicted labels for test features  
**theCall:** Object of class "call" call used to generate this wonderful thing

### Methods

**learnerIn3D** signature(`x = "projectedLearner"`): uses `rgl` to give a dynamic 3d-like projection of labels in colored regions. See [projectLearnerToGrid](#) for an example.  
**plot** signature(`x = "projectedLearner"`, `y = "ANY"`): pairs plot of the tessellated PCA of the training features  
**plotOne** signature(`x = "projectedLearner"`): a 2d plot of tessellation projection for selected axes of the PCA  
**show** signature(`object = "projectedLearner"`): object housing numerical resources for the renderings

**Note**

plot may need to be modified when there are many features/PCs in use

plotOne has additional arguments ind1, ind2, and type. ind1 and ind2 specify the PCs to display. type is one of "showTestPredictions" (default), "showTrainPredictions", "showTestLabels", "showTrainLabels". These indicate what will be used to locate glyphs with labels in the projected scatterplots.

**Author(s)**

VJ Carey <stvjc@channing.harvard.edu>

**References**

None.

**Examples**

```
showClass("projectedLearner")
```

---

projectLearnerToGrid *create learned tessellation of feature space after PC transformation*

---

**Description**

create learned tessellation of feature space after PC transformation

**Usage**

```
projectLearnerToGrid(formula, data, learnerSchema,
  trainInds, ...,
  dropIntercept = TRUE,
  ngpts = 20,
  predExtras = list(),
  predWrapper = force)
```

**Arguments**

formula	standard formula, typically of the form "y~." where y denotes the class label variable to be predicted by all remaining features in the input data frame
data	a data.frame instance
learnerSchema	an instance of <a href="#">learnerSchema-class</a>
trainInds	integer vector of rows of data to use for training
...	additional parameters for use with learnerSchema
dropIntercept	logical indicating whether to include column of 1s among feature column-vectors

ngpts	number of equispaced points along the range of each input feature to use in forming a grid in feature space
predExtras	a list with named elements giving binding to extra parameters needed to predict labels for the learner in use. For example, with ldaI, set predExtras=list(type="class")
predWrapper	Sometimes a function call is needed to extract the predicted labels from the RObject applied to the fittedLearner slot of the output object; this parameter defines that call.

**Value**

instance of `projectedLearner-class`

**Author(s)**

VJ Carey <stvjc@channing.harvard.edu>

**References**

none.

**Examples**

```
library(mlbench)
# demonstrate with 3 dimensional hypercube problem
kk = mlbench.hypercube()
colnames(kk$x) = c("f1", "f2", "f3")
hcu = data.frame(cl=kk$classes, kk$x)
set.seed(1234)
sam = sample(1:nrow(kk$x), size=nrow(kk$x)/2)
ldap = projectLearnerToGrid(cl~., data=hcu, ldaI,
  sam, predWrapper=function(x)x$class)
plot(ldap)
confuMat(ldap@fittedLearner)
nnetp = projectLearnerToGrid(cl~., data=hcu, nnetI, sam, size=2,
  decay=.01, predExtras=list(type="class"))
plot(nnetp)
confuMat(nnetp@fittedLearner)
#if (requireNamespace("rgl") && interactive()) {
#   learnerIn3D(nnetp)
#   ## customising the rgl plot
#   learnerIn3D(nnetp, size = 10, alpha = 0.1)
#}
```

---

RAB

*real adaboost (Friedman et al)*

---

### Description

read adaboost ... a demonstration version

### Usage

```
RAB(formula, data, maxiter=200, maxdepth=1)
```

### Arguments

formula	formula – the response variable must be coded -1, 1
data	data
maxiter	maxiter
maxdepth	maxdepth – passed to rpart

### Value

an instance of raboostCont

### Author(s)

Vince Carey <stvjc@channing.harvard.edu>

### References

Friedman et al Ann Stat 28/2 337

### Examples

```
library(MASS)
library(rpart)
data(Pima.tr)
data(Pima.te)
Pima.all = rbind(Pima.tr, Pima.te)
tonp = ifelse(Pima.all$type == "Yes", 1, -1)
tonp = factor(tonp)
Pima.all = data.frame(Pima.all[,1:7], mtype=tonp)
fit1 = RAB(mtype~ped+glu+npreg+bmi+age, data=Pima.all[1:200,], maxiter=10, maxdepth=5)
pfit1 = Predict(fit1, newdata=Pima.tr)
table(Pima.tr$type, pfit1)
```

---

raboostCont-class      *Class "raboostCont" ~~~*

---

**Description**

~~ A concise (1-5 lines) description of what the class is. ~~

**Objects from the Class**

Objects can be created by calls of the form `new("raboostCont", ...)`. ~~ describe objects here  
~~

**Slots**

.Data: Object of class "list" ~~  
formula: Object of class "formula" ~~  
call: Object of class "call" ~~

**Extends**

Class "[list](#)", from data part. Class "[vector](#)", by class "list", distance 2.

**Methods**

Predict is an S4 method that can apply to instances of this class.

**Author(s)**

VJ Carey <stvjc@channing.harvard.edu>

**Examples**

```
showClass("raboostCont")
```

---

varImpStruct-class      *Class "varImpStruct" – collect data on variable importance from various machine learning methods*

---

**Description**

collects data on variable importance

**Objects from the Class**

Objects can be created by calls of the form `new("varImpStruct", ...)`. These are matrices of importance measures with separate slots identifying algorithm generating the measures and variable names.

**Slots**

**.Data:** Object of class "matrix" actual importance measures  
**method:** Object of class "character" tag  
**varnames:** Object of class "character" conformant vector of names of variables

**Extends**

Class "matrix", from data part. Class "structure", by class "matrix". Class "array", by class "matrix". Class "vector", by class "matrix", with explicit coerce. Class "vector", by class "matrix", with explicit coerce.

**Methods**

**plot** signature(x = "varImpStruct"): make a bar plot, you can supply arguments plat and toktype which will use lookUp(..., plat, toktype) from the annotate package to translate probe names to, e.g., gene symbols.

**show** signature(object = "varImpStruct"): simple abbreviated display

**getVarImp** signature(object = "classifOutput", fixNames="logical"): extractor of variable importance structure; fixNames parameter is to remove leading X used to make variable names syntactic by randomForest (ca 1/2008). You can set fixNames to false if using hu6800 platform, because all featureNames are syntactic as given.

**report** signature(object = "classifOutput", fixNames="logical"): extractor of variable importance data, with annotation; fixNames parameter is to remove leading X used to make variable names syntactic by randomForest (ca 1/2008). You can set fixNames to false if using hu6800 platform, because all featureNames are syntactic as given.

**Examples**

```
library(golubEsets)
data(Golub_Merge)
library(hu6800.db)
smallG <- Golub_Merge[1001:1060,]
set.seed(1234)
opar=par(no.readonly=TRUE)
par(las=2, mar=c(10,11,5,5))
rf2 <- MLearn(ALL.AML~, smallG, randomForestI, 1:40, importance=TRUE,
  sampsize=table(smallG$ALL.AML[1:40]), mtry=sqrt(ncol(exprs(smallG))))
plot( getVarImp( rf2, FALSE ), n=10, plat="hu6800", toktype="SYMBOL")
par(opar)
report( getVarImp( rf2, FALSE ), n=10, plat="hu6800", toktype="SYMBOL")
```

**Description**

Use cross-validation in a clustered computing environment

**Usage**

```
xvalLoop( cluster, ... )
```

**Arguments**

<code>cluster</code>	Any S4-class object, used to indicate how to perform clustered computations.
<code>...</code>	Additional arguments used to inform the clustered computation.

**Details**

Cross-validation usually involves repeated calls to the same function, but with different arguments. This provides an obvious place for using clustered computers to enhance execution. The method `xval` is structured to exploit this; `xvalLoop` provides an easy mechanism to change how `xval` performs cross-validation.

The idea is to write an `xvalLoop` method that returns a function. The function is then used to execute the cross-validation. For instance, the default method returns the function `lapply`, so the cross-validation is performed by using `lapply`. A different method might return a function that executed `lapply`-like functions, but sent different parts of the function to different computer nodes.

An accompanying vignette illustrates the technique in greater detail. An effective division of labor is for experienced cluster programmers to write `lapply`-like methods for their favored clustering environment. The user then only has to add the cluster object to the list of arguments to `xval` to get clustered calculations.

**Value**

A function taking arguments like those for [lapply](#)

**Examples**

```
## Not run:
library(golubEsets)
data(Golub_Merge)
smallG <- Golub_Merge[200:250,]

# Evaluation on one node

lk1 <- xval(smallG, "ALL.AML", knnB, xvalMethod="L00", group=as.integer(0))
table(lk1, smallG$ALL.AML)
```

```

# Evaluation on several nodes -- a cluster programmer might write the following...

library(snow)
setOldClass("spawnedMPIcluster")

setMethod("xvalLoop", signature( cluster = "spawnedMPIcluster"),
## use the function returned below to evaluate
## the central cross-validation loop in xval
function( cluster, ... ) {
  clusterExportEnv <- function( cl, env = .GlobalEnv )
  {
    unpackEnv <- function(env) {
      for ( name in ls(env) ) assign(name, get(name, env), .GlobalEnv )
      NULL
    }
    clusterCall(cl, unpackEnv, env)
  }
  function(X, FUN, ...) { # this gets returned to xval
    ## send all visible variables from the parent (i.e., xval) frame
    clusterExportEnv( cluster, parent.frame(1) )
    parLapply( cluster, X, FUN, ... )
  }
})

# ... and use the cluster like this...

cl <- makeCluster(2, "MPI")
clusterEvalQ(cl, library(MLInterfaces))

lk1 <- xval(smallG, "ALL.AML", knnB, xvalMethod="L00", group=as.integer(0), cluster = cl)
table(lk1,smallG$ALL.AML)

## End(Not run)

```

---

xvalSpec	<i>container for information specifying a cross-validated machine learning exercise</i>
----------	---

---

## Description

container for information specifying a cross-validated machine learning exercise

## Usage

```

xvalSpec(type, niter=0,
  partitionFunc = function(data, classLab, iternum) { (seq_len(nrow(data)))[-iternum] },
  fsFun = function(formula, data) formula )

```



**Arguments**

type	a string, "LOO" indicating leave-one-out cross-validation, or "LOG" indicating leave-out-group, or "NOTEST", indicating the entire dataset is used in a single training run.
niter	numeric specification of the number of cross-validation iterations to use. Ignored if type is "LOO".
partitionFunc	function, with parameters data (bound to data.frame), clab (bound to character string), iternum (bound to numeric index into sequence of 1:niter). This function's job is to provide the indices of training cases for each cross-validation step. An example is <a href="#">balKfold.xvspec</a> , which computes a series of indices that are approximately balanced with respect to frequency of outcome types.
fsFun	function, with parameters formula, data. The function must return a formula suitable for defining a model on the basis of the main input data. A candidate fsFun is given in example for fsHistory function.

**Details**

If type == "LOO", no other parameters are inspected. If type == "LOG" a value for partitionFunc must be supplied. We recommend using [balKfold.xvspec\(K\)](#). The values of niter and K in this usage must be the same. This redundancy will be removed in a future upgrade.

If the parallel package is attached and symbol mc\_fork is loaded, cross-validation will be distributed to cores using [mclapply](#).

**Value**

An instance of [classifierOutput](#), with a special structure. The RObject return slot is populated with a list of niter cross-validation results. Each element of this list is itself a list with two elements: test.idx (the indices of the test set for the associated cross-validation iteration, and mlans, the [classifierOutput](#) generated at each iteration. Thus there are classifierOutput instances nested within the main classifierOutput returned when a xvalSpec is used.

**Author(s)**

Vince Carey <stvjc@channing.harvard.edu>

**Examples**

```
library("MASS")
data(crabs)
set.seed(1234)
#
# demonstrate cross validation
#
nn1cv = MLearn(sp~CW+RW, data=crabs, nnetI, xvalSpec("LOG",
  5, balKfold.xvspec(5)), size=3, decay=.01 )
nn1cv
confuMat(nn1cv)
names(RObject(nn1cv)[[1]])
RObject(RObject(nn1cv)[[1]]$mlans)
```

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