Package 'ComPrAn'

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Type Package

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Description This package is for analysis of SILAC labeled complexome profiling data. It uses peptide table in tab-delimited format as an input and produces ready-to-use tables and plots.

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`allPeptidesPlot` *Create scatter plot*

Description

This function creates a plot of all peptides that belong to a single protein

Usage

```
allPeptidesPlot(
  .listDF,
  protein,
  max_frac,
  meanLine = FALSE,
  repPepLine = FALSE,
  separateLabStates = FALSE,
  grid = TRUE,
  titleLabel = "all",
  titleAlign = "left",
  ylabel = "Precursor Area",
  xlabel = "Fraction",
  legendLabel = "Condition",
  labelled = "Labeled",
  unlabelled = "Unlabeled",
  controlSample = "",
  textSize = 12,
  axisTextSize = 8
)
```

Arguments

.listDF	list, list containing data frames of peptides for each protein indexed by 'Protein Group Accessions'
protein	character, 'Protein Group Accession' to show in the plot
max_frac	numeric, total number of fractions
meanLine	logical, specifies whether to plot a mean line
repPepLine	logical, specifies whether to plot a representative peptide line
separateLabStates	logical, specifies whether label states will be separated into facets
grid	logical, specifies presence/absence of gridline in the plot
titleLabel	character, what to call the plot
titleAlign	character, one of the 'left', 'center'/'centre', 'right', specifies alignment of the title in plot
ylabel	character
xlabel	character
legendLabel	character
labelled	character, label to be used for isLabel == TRUE
unlabelled	character, label to be used for isLabel == FALSE
controlSample	character, either labelled or unlabelled, this setting will adjust coloring based on which sample is a control
textSize	numeric, size of text in the plot
axisTextSize	numeric, size of axis labels in the plot

Value

plot

Examples

```
##Use example peptide data set, read in and clean data
inputFile <- system.file("extData", "data.txt", package = "ComPrAn")
peptides <- peptideImport(inputFile)
peptides <- cleanData(peptides, fCol = "Search ID")
## separate chemical modifications and labelling into separate columns
peptides <- splitModLab(peptides)
## remove unneccessary columns, simplify rows
peptides <- simplifyProteins(peptides)
## Pick representative peptide for each protein for both scenarios
peptide_index <- pickPeptide(peptides)

##create a plot showing all peptides of selected protein
protein <- "P52815"
max_frac <- 23
#default plot
allPeptidesPlot(peptide_index,protein, max_frac = max_frac)
#other plot version
allPeptidesPlot(peptide_index,protein, max_frac = max_frac,
repPepLine = TRUE, grid = FALSE, titleAlign = "center")
#other plot version
```

```
allPeptidesPlot(peptide_index,protein, max_frac = max_frac,
repPepLine = TRUE, meanLine = TRUE, separateLabStates =TRUE,
titleLabel = "GN")
```

---

assignClusters *Create a data frames with cluster assignment*

Description

This function creates a data frame with column specifying clusters assigned ot each protein using the table and distance matrix produced by clusterComp() function.

Usage

```
assignClusters(.listDf, sample, method = "complete", cutoff = 0.5)
```

Arguments

.listDf	list of data frames produced by clusterComp() function
sample	which of the two samples you want to apply the function to (labeled/unlabeled).
method	character, One of 'average', 'single' or 'complete' (default), specifies the linkage method to be used inside R hclust() function
cutoff	numeric, specifies the h value in R cutree() function, height at which to 'cut the tree', everything with distance below this value is assigned into same cluster everything with larger distance is in a different cluster extreme possible values are 0 to 2 (might not be reached for all data sets)

Value

dataframe

See Also

clusterComp

Examples

```
##Use example normalised proteins file
inputFile <- system.file("extData", "dataNormProts.txt", package = "ComPrAn")
#read file in and change structure of table to required format
forAnalysis <- protImportForAnalysis(inputFile)
# create components necessary for clustering
clusteringDF <- clusterComp(forAnalysis,scenar = "A", PearsCor = "centered")
#assign clusters
labTab_clust <- assignClusters(.listDf = clusteringDF,sample = "labeled",
method = 'complete', cutoff = 0.5)
unlabTab_clust <- assignClusters(.listDf = clusteringDF,sample = "unlabeled",
                                 method = 'complete', cutoff = 0.5)
```

`cleanData` *Clean raw peptide complexomics data*

Description

Perform initial, mandatory, cleaning of data Function to process raw input data into format required for subsequent analysis. .data is a data frame containing raw input data. This function checks (not neccessarily in this order):

- renames Sequence ID column to Fraction and converts values in this column from letters to numbers
- reorders Protein Group Accessions containing multiple proteins
- removes rows in which PSM Ambiguity == 'Rejected'
- removes rows in which # Protein Groups == 0
- removes rows in which Precursor Area is NA
- removes cols that are not used at all

Usage

```
cleanData(.data, fCol = "Search ID")
```

Arguments

`.data`	dataframe
`fCol`	character The column containing the fractions, e.g. "Search ID" (default)

Value

dataframe

Author(s)

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Examples

```
##Use example peptide data set, read in and clean data
inputFile <- system.file("extData", "data.txt", package = "ComPrAn")
peptides <- peptideImport(inputFile)
peptides <- cleanData(peptides, fCol = "Search ID")
```

clusterComp *Create components necessary for clustering*

Description

Reformat the table for the one neccessary for assignClusters function. Calculate the distance matirx using selected variant of correlation.

Usage

```
clusterComp(.df, scenar = "A", PearsCor = "centered")
```

Arguments

.df	data frame, table of normalised protein values
scenar	character, scenario intended for clustering, either "A" or "B"
PearsCor	character, pearsons correlation variant (centered/uncentered)

Value

list of data frames

Examples

```
##Use example normalised proteins file
inputFile <- system.file("extData", "dataNormProts.txt", package = "ComPrAn")
#read file in and change structure of table to required format
forAnalysis <- protImportForAnalysis(inputFile)
# create components necessary for clustering
clusteringDF <- clusterComp(forAnalysis,scenar = "A", PearsCor = "centered")
```

compranApp *Execute the complexomics Shiny app*

Description

Execute the complexomics Shiny app

Usage

```
compranApp()
```

Value

Shiny app

Examples

```
#' @examples
##to start the shiny app associated with ComPrAn package run
if(interactive()){
    compranApp()
}
```

`exportClusterAssignments` *Covert clustered tables into format for export*

Description

Covert clustered tables into format for export

Usage

```
exportClusterAssignments(labClustTable, unlabClustTable)
```

Arguments

`labClustTable`	output: data frame containing columns: 'Protein Group Accessions' character 'Protein Descriptions' character 'Cluster number - unlabeled' integer 'Cluster number - labeled' integer
`unlabClustTable`	labClustTable, unlabClustTable: data frames, contain columns: 'Protein Group Accessions' character 'Protein Descriptions' character isLabel character ('TRUE'/'FALSE') - here in one data frame all are TRUE in second one all are FALSE columns 1 to n, numeric, n is the total number of fractions/slices, each of this columns contains 'Precursor Area' values in a given fraction(columns) for a protein(rows) cluster integer

Value

dataframe

Examples

```
##Use example normalised proteins file
inputFile <- system.file("extData", "dataNormProts.txt", package = "ComPrAn")
#read file in and change structure of table to required format
forAnalysis <- protImportForAnalysis(inputFile)
# create components necessary for clustering
clusteringDF <- clusterComp(forAnalysis,scenar = "A", PearsCor = "centered")
#assign clusters
labTab_clust <- assignClusters(.listDf = clusteringDF,sample = "labeled",
method = 'complete', cutoff = 0.5)
unlabTab_clust <- assignClusters(.listDf = clusteringDF,sample = "unlabeled",
                                 method = 'complete', cutoff = 0.5)
#make table of cluster assginment
tableClusterExport <- exportClusterAssignments(labTab_clust,unlabTab_clust)
```

extractRepPeps *Extract Only Data Belonging to Representative Peptide*

Description

Incomplete labelling - there are cases when in peptides containing multiple Lys/Arg not all of them are heavy in labelled samples. As in SILAC we assume that addition of label does not affect peptide properties, we are taking a mean ‘Precursor Area‘ value as the representative ‘Precursor Area‘ in such cases.

Usage

```
extractRepPeps(.data, scenario, label = "Label neccessary for scenario A")
```

Arguments

.data	dataframe containing all peptides of one protein
scenario	character "A", or "B"
label	character, selects for which label state the representative peptides will be exported, can have value of "TRUE" or "FALSE", required only for scenario "A"

Value

dataframe containing only representative peptide

getNormTable *Get normalised table for all proteins*

Description

Extracts values for representative peptides for each protein, for both scenario A and scenario B. Results are combined into one data frame in a format either indended for further analysis or for export.

Usage

```
getNormTable(.listDf, purpose = "analysis")
```

Arguments

.listDf	list of data frames
purpose	character, purpose of use of function output, values either "analysis" of "export"

Value

dataframe

Examples

```
##Use example peptide data set, read in and clean data
inputFile <- system.file("extData", "data.txt", package = "ComPrAn")
peptides <- peptideImport(inputFile)
peptides <- cleanData(peptides, fCol = "Search ID")
## separate chemical modifications and labelling into separate columns
peptides <- splitModLab(peptides)
## remove unneccessary columns, simplify rows
peptides <- simplifyProteins(peptides)
## Pick representative peptide for each protein for both scenarios
peptide_index <- pickPeptide(peptides)
## extract table with normalised protein values for both scenarios
forAnalysis <- getNormTable(peptide_index,purpose = "analysis")
```

---

groupHeatMap *Make heatmap*

Description

This function creates a heatmap for a subset of proteins in dataFrame specified in groupData, heatmap is divided into facets according to isLabel

Usage

```
groupHeatMap(
  dataFrame,
  groupData,
  groupName,
  titleAlign = "left",
  newNamesCol = NULL,
  colNumber = 2,
  ylabel = "Protein",
  xlabel = "Fraction",
  legendLabel = "Relative Protein Abundance",
  legendPosition = "right",
  grid = TRUE,
  labelled = "labeled",
  unlabelled = "unlabeled",
  orderColumn = NULL
)
```

Arguments

dataFrame	data frame, contains columns: 'Protein Group Accessions' character 'Protein Descriptions' character Fraction integer isLabel character ('TRUE'/'FALSE' values) 'Precursor Area' double scenario character
groupData	data frame, mandatory column: 'Protein Group Accessions' character - this column is used for filtering optional columns: any other column of type character that should be used for renaming
groupName	character, name that should be used for the group specified in groupData

titleAlign	character, one of the 'left', 'center'/'centre', 'right', specifies alignment of the title in plot
newNamesCol	character, if groupData contains column for re-naming and you want to use it, specify the column name in here
colNumber	numeric, values of 1 or 2, specifies whether facets will be shown side-by-side or above each other
ylabel	character
xlabel	character
legendLabel	character
legendPosition	character, one of "right" or "bottom"
grid	logical, specifies presence/absence of gridline in the plot
labelled	character, label to be used for isLabel == TRUE
unlabelled	character, label to be used for isLabel == FALSE
orderColumn	character, if groupData contains column for re-ordering and you want to use it, specify the column name in here

Value

plot

Examples

```
##Use example normalised proteins file
inputFile <- system.file("extData", "dataNormProts.txt", package = "ComPrAn")
#read file in and change structure of table to required format
forAnalysis <- protImportForAnalysis(inputFile)
##example plot:
groupDfn <- system.file("extData", "exampleGroup.txt", package = "ComPrAn")
groupName <- 'group1'
groupData <- data.table::fread(groupDfn)
groupHeatMap(forAnalysis[forAnalysis$scenario == "B",], groupData, groupName)
```

---

makeBarPlotClusterSummary *Title*

Description

Title

Usage

```
makeBarPlotClusterSummary(df, name = "sample 1")
```

Arguments

df	data frame, contains columns: 'Protein Group Accessions' character 'Protein Descriptions' character isLabel character ('TRUE'/'FALSE') columns 1 to n, numeric, n is the total number of fractions/slices, each of this columns contains 'Precursor Area' values in a given fraction(columns) for a protein(rows) cluster integer
name	character, specifies the name of the sample

Value

plot

Examples

```
##Use example normalised proteins file
inputFile <- system.file("extData", "dataNormProts.txt", package = "ComPrAn")
#read file in and change structure of table to required format
forAnalysis <- protImportForAnalysis(inputFile)
# create components necessary for clustering
clusteringDF <- clusterComp(forAnalysis,scenar = "A", PearsCor = "centered")
#assign clusters
labTab_clust <- assignClusters(.listDf = clusteringDF,sample = "labeled",
method = 'complete', cutoff = 0.5)
unlabTab_clust <- assignClusters(.listDf = clusteringDF,sample = "unlabeled",
                            method = 'complete', cutoff = 0.5)
#Make bar plots for labeled and unlabeled samples
makeBarPlotClusterSummary(labTab_clust, name = 'labeled')
makeBarPlotClusterSummary(unlabTab_clust, name = 'unlabeled')
```

---

makeDist *Make disstance matrix*

Description

This function calculates distance matrix for a data frame, column by column requires uncenteredCor function to work

Usage

```
makeDist(df, centered = FALSE)
```

Arguments

df	data frame, contains columns: 'Protein Group Accessions' character 'Protein Descriptions' character isLabel character ('TRUE'/'FALSE') columns 1 to n, numeric, n is the total number of fractions/slices, each of this columns contains 'Precursor Area' values in a given fraction(columns) for a protein(rows)
centered	centered: logical,if TRUE return dist matrix based on centered Pearson correlation (uses R cor() function, fast) ,if FALSE return dist matrix based on uncentered Pearson correlation (uses custom uncenteredCor() function, slow)

Value

matrix

---

normalizeTable *Convert extractRepPeps output to a Matrix*

Description

Convert the dataframe as output from extractRepPeps() to matrix-like table return normalized or raw values of Precursor Area, by default return normalized values

Usage

```
normalizeTable(.data, applyNormalization = TRUE)
```

Arguments

.data	a dataframe
applyNormalization	logical apply normalization or not

Value

a matrix

---

normTableForExport *Convert Normalized Dataframe to Export format*

Description

This is a convenient function for plotting

Usage

```
normTableForExport(labTab, unlabTab, comboTab)
```

Arguments

labTab	a dataframe
unlabTab	a dataframe
comboTab	a dataframe

Value

a dataframe

normTableWideToLong *Convert Normalized Dataframe To Long format*

Description

This is a convenient function for plotting

Usage

```
normTableWideToLong(labTab, unlabTab, comboTab)
```

Arguments

labTab	a dataframe
unlabTab	a dataframe
comboTab	a dataframe

Value

a dataframe

---

oneGroupTwoLabelsCoMigration
Compare a Single Group of Proteins Between Two Label States

Description

This function creates a ?scatter plot? for a subset of proteins in dataFrame specified in groupData. Intended use of the function - using scenario A data, compare shape of the migration profile for a SINGLE GROUP of proteins BETWEEN the two LABEL STATES.

Usage

```
oneGroupTwoLabelsCoMigration(
  dataFrame,
  max_frac,
  groupData = NULL,
  groupName = "group1",
  meanLine = FALSE,
  medianLine = FALSE,
  ylabel = "Relative Protein Abundance",
  xlabel = "Fraction",
  legendLabel = "Condition",
  labelled = "Labeled",
  unlabelled = "Unlabeled",
  jitterPoints = 0.3,
  pointSize = 2.5,
  grid = FALSE,
  titleAlign = "left",
```

```
  alphaValue = 1,
  controlSample = "",
  textSize = 12,
  axisTextSize = 8
)
```

Arguments

dataFrame	dataFrame: data frame, data frame of normalised values for proteins from SCENARIO A, contains columns: 'Protein Group Accessions' character 'Protein Descriptions' character Fraction integer isLabel character ('TRUE'/'FALSE' values) 'Precursor Area' double scenario character
max_frac	numeric, total number of fractions
groupData	character vector, contins list of Protein Group Accessions that belong to the group we want to plot
groupName	character, name that should be used for the group specified in groupData
meanLine	logical, specifies whether to plot a mean line for all values in the group
medianLine	logical, specifies whether to plot a median line for all values in the group
ylabel	character
xlabel	character
legendLabel	character
labelled	character, label to be used for isLabel == TRUE
unlabelled	character, label to be used for isLabel == FALSE
jitterPoints	numeric
pointSize	numeric, size of the point in the plot
grid	logical, specifies presence/absence of gridline in the plot
titleAlign	character, one of the 'left', 'center'/'centre', 'right', specifies alignment of the title in plot
alphaValue	numeric, transparency of the point, values 0 to 1
controlSample	character, either labelled or unlabelled, this setting will adjust plot coloring based on which sample is a control
textSize	numeric, size of text in the plot
axisTextSize	numeric, size of axis labels in the plot

Value

plot

Examples

```
##Use example normalised proteins file
inputFile <- system.file("extData", "dataNormProts.txt", package = "ComPrAn")
#read file in and change structure of table to required format
forAnalysis <- protImportForAnalysis(inputFile)
##example plot:
groupDV <- c("Q16540","P52815","P09001","Q13405","Q9H2W6")
groupName <- 'group1'
max_frac <- 23
oneGroupTwoLabelsCoMigration(forAnalysis, max_frac, groupDV,groupName)
```

onlyInOneLabelState *Report Proteins Present In Only One Label State*

Description

This function returns NAMES of proteins present in only labelled/only unlabelld or both label states

Usage

```
onlyInOneLabelState(.data)
```

Arguments

.data	An environment containing dataframes

Value

a list with 3 items, each item is a vector containing names belonging to one of 3 groups

Examples

```
##Use example peptide data set, read in and clean data
inputFile <- system.file("extData", "data.txt", package = "ComPrAn")
peptides <- peptideImport(inputFile)
peptides <- cleanData(peptides, fCol = "Search ID")
## separate chemical modifications and labelling into separate columns
peptides <- splitModLab(peptides)
## remove unneccessary columns, simplify rows
peptides <- simplifyProteins(peptides)
## Pick representative peptide for each protein for both scenarios
peptide_index <- pickPeptide(peptides)
## extract list of names of proteins present in one/both samples
oneStateList <- onlyInOneLabelState(peptide_index)
```

peptideImport *Import raw peptide complexomics data*

Description

Check presence of required columns inputFile is a character vector containing the location of peptide file This function checks:

- are all required columns present
- are these columns in correct format

Usage

```
peptideImport(inputFile)
```

Arguments

inputFile	character

Value

dataframe

Author(s)

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Rick Scavetta <office@scavetta.academy>

Examples

```
##Use example peptide data set, read in data
inputFile <- system.file("extData", "data.txt", package = "ComPrAn")
peptides <- peptideImport(inputFile)
```

---

pickPeptide *Select Top Peptide For Various Scenarios*

Description

This function selects a single unique peptide to represent each 'Protein Group Accession' There are 2 ways of selecting peptides, both are perform as they are needed for different tasks later on.

1. Scenario A: select peptide occuring in most fractions, do this individually for labelled/unlabelled (max value for any peptide is equal to number of fractions) in case of ties, pick peptide whith highest 'Precursor Area' in any fraction.
2. Scenario B: select peptide occuring in most fractions counting both label states together (max value for any peptide is equal to twice the number of fractions) in case of ties, pick peptide with highest 'Precursor Area' in any fraction. Representative peptide in Scenario B is picked only for proteins that have shared peptide between label states.

Usage

```
pickPeptide(.data)
```

Arguments

.data	a dataframe

Value

list of data frames

Examples

```
##Use example peptide data set, read in and clean data
inputFile <- system.file("extData", "data.txt", package = "ComPrAn")
peptides <- peptideImport(inputFile)
peptides <- cleanData(peptides, fCol = "Search ID")
## separate chemical modifications and labelling into separate columns
peptides <- splitModLab(peptides)
## remove unneccessary columns, simplify rows
peptides <- simplifyProteins(peptides)
## Pick representative peptide for each protein for both scenarios
peptide_index <- pickPeptide(peptides)
```

proteinPlot *Create Line Plots*

Description

This function creates a line plot for a proteins in dataFrame specified by protein

Usage

```
proteinPlot(
  dataFrame,
  protein,
  max_frac,
  grid = TRUE,
  titleLabel = "all",
  titleAlign = "left",
  ylabel = "Relative Protein Abundance",
  xlabel = "Fraction",
  legendLabel = "Condition",
  labelled = "Labeled",
  unlabelled = "Unlabeled",
  controlSample = "",
  textSize = 12,
  axisTextSize = 8
)
```

Arguments

dataFrame	data frame, contains columns: 'Protein Group Accessions' character; 'Protein Descriptions' character;bFraction integer; isLabel character ("TRUE"/"FALSE" values);'Precursor Area' double; scenario character
protein	character the protein of interest
max_frac	integer total number of fractions
grid	logical specifies presence/absence of gridline in the plot
titleLabel	character, if it is 'all' or 'GN', it specifies whether to show full label or just the gene name, if any other character is used, the value of titleLabel will be used as plot title
titleAlign	character one of the 'left', 'center'/'centre', 'right', specifies alignment of the title in plot
ylabel	character
xlabel	character
legendLabel	character
labelled	character label to be used for isLabel == TRUE
unlabelled	character label to be used for isLabel == FALSE
controlSample	character, either labelled or unlabelled, this setting will adjust plot coloring based on which sample is a control
textSize	numeric, size of text in the plot
axisTextSize	numeric, size of axis labels in the plot

Value

a plot

Examples

```
##Use example normalised proteins file
inputFile <- system.file("extData", "dataNormProts.txt", package = "ComPrAn")
#read file in and change structure of table to required format
forAnalysis <- protImportForAnalysis(inputFile)
##example plot:
protein <- "P52815"
max_frac <- 23
proteinPlot(forAnalysis, protein, max_frac)
```

---

protImportForAnalysis *Modify import protein data*

Description

This function converts imported protien table into format compatible with downstream analysis Imported file needs to contain following columns:

- "Protein Group Accessions" - character/factor
- "Protein Descriptions" - character
- "scenario" - character/factor
- "label" - logical
- columns "1" to "n" - numeric

Usage

```
protImportForAnalysis(inputFile)
```

Arguments

inputFile	- character vector containing the location of protein file

Value

data frame

Examples

```
##Use example normalised proteins file
inputFile <- system.file("extData", "dataNormProts.txt", package = "ComPrAn")
#read file in and change structure of table to required format
forAnalysis <- protImportForAnalysis(inputFile)
```

simplifyProteins *Simplify Raw Proteins file*

Description

- For rows: Keep only one row with highest Precursor Area in cases where for a single Protein Group Accession in a single fraction there are multiple rows with the same combination of Sequence, Mods and Charge
- For cols: remove columns that are not neccessary any more

Usage

```
simplifyProteins(.data, direction = c("rows", "cols"))
```

Arguments

.data	a dataframe
direction	character, rows, cols or both

Value

a dataframe

Examples

```
##Use example peptide data set, read in and clean data
inputFile <- system.file("extData", "data.txt", package = "ComPrAn")
peptides <- peptideImport(inputFile)
peptides <- cleanData(peptides, fCol = "Search ID")
## separate chemical modifications and labelling into separate columns
peptides <- splitModLab(peptides)
## remove unneccessary columns, simplify rows
peptides <- simplifyProteins(peptides)
```

splitModLab *Split Modification and Label tags*

Description

Splits up the Modifications column into lists of vectors for modifications(Mods) and labels(Labels) It adds two more columns to the data frame:

- UniqueCombinedID_A: Unique combinations of Sequence, Mods and Charge for "scenario A".
- UniqueCombinedID_B: Unique combinations of Sequence, Mods, Charge and Labels for "scenario B"

Usage

```
splitModLab(.data)
```

Arguments

.data	dataframe

Value

dataframe

Examples

```
##Use example peptide data set, read in and clean data
inputFile <- system.file("extData", "data.txt", package = "ComPrAn")
peptides <- peptideImport(inputFile)
peptides <- cleanData(peptides, fCol = "Search ID")
## separate chemical modifications and labelling into separate columns
peptides <- splitModLab(peptides)
```

---

toFilter *Optional Filtering For Raw Data*

Description

Filters only rows with specified values in columns Rank and Confidence Level , specified as cl

Usage

```
toFilter(.data, rank = 1, cl = c("Low", "Middle", "High"))
```

Arguments

.data	dataframe
rank	integer
cl	charater any combination of one or more of 'Low', 'Middle', or 'High'

Value

a dataframe

Examples

```
##Use example peptide data set, read in and clean data
inputFile <- system.file("extData", "data.txt", package = "ComPrAn")
peptides <- peptideImport(inputFile)
peptides <- cleanData(peptides, fCol = "Search ID")
##optional filtering based on rank and confidence level
peptides <- toFilter(peptides, rank = 1)
```

`twoGroupsWithinLabelCoMigration` *Compare a Two Groups of Proteins Within One Label State*

Description

This function creates a scatter plot for a subset of proteins in dataFrame specified in group1Data and group2Data, label states are always separated into facets

Usage

```
twoGroupsWithinLabelCoMigration(
  dataFrame,
  max_frac,
  group1Data = NULL,
  group1Name = "group1",
  group2Data = NULL,
  group2Name = "group2",
  meanLine = FALSE,
  medianLine = FALSE,
  ylabel = "Relative Protein Abundance",
  xlabel = "Fraction",
  legendLabel = "Group",
  labelled = "Labeled",
  unlabelled = "Unlabeled",
  jitterPoints = 0.3,
  pointSize = 2.5,
  grid = FALSE,
  showTitle = FALSE,
  titleAlign = "left",
  alphaValue = 1,
  textSize = 12,
  axisTextSize = 8
)
```

Arguments

`dataFrame`	dataFrame: data frame, data frame of normalised values for proteins from SCENARIO A, contains columns: 'Protein Group Accessions' character 'Protein Descriptions' character Fraction integer isLabel character ('TRUE'/'FALSE' values) 'Precursor Area' double scenario character
`max_frac`	numeric, total number of fractions
`group1Data`	character vector, contins list of Protein Group Accessions that belong to the group we want to plot for group 1
`group1Name`	character, name that should be used for the group specified in group1Data
`group2Data`	character vector, contins list of Protein Group Accessions that belong to the group we want to plot for group 2
`group2Name`	character, name that should be used for the group specified in group2Data
`meanLine`	logical, specifies whether to plot a mean line for all values in the group

medianLine	logical, specifies whether to plot a median line for all values in the group
ylabel	character
xlabel	character
legendLabel	character
labelled	character, label to be used for isLabel == TRUE
unlabelled	character, label to be used for isLabel == FALSE
jitterPoints	numeric
pointSize	numeric, size of the point in the plot
grid	logical, specifies presence/absence of gridline in the plot
showTitle	logical
titleAlign	character, one of the 'left', 'center'/'centre', 'right', specifies alignment of the title in plot
alphaValue	numeric, transparency of the point, values 0 to 1
textSize	numeric, size of text in the plot
axisTextSize	numeric, size of axis labels in the plot

Details

Intended use of the function - using scenario A data, compare shape of the migration profile between a TWO GROUPS of proteins WITHIN the ONE LABEL STATE

Value

plot

Examples

```
##Use example normalised proteins file
inputFile <- system.file("extData", "dataNormProts.txt", package ="ComPrAn")
#read file in and change structure of table to required format
forAnalysis <- protImportForAnalysis(inputFile)
##example plot:
g1D <- c("Q16540","P52815","P09001","Q13405","Q9H2W6") #group 1 data vector
g1N <- 'group1'                                        #group 1 name
g2D <- c("Q9NVS2","Q9NWU5","Q9NX20","Q9NYK5","Q9NZE8") #group 2 data vector
g2N <- 'group2'                                        #group 2 name
max_frac <- 23
twoGroupsWithinLabelCoMigration(forAnalysis, max_frac, g1D, g1N, g2D, g2N)
```

---

uncenteredCor *Perform uncentered correlation*

Description

Perform uncentered correlation

Usage

```
uncenteredCor(xx, yy)
```

Arguments

`xx`	numeric vector
`yy`	numeric vector

Value

vector

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