Package 'cTRAP'

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Title Identification of candidate causal perturbations from differential gene expression data

Version 1.30.0

Description Compare differential gene expression results with those from known cellular perturbations (such as gene knock-down, overexpression or small molecules) derived from the Connectivity Map. Such analyses allow not only to infer the molecular causes of the observed difference in gene expression but also to identify small molecules that could drive or revert specific transcriptomic alterations.

Depends R (>= 4.0)

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.plotBubbles *Plot packed bubbles*

Description

Plot packed bubbles

Usage

```
.plotBubbles(data, title, colour = "orange")
```

Arguments

data	Data to plot
title	Character: plot title
colour	Character: bubble colour

Value

highchart object

.prepareNavPage *Prepare Shiny page template*

Description

Prepare Shiny page template

Usage

```
.prepareNavPage(...)
```

Value

HTML elements

.traceInList *Find an item in list of lists and return its coordinates*

Description

Find an item in list of lists and return its coordinates

Usage

```
.traceInList(ll, item)
```

`analyseDrugSetEnrichment` *Analyse drug set enrichment*

Description

Analyse drug set enrichment

Usage

```
analyseDrugSetEnrichment(
  sets,
  stats,
  col = NULL,
  nperm = 10000,
  maxSize = 500,
  ...,
  keyColSets = NULL,
  keyColStats = NULL
)
```

Arguments

`sets`	Named list of characters: named sets containing compound identifiers (obtain drug sets by running `prepareDrugSets()`)
`stats`	Named numeric vector or either a `similarPerturbations` or a `targetingDrugs` object (obtained after running `rankSimilarPerturbations` or `predictTargetingDrugs`, respectively)
`col`	Character: name of the column to use for statistics (only required if class of `stats` is either `similarPerturbations` or `targetingDrugs`)
`nperm`	Number of permutations to do. Minimial possible nominal p-value is about 1/nperm
`maxSize`	Maximal size of a gene set to test. All pathways above the threshold are excluded.
`...`	Arguments passed on to `fgsea::fgseaSimple` `minSize` Minimal size of a gene set to test. All pathways below the threshold are excluded. `scoreType` This parameter defines the GSEA score type. Possible options are ("std", "pos", "neg"). By default ("std") the enrichment score is computed as in the original GSEA. The "pos" and "neg" score types are intended to be used for one-tailed tests (i.e. when one is interested only in positive ("pos") or negateive ("neg") enrichment). `nproc` If not equal to zero sets BPPARAM to use nproc workers (default = 0). `gseaParam` GSEA parameter value, all gene-level statis are raised to the power of ‘gseaParam‘ before calculation of GSEA enrichment scores.

BPPARAM Parallelization parameter used in bplapply. Can be used to specify cluster to run. If not initialized explicitly or by setting 'nproc' default value 'bpparam()' is used.

keyColSets Character: column from sets to compare with column keyColStats from stats; automatically selected if NULL

keyColStats Character: column from stats to compare with column keyColSets from sets; automatically selected if NULL

Value

Enrichment analysis based on GSEA

See Also

Other functions for drug set enrichment analysis: loadDrugDescriptors(), plotDrugSetEnrichment(), prepareDrugSets()

Examples

```
descriptors <- loadDrugDescriptors()
drugSets <- prepareDrugSets(descriptors)

# Analyse drug set enrichment in ranked targeting drugs for a differential
# expression profile
data("diffExprStat")
gdsc      <- loadExpressionDrugSensitivityAssociation("GDSC")
predicted <- predictTargetingDrugs(diffExprStat, gdsc)

analyseDrugSetEnrichment(drugSets, predicted)
```

---

as.table.referenceComparison

Cross Tabulation and Table Creation

Description

Cross Tabulation and Table Creation

Usage

```
## S3 method for class 'referenceComparison'
as.table(x, ..., clean = TRUE)
```

Arguments

x referenceComparison object

... Extra parameters not currently used

clean Boolean: only show certain columns (to avoid redundancy)?

Value

Complete table with metadata based on a `targetingDrugs` object

See Also

Other functions related with the ranking of CMap perturbations: `filterCMapMetadata()`, `getCMapConditions()`, `getCMapPerturbationTypes()`, `loadCMapData()`, `loadCMapZscores()`, `parseCMapID()`, `plot.perturbationChanges()`, `plot.referenceComparison()`, `plotTargetingDrugsVSsimilarPerturbations()`, `prepareCMapPerturbations()`, `print.similarPerturbations()`, `rankSimilarPerturbations()`

Other functions related with the prediction of targeting drugs: `listExpressionDrugSensitivityAssociation()`, `loadExpressionDrugSensitivityAssociation()`, `plot.referenceComparison()`, `plotTargetingDrugsVSsimilarPer`, `predictTargetingDrugs()`

`calculateCellLineMean` *Calculate cell line mean*

Description

Calculate cell line mean

Usage

```
calculateCellLineMean(data, cellLine, metadata, rankPerCellLine)
```

Arguments

`data`	Data table: comparison against CMap data
`cellLine`	Character: perturbation identifiers as names and respective cell lines as values
`metadata`	Data table: `data` metadata
`rankPerCellLine`	Boolean: rank results based on both individual cell lines and mean scores across cell lines (`TRUE`) or based on mean scores alone (`FALSE`)? If `cellLineMean = FALSE`, individual cell line conditions are always ranked.

Value

A list with two items:

`data` input `data` with extra rows containing cell line average scores (if calculated)

`rankingInfo` data table with ranking information

`metadata` metadata associated with output `data`, including for identifiers regarding mean cell line scores

`calculateEvenlyDistributedBins` *Calculate evenly-distributed bins*

Description

Calculate evenly-distributed bins

Usage

```
calculateEvenlyDistributedBins(
  numbers,
  maxBins = 15,
  k = 5,
  minPoints = NULL,
  ...,
  ids = NULL
)
```

Arguments

`numbers`	Numeric
`maxBins`	Numeric: maximum number of bins for numeric columns
`k`	Numeric: constant; the higher the constant, the smaller the bin size (check `minpts`)
`minPoints`	Numeric: minimum number of points in a bin (if `NULL`, the minimum number of points is the number of non-missing values divided by `maxBins` divided by `k`)
`...`	Arguments passed on to `binr::bins` `max.breaks` Used for initial cut. If `exact.groups` is `FALSE`, bins are merged until there's no bins with fewer than `length(x) / max.breaks` points. In `bins`, one of `max.breaks` and `minpts` must be supplied. `exact.groups` if TRUE, the result will have exactly the number of target.bins; if FALSE, the result may contain fewer than target.bins bins `verbose` Indicates verbose output. `errthresh` If the error is below the provided value, stops after the first rough estimate of the bins.

Value

Factor containing the respective group of each element in `numbers`

checkColnames *Check whether* test_names *are columns in the* data.frame

Description

Check whether test_names are columns in the data.frame

Usage

```
checkColnames(test_names, df, throw_error = TRUE)
```

Arguments

test_names	a vector of column names to test
df	the data.frame to test against
throw_error	boolean indicating whether to throw an error if any test_names are not found in df

Value

boolean indicating whether or not all test_names are columns of df

Source

https://github.com/cmap/cmapR

chunkColumns *Assign columns into chunks*

Description

Assign columns into chunks

Usage

```
chunkColumns(x, nrows, chunkGiB)
```

Arguments

x	Vector of elements
nrows	Numeric: number of rows
chunkGiB	Numeric: size (in gibibytes) of chunks to load reference file; only if argument reference is a file path

Value

List of chunks with equally distributed columns

closeOpenHandles *Close open handles*

Description

Close open handles

Usage

```
closeOpenHandles()
```

Value

Closes all open identifiers

cmapMetadata *CMap metadata*

Description

CMap metadata obtained by running the following code:

```
cmapMetadata <- filterCMapMetadata("cmapMetadata.txt", cellLine = "HEPG2",
                                   timepoint = "2 h")
```

cmapPerturbationsCompounds *CMap perturbations sample for small molecules*

Description

CMap perturbations sample for small molecules obtained by running the following code:

```
cellLine <- c("HepG2", "HUH7")
cmapMetadataCompounds <- filterCMapMetadata(
    "cmapMetadata.txt", cellLine=cellLine, timepoint="24 h",
    dosage="5 \u00B5M", perturbationType="Compound")

cmapPerturbationsCompounds <- prepareCMapPerturbations(
    cmapMetadataCompounds, "cmapZscores.gctx", "cmapGeneInfo.txt",
    "cmapCompoundInfo_drugs.txt", loadZscores=TRUE)

# Remove non-ASCII characters for portability reasons
```

```
metadata <- attr(cmapPerturbationsCompounds, "metadata")
metadata$pert_idose <- gsub("\u00B5", "micro", metadata$pert_idose)
metadata$pert_dose_unit <- gsub("\u00B5", "micro", metadata$pert_dose_unit)
attr(cmapPerturbationsCompounds, "metadata") <- metadata
```

`cmapPerturbationsKD` *CMap perturbations sample for knockdown experiments*

Description

CMap perturbations sample for knockdown experiments obtained by running the following code:

```
# Code for loading CMap gene KD HepG2 data
cellLine <- "HepG2"
cmapMetadataKD <- filterCMapMetadata(
  "cmapMetadata.txt", cellLine=cellLine,
  perturbationType="Consensus signature from shRNAs targeting the same gene")

cmapPerturbationsKD <- prepareCMapPerturbations(
  cmapMetadataKD, "cmapZscores.gctx", "cmapGeneInfo.txt",
  loadZscores=TRUE)

data("diffExprStat")
compareKD <- rankSimilarPerturbations(diffExprStat, cmapPerturbationsKD)

# Select only some perturbations (to reduce file size)
filter <- c(head(order(compareKD$spearman_rank)),
            tail(order(compareKD$spearman_rank)),
            head(order(compareKD$pearson_rank)),
            tail(order(compareKD$pearson_rank)),
            head(order(compareKD$gsea_rank)),
            tail(order(compareKD$gsea_rank)))
filter <- unique(compareKD[[1]][filter])
cmapPerturbationsKD <- cmapPerturbationsKD[ , filter]

# Remove non-ASCII characters for portability reasons
metadata <- attr(cmapPerturbationsKD, "metadata")
metadata$pert_idose <- gsub("\u00B5", "micro", metadata$pert_idose)
metadata$pert_dose_unit <- gsub("\u00B5", "micro", metadata$pert_dose_unit)
attr(cmapPerturbationsKD, "metadata") <- metadata
```

compareQuantile *Compare vector against its quantile*

Description

Check which elements of the vector are lower/greater than or equal to the quantile of a given vector.

Usage

```
compareQuantile(vec, prob, lte = FALSE)
```

Arguments

vec	Numeric vector
prob	Numeric: probability value between [0,1] to produce sample quantiles
lte	Boolean: check if values are <= quantile? If FALSE, checks if values are >= quantile

Value

Boolean vector regarding compared elements

compareWithAllMethods *Compare reference using all methods*

Description

Compare reference using all methods

Usage

```
compareWithAllMethods(
  method,
  input,
  reference,
  geneSize = 150,
  cellLines = NULL,
  cellLineMean = "auto",
  rankPerCellLine = FALSE,
  threads = 1,
  chunkGiB = 1,
  verbose = FALSE
)
```

Arguments

method	Character: comparison method (spearman, pearson or gsea; multiple methods may be selected at once)
input	Named numeric vector of differentially expressed genes whose names are gene identifiers and respective values are a statistic that represents significance and magnitude of differentially expressed genes (e.g. t-statistics); or character of gene symbols composing a gene set that is tested for enrichment in reference data (only used if method includes gsea)
reference	Data matrix or character object with file path to CMap perturbations (see prepareCMapPerturbations()) or gene expression and drug sensitivity association (see loadExpressionDrugSensitivityAssociation())
geneSize	Numeric: number of top up-/down-regulated genes to use as gene sets to test for enrichment in reference data; if a 2-length numeric vector, the first index is the number of top up-regulated genes and the second index is the number of down-regulated genes used to create gene sets; only used if method includes gsea and if input is not a gene set
cellLines	Integer: number of unique cell lines
cellLineMean	Boolean: add rows with the mean of method across cell lines? If cellLineMean = "auto" (default), rows will be added when data for more than one cell line is available.
rankPerCellLine	Boolean: rank results based on both individual cell lines and mean scores across cell lines (TRUE) or based on mean scores alone (FALSE)? If cellLineMean = FALSE, individual cell line conditions are always ranked.
threads	Integer: number of parallel threads
chunkGiB	Numeric: size (in gibibytes) of chunks to load reference file; only if argument reference is a file path
verbose	Boolean: print additional details?
rankByAscending	Boolean: rank values based on their ascending (TRUE) or descending (FALSE) order?

Value

List of data tables with correlation and/or GSEA score results

GSEA score

When method = "gsea", weighted connectivity scores (WTCS) are calculated (https://clue.io/connectopedia/cmap_algorithms).

convertENSEMBLtoGeneSymbols

Convert ENSEMBL gene identifiers to gene symbols

Description

Convert ENSEMBL gene identifiers to gene symbols

Usage

```
convertENSEMBLtoGeneSymbols(
  genes,
  dataset = "hsapiens_gene_ensembl",
  mart = "ensembl"
)
```

Arguments

genes	Character: ENSEMBL gene identifiers
dataset	Character: biomaRt dataset name
mart	Character: biomaRt database name

Value

Named character vector where names are the input ENSEMBL gene identifiers and the values are the matching gene symbols

convertGeneIdentifiers

Convert gene identifiers

Description

Convert gene identifiers

Usage

```
convertGeneIdentifiers(
  genes,
  annotation = "Homo sapiens",
  key = "ENSEMBL",
  target = "SYMBOL",
  ignoreDuplicatedTargets = TRUE
)
```

Arguments

genes	Character: genes to be converted
annotation	OrgDb with genome wide annotation for an organism or character with species name to query OrgDb, e.g. "Homo sapiens"
key	Character: type of identifier used, e.g. ENSEMBL; read ?AnnotationDbi::columns
target	Character: type of identifier to convert to; read ?AnnotationDbi::columns
ignoreDuplicatedTargets	Boolean: if TRUE, identifiers that share targets with other identifiers will not be converted

Value

Character vector of the respective targets of gene identifiers. The previous identifiers remain other identifiers have the same target (in case ignoreDuplicatedTargets = TRUE) or if no target was found.

Examples

```
genes <- c("ENSG00000012048", "ENSG00000083093", "ENSG00000141510",
           "ENSG00000051180")
convertGeneIdentifiers(genes)
convertGeneIdentifiers(genes, key="ENSEMBL", target="UNIPROT")

# Explicit species name to automatically look for its OrgDb database
sp <- "Homo sapiens"
genes <- c("ENSG00000012048", "ENSG00000083093", "ENSG00000141510",
           "ENSG00000051180")
convertGeneIdentifiers(genes, sp)

# Alternatively, set the annotation database directly
ah <- AnnotationHub::AnnotationHub()
sp <- AnnotationHub::query(ah, c("OrgDb", "Homo sapiens"))[[1]]
columns(sp) # these attributes can be used to change the attributes

convertGeneIdentifiers(genes, sp)
```

---

counts *Gene expression data sample*

Description

Gene expression data sample obtained by running the following code:

```
data("ENCODEmetadata")
ENCODEsamples <- loadENCODEsamples(ENCODEmetadata)[[1]]
counts <- prepareENCODEgeneExpression(ENCODEsamples)
```

```
# Remove low coverage (at least 10 counts shared across two samples)
minReads   <- 10
minSamples <- 2
filter <- rowSums(counts[ , -c(1, 2)] >= minReads) >= minSamples
counts <- counts[filter, ]

# Convert ENSEMBL identifier to gene symbol
counts$gene_id <- convertGeneIdentifiers(counts$gene_id)
```

---

cTRAP *cTRAP package*

Description

Compare differential gene expression results with those from big datasets (e.g. CMap), allowing to infer which types of perturbations may explain the observed difference in gene expression.

Optimised to run in ShinyProxy with Celery/Flower backend with argument `shinyproxy = TRUE`.

Usage

```
cTRAP(
  ...,
  commonPath = "data",
  expire = 14,
  fileSizeLimitMiB = 50,
  flowerURL = NULL,
  port = getOption("shiny.port"),
  host = getOption("shiny.host", "127.0.0.1")
)
```

Arguments

`...`	Objects
`commonPath`	Character: path where to store data common to all sessions
`expire`	Character: days until a session expires (message purposes only)
`fileSizeLimitMiB`	Numeric: file size limit in MiB
`flowerURL`	Character: Flower REST API's URL (`NULL` to avoid using Celery/Flower backend)
`port`	The TCP port that the application should listen on. If the `port` is not specified, and the `shiny.port` option is set (with `options(shiny.port = XX)`), then that port will be used. Otherwise, use a random port between 3000:8000, excluding ports that are blocked by Google Chrome for being considered unsafe: 3659, 4045, 5060, 5061, 6000, 6566, 6665:6669 and 6697. Up to twenty random ports will be tried.
`host`	The IPv4 address that the application should listen on. Defaults to the `shiny.host` option, if set, or `"127.0.0.1"` if not. See Details.

Details

Input: To use this package, a named vector of differentially expressed gene metric is needed, where its values represent the significance and magnitude of the differentially expressed genes (e.g. t-statistic) and its names are gene symbols.

Workflow: The differentially expressed genes will be compared against selected perturbation conditions by:

- Spearman or Pearson correlation with z-scores of differentially expressed genes after perturbations from CMap. Use function `rankSimilarPerturbations` with `method = "spearman"` or `method = "pearson"`
- Gene set enrichment analysis (GSEA) using the (around) 12 000 genes from CMap. Use function `rankSimilarPerturbations` with `method = gsea`.

Available perturbation conditions for CMap include:

- Cell line(s).
- Perturbation type (gene knockdown, gene upregulation or drug intake).
- Drug concentration.
- Time points.

Values for each perturbation type can be listed with `getCMapPerturbationTypes()`

Output: The output includes a data frame of ranked perturbations based on the associated statistical values and respective p-values.

Value

Launches result viewer and plotter (returns `NULL`)

Author(s)

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See Also

Useful links:

- https://nuno-agostinho.github.io/cTRAP
- https://github.com/nuno-agostinho/cTRAP
- Report bugs at https://github.com/nuno-agostinho/cTRAP/issues

Other visual interface functions: `launchCMapDataLoader()`, `launchDiffExprLoader()`, `launchDrugSetEnrichmentAnaly`
`launchMetadataViewer()`, `launchResultPlotter()`

diffExprStat *Differential expression's t-statistics sample*

Description

Differential expression's t-statistics sample obtained by running the following code:

```
data("counts")

# Perform differential gene expression analysis
diffExpr <- performDifferentialExpression(counts)

# Get t-statistics of differential expression with respective gene names
diffExprStat <- diffExpr$t
names(diffExprStat) <- diffExpr$Gene_symbol
```

dimnames.expressionDrugSensitivityAssociation *Operations on* expressionDrugSensitivityAssociation *objects*

Description

Operations on expressionDrugSensitivityAssociation objects

Usage

```
## S3 method for class 'expressionDrugSensitivityAssociation'
dimnames(x)

## S3 method for class 'expressionDrugSensitivityAssociation'
dim(x)

## S3 method for class 'expressionDrugSensitivityAssociation'
x[i, j, drop = FALSE, ...]
```

Arguments

x	An expressionDrugSensitivityAssociation object
i, j	Character or numeric indexes specifying elements to extract
drop	Boolean: coerce result to the lowest possible dimension?
...	Extra arguments given to other methods

Value

Subset, dimension or dimension names

downloadENCODEknockdownMetadata *Download metadata for ENCODE knockdown experiments*

Description

Download metadata for ENCODE knockdown experiments

Usage

```
downloadENCODEknockdownMetadata(
  cellLine = NULL,
  gene = NULL,
  file = "ENCODEmetadata.rds"
)
```

Arguments

cellLine	Character: cell line
gene	Character: target gene
file	Character: RDS filepath with metadata (if file doesn't exist, it will be created)

Value

Data frame containing ENCODE knockdown experiment metadata

See Also

Other functions related with using ENCODE expression data: loadENCODEsamples(), performDifferentialExpression(), prepareENCODEgeneExpression()

Examples

```
downloadENCODEknockdownMetadata("HepG2", "EIF4G1")
```

downloadIfNotFound *Download data if given file is not found*

Description

Download data if given file is not found

Usage

```
downloadIfNotFound(link, file, ask = FALSE, toExtract = NULL)
```

Arguments

link	Character: link to download file
file	Character: filepath
ask	Boolean: ask to download file?
toExtract	Character: files to extract (if NULL, extract all)

Value

Download file if file is not found

ENCODEmetadata *ENCODE metadata sample*

Description

ENCODE metadata sample obtained by running the following code:

```
gene <- "EIF4G1"
cellLine <- "HepG2"
ENCODEmetadata <- downloadENCODEknockdownMetadata(cellLine, gene)

table(ENCODEmetadata$`Experiment target`)
length(unique(ENCODEmetadata$`Experiment target`))
```

---

filterCMapMetadata *Filter CMap metadata*

Description

Filter CMap metadata

Usage

```
filterCMapMetadata(
  metadata,
  cellLine = NULL,
  timepoint = NULL,
  dosage = NULL,
  perturbationType = NULL
)
```

Arguments

metadata	Data frame (CMap metadata) or character (respective filepath)
cellLine	Character: cell line (if NULL, all values are loaded)
timepoint	Character: timepoint (if NULL, all values are loaded)
dosage	Character: dosage (if NULL, all values are loaded)
perturbationType	Character: type of perturbation (if NULL, all perturbation types are loaded)

Value

Filtered CMap metadata

See Also

Other functions related with the ranking of CMap perturbations: as.table.referenceComparison(), getCMapConditions(), getCMapPerturbationTypes(), loadCMapData(), loadCMapZscores(), parseCMapID(), plot.perturbationChanges(), plot.referenceComparison(), plotTargetingDrugsVSsimilarPertu prepareCMapPerturbations(), print.similarPerturbations(), rankSimilarPerturbations()

Examples

```
cmapMetadata <- loadCMapData("cmapMetadata.txt", "metadata")
filterCMapMetadata(cmapMetadata, cellLine="HEPG2", timepoint="2 h",
                   dosage="25 ng/mL")
```

---

findIntersectingCompounds

Check for intersecting compounds across specific columns on both datasets

---

Description

Check for intersecting compounds across specific columns on both datasets

Usage

```
findIntersectingCompounds(data1, data2, keys1 = NULL, keys2 = NULL)
```

Value

List containing three elements: matching compounds commonCompounds between column key 1 and key 2 from the first and second datasets, respectively

fix.datatypes *Adjust the data types for columns of a meta data frame*

Description

GCT(X) parsing initially returns data frames of row and column descriptors where all columns are of type character. This is inconvenient for analysis, so the goal of this function is to try and guess the appropriate data type for each column.

Usage

```
fix.datatypes(meta)
```

Arguments

meta	a data.frame

Details

This is a low-level helper function which most users will not need to access directly

Value

meta the same data frame with (potentially) adjusted column types

Source

https://github.com/cmap/cmapR

See Also

Other GCTX parsing functions: processIds(), readGctxIds(), readGctxMeta()

GCT-class *An S4 class to represent a GCT object*

Description

The GCT class serves to represent annotated matrices. The mat slot contains said data and the rdesc and cdesc slots contain data frames with annotations about the rows and columns, respectively

Slots

`mat` a numeric matrix

`rid` a character vector of row ids

`cid` a character vector of column ids

`rdesc` a `data.frame` of row descriptors

`rdesc` a `data.frame` of column descriptors

`src` a character indicating the source (usually file path) of the data

Source

https://github.com/cmap/cmapR

See Also

http://clue.io/help for more information on the GCT format

`getCMapConditions` *List available conditions in CMap datasets*

Description

Downloads metadata if not available

Usage

```
getCMapConditions(
  metadata,
  cellLine = NULL,
  timepoint = NULL,
  dosage = NULL,
  perturbationType = NULL,
  control = FALSE
)
```

Arguments

`metadata`	Data frame (CMap metadata) or character (respective filepath)
`cellLine`	Character: cell line (if `NULL`, all values are loaded)
`timepoint`	Character: timepoint (if `NULL`, all values are loaded)
`dosage`	Character: dosage (if `NULL`, all values are loaded)
`perturbationType`	Character: type of perturbation (if `NULL`, all perturbation types are loaded)
`control`	Boolean: show controls for perturbation types?

Value

List of conditions in CMap datasets

See Also

Other functions related with the ranking of CMap perturbations: as.table.referenceComparison(), filterCMapMetadata(), getCMapPerturbationTypes(), loadCMapData(), loadCMapZscores(), parseCMapID(), plot.perturbationChanges(), plot.referenceComparison(), plotTargetingDrugsVSsimilarPertu prepareCMapPerturbations(), print.similarPerturbations(), rankSimilarPerturbations()

Examples

```
## Not run:
cmapMetadata <- loadCMapData("cmapMetadata.txt", "metadata")

## End(Not run)
getCMapConditions(cmapMetadata)
```

---

getCMapPerturbationTypes

Get CMap perturbation types

Description

Get CMap perturbation types

Usage

```
getCMapPerturbationTypes(control = FALSE)
```

Arguments

control	Boolean: return perturbation types used as control?

Value

Perturbation types and respective codes as used by CMap datasets

See Also

Other functions related with the ranking of CMap perturbations: as.table.referenceComparison(), filterCMapMetadata(), getCMapConditions(), loadCMapData(), loadCMapZscores(), parseCMapID(), plot.perturbationChanges(), plot.referenceComparison(), plotTargetingDrugsVSsimilarPerturbations(), prepareCMapPerturbations(), print.similarPerturbations(), rankSimilarPerturbations()

Examples

```
getCMapPerturbationTypes()
```

getENCODEcontrols *Get experiments files for a given control*

Description

Get experiments files for a given control

Usage

```
getENCODEcontrols(control, table)
```

Arguments

control	Character: control identifier
table	Data frame

Value

Character vector with respective experiment identifiers

HTMLfast *Faster version of* shiny::HTML

Description

Faster version of shiny::HTML

Usage

```
HTMLfast(text)
```

Arguments

text	Character: text

Value

HTML element

`launchCMapDataLoader` *Load CMap data via a visual interface*

Description

Load CMap data via a visual interface

Usage

```
launchCMapDataLoader(
  metadata = "cmapMetadata.txt",
  zscores = "cmapZscores.gctx",
  geneInfo = "cmapGeneInfo.txt",
  compoundInfo = "cmapCompoundInfo.txt",
  cellLine = NULL,
  timepoint = NULL,
  dosage = NULL,
  perturbationType = NULL
)
```

Arguments

`metadata`	Data frame (CMap metadata) or character (respective filepath)
`zscores`	Data frame (GCTX z-scores) or character (respective filepath to load data from file)
`geneInfo`	Data frame (CMap gene info) or character (respective filepath to load data from file)
`compoundInfo`	Data frame (CMap compound info) or character (respective filepath to load data from file)
`cellLine`	Character: cell line (if `NULL`, all values are loaded)
`timepoint`	Character: timepoint (if `NULL`, all values are loaded)
`dosage`	Character: dosage (if `NULL`, all values are loaded)
`perturbationType`	Character: type of perturbation (if `NULL`, all perturbation types are loaded)

Value

CMap data

See Also

Other visual interface functions: `cTRAP()`, `launchDiffExprLoader()`, `launchDrugSetEnrichmentAnalyser()`, `launchMetadataViewer()`, `launchResultPlotter()`

launchDiffExprLoader *Load differential expression data via a visual interface*

Description

Currently only supports loading data from ENCODE knockdown experiments

Usage

```
launchDiffExprLoader(
  cellLine = NULL,
  gene = NULL,
  file = "ENCODEmetadata.rds",
  path = "."
)
```

Arguments

cellLine	Character: cell line
gene	Character: target gene
file	Character: RDS filepath with metadata (if file doesn't exist, it will be created)
path	Character: path where to download files

Value

Differential expression data

See Also

Other visual interface functions: cTRAP(), launchCMapDataLoader(), launchDrugSetEnrichmentAnalyser(), launchMetadataViewer(), launchResultPlotter()

launchDrugSetEnrichmentAnalyser *View and plot results via a visual interface*

Description

View and plot results via a visual interface

Usage

```
launchDrugSetEnrichmentAnalyser(sets, ...)
```

Arguments

sets	Named list of characters: named sets containing compound identifiers (obtain drug sets by running prepareDrugSets())
...	Objects

Value

Launches result viewer and plotter (returns NULL)

See Also

Other visual interface functions: cTRAP(), launchCMapDataLoader(), launchDiffExprLoader(), launchMetadataViewer(), launchResultPlotter()

---

launchMetadataViewer *View metadata via a visual interface*

Description

View metadata via a visual interface

Usage

```
launchMetadataViewer(...)
```

Arguments

...	Objects

Value

Metadata viewer (returns NULL)

See Also

Other visual interface functions: cTRAP(), launchCMapDataLoader(), launchDiffExprLoader(), launchDrugSetEnrichmentAnalyser(), launchResultPlotter()

launchResultPlotter *View and plot results via a visual interface*

Description

View and plot results via a visual interface

Usage

```
launchResultPlotter(...)
```

Arguments

...	Objects

Value

Launches result viewer and plotter (returns NULL)

See Also

Other visual interface functions: cTRAP(), launchCMapDataLoader(), launchDiffExprLoader(), launchDrugSetEnrichmentAnalyser(), launchMetadataViewer()

listExpressionDrugSensitivityAssociation *List available gene expression and drug sensitivity correlation matrices*

Description

List available gene expression and drug sensitivity correlation matrices

Usage

```
listExpressionDrugSensitivityAssociation(url = FALSE)
```

Arguments

url	Boolean: return download link?

Value

Character vector of available gene expression and drug sensitivity correlation matrices

See Also

Other functions related with the prediction of targeting drugs: `as.table.referenceComparison()`, `loadExpressionDrugSensitivityAssociation()`, `plot.referenceComparison()`, `plotTargetingDrugsVSsimilarPe`
`predictTargetingDrugs()`

Examples

```
listExpressionDrugSensitivityAssociation()
```

`loadCMapData` *Load CMap data*

Description

Load CMap data (if not found, `file` will be automatically downloaded)

Usage

```
loadCMapData(
  file,
  type = c("metadata", "geneInfo", "zscores", "compoundInfo"),
  zscoresID = NULL
)
```

Arguments

`file`	Character: path to file
`type`	Character: type of data to load (`metadata`, `geneInfo`, `zscores` or `compoundInfo`)
`zscoresID`	Character: identifiers to partially load z-scores file (for performance reasons; if `NULL`, all identifiers will be loaded)

Value

Metadata as a data table

Note

If `type = "compoundInfo"`, two files from **The Drug Repurposing Hub** will be downloaded containing information about drugs and perturbations. The files will be named `file` with `_drugs` and `_samples` before their extension, respectively.

See Also

Other functions related with the ranking of CMap perturbations: `as.table.referenceComparison()`, `filterCMapMetadata()`, `getCMapConditions()`, `getCMapPerturbationTypes()`, `loadCMapZscores()`, `parseCMapID()`, `plot.perturbationChanges()`, `plot.referenceComparison()`, `plotTargetingDrugsVSsimilarPertu`
`prepareCMapPerturbations()`, `print.similarPerturbations()`, `rankSimilarPerturbations()`

Examples

```
# Load CMap metadata (data is automatically downloaded if not available)
cmapMetadata <- loadCMapData("cmapMetadata.txt", "metadata")

# Load CMap gene info
loadCMapData("cmapGeneInfo.txt", "geneInfo")
## Not run:
# Load CMap zscores based on filtered metadata
cmapMetadataKnockdown <- filterCMapMetadata(
  cmapMetadata, cellLine="HepG2",
  perturbationType="Consensus signature from shRNAs targeting the same gene")
loadCMapData("cmapZscores.gctx.gz", "zscores", cmapMetadataKnockdown$sig_id)

## End(Not run)
```

---

loadCMapZscores *Load matrix of CMap perturbation's differential expression z-scores (optional)*

Description

Load matrix of CMap perturbation's differential expression z-scores (optional)

Usage

```
loadCMapZscores(data, inheritAttrs = FALSE, verbose = TRUE)
```

Arguments

data	perturbationChanges object
inheritAttrs	Boolean: convert to perturbationChanges object and inherit attributes from data?
verbose	Boolean: print additional details?

Value

Matrix containing CMap perturbation z-scores (genes as rows, perturbations as columns)

See Also

Other functions related with the ranking of CMap perturbations: as.table.referenceComparison(), filterCMapMetadata(), getCMapConditions(), getCMapPerturbationTypes(), loadCMapData(), parseCMapID(), plot.perturbationChanges(), plot.referenceComparison(), plotTargetingDrugsVSsimilarPertu prepareCMapPerturbations(), print.similarPerturbations(), rankSimilarPerturbations()

Examples

```
metadata <- loadCMapData("cmapMetadata.txt", "metadata")
metadata <- filterCMapMetadata(metadata, cellLine="HepG2")
## Not run:
perts <- prepareCMapPerturbations(metadata, "cmapZscores.gctx",
                                  "cmapGeneInfo.txt")
zscores <- loadCMapZscores(perts[ , 1:10])

## End(Not run)
```

---

loadCTRPgeneExpression

Load CTRP data

Description

If given paths direct to non-existing files, those files will be downloaded

Usage

```
loadCTRPgeneExpression(
  geneExpressionFile = "CTRP 2.1/geneExpr.txt",
  geneInfoFile = "CTRP 2.1/geneInfo.txt",
  cellLineInfoFile = "CTRP 2.1/cellLineInfo.txt"
)

loadCTRPdrugSensitivity(
  drugSensitivityFile = "CTRP 2.1/drugSensitivity.txt",
  experimentFile = "CTRP 2.1/experimentInfo.txt",
  compoundFile = "CTRP 2.1/compoundInfo.txt"
)

loadCTRPcompoundInfo(compoundFile = "CTRP 2.1/compoundInfo.txt")

loadNCI60geneExpression(
  file = "NCI60/geneExpr.xls",
  cellLineInfoFile = "cellLineInfo.xls"
)

loadGDSC7file(file, filename, type, ...)

loadGDSC7cellLineInfo(file = "GDSC_7/cellLineInfo.xlsx")

loadGDSC7compoundInfo(file = "GDSC_7/compoundInfo.xlsx")

loadGDSC7geneExpression(file = "GDSC_7/geneExpr.txt")

loadGDSC7drugSensitivity(file = "GDSC_7/drugs.xlsx")
```

Arguments

geneExpressionFile	Character: path to file with gene expression
geneInfoFile	Character: path to file with gene information
cellLineInfoFile	Character: path to file with cell line information
drugSensitivityFile	Character: path to file with drug sensitivity
experimentFile	Character: path to file with experiment information
compoundFile	Character: path to file with compound information
file	Character: file path

Value

Data frame

---

loadDrugDescriptors *Load table with drug descriptors*

Description

Load table with drug descriptors

Usage

```
loadDrugDescriptors(
  source = c("NCI60", "CMap"),
  type = c("2D", "3D"),
  file = NULL,
  path = NULL
)
```

Arguments

source	Character: source of compounds used to calculate molecular descriptors (NCI60 or CMap)
type	Character: load 2D or 3D molecular descriptors
file	Character: filepath to drug descriptors (automatically downloaded if file does not exist)
path	Character: folder where to find files (optional; file may contain the full filepath if preferred)

Value

Data table with drug descriptors

See Also

Other functions for drug set enrichment analysis: `analyseDrugSetEnrichment()`, `plotDrugSetEnrichment()`, `prepareDrugSets()`

Examples

```
loadDrugDescriptors()
```

---

loadENCODEsample *Load ENCODE sample*

Description

Load ENCODE sample

Usage

```
loadENCODEsample(metadata, replicate, control = FALSE, path = ".")
```

Arguments

`metadata`	Data frame: ENCODE metadata
`replicate`	Number: replicate
`control`	Boolean: load control experiment?
`path`	Character: path where to download files

Value

Data table with ENCODE sample data

---

loadENCODEsamples *Load ENCODE samples*

Description

Samples are automatically downloaded if they are not found in the current working directory.

Usage

```
loadENCODEsamples(metadata, path = ".")
```

Arguments

`metadata`	Character: ENCODE metadata
`path`	Character: path where to download files

Value

List of loaded ENCODE samples

See Also

Other functions related with using ENCODE expression data: `downloadENCODEknockdownMetadata()`, `performDifferentialExpression()`, `prepareENCODEgeneExpression()`

Examples

```
if (interactive()) {
  # Load ENCODE metadata for a specific cell line and gene
  cellLine <- "HepG2"
  gene <- c("EIF4G1", "U2AF2")
  ENCODEmetadata <- downloadENCODEknockdownMetadata(cellLine, gene)

  # Load samples based on filtered ENCODE metadata
  loadENCODEsamples(ENCODEmetadata)
}
```

---

loadExpressionDrugSensitivityAssociation

Load gene expression and drug sensitivity correlation matrix

Description

Load gene expression and drug sensitivity correlation matrix

Usage

```
loadExpressionDrugSensitivityAssociation(
  source,
  file = NULL,
  path = NULL,
  rows = NULL,
  cols = NULL,
  loadValues = FALSE
)
```

Arguments

`source`	Character: source of matrix to load; see `listExpressionDrugSensitivityAssociation`
`file`	Character: filepath to gene expression and drug sensitivity association dataset (automatically downloaded if file does not exist)
`path`	Character: folder where to find files (optional; `file` may contain the full filepath if preferred)
`rows`	Character or integer: rows

cols	Character or integer: columns
loadValues	Boolean: load data values (if available)? If FALSE, downstream functions will load and process directly from the file chunk by chunk, resulting in a lower memory footprint

Value

Correlation matrix between gene expression (rows) and drug sensitivity (columns)

See Also

Other functions related with the prediction of targeting drugs: as.table.referenceComparison(), listExpressionDrugSensitivityAssociation(), plot.referenceComparison(), plotTargetingDrugsVSsimilarPe
predictTargetingDrugs()

Examples

```
gdsc <- listExpressionDrugSensitivityAssociation()[[1]]
loadExpressionDrugSensitivityAssociation(gdsc)
```

---

loadNCI60drugSensitivity

Load CTRP data

Description

If given paths direct to non-existing files, those files will be downloaded

Usage

```
loadNCI60drugSensitivity(file = "NCI60/drugSensitivity.xls")
```

Arguments

file	Character: file path

Value

Data frame

`matchStatsWithDrugSetsID` *Match identifiers between data and drug sets*

Description

Match identifiers between data and drug sets

Usage

```
matchStatsWithDrugSetsID(
  sets,
  stats,
  col = "values",
  keyColSets = NULL,
  keyColStats = NULL
)
```

Arguments

`sets`	Named list of characters: named sets containing compound identifiers (obtain drug sets by running `prepareDrugSets()`)
`stats`	Named numeric vector or either a `similarPerturbations` or a `targetingDrugs` object (obtained after running `rankSimilarPerturbations` or `predictTargetingDrugs`, respectively)
`col`	Character: name of the column to use for statistics (only required if class of `stats` is either `similarPerturbations` or `targetingDrugs`)
`keyColSets`	Character: column from `sets` to compare with column `keyColStats` from `stats`; automatically selected if `NULL`
`keyColStats`	Character: column from `stats` to compare with column `keyColSets` from `sets`; automatically selected if `NULL`

Value

Statistic values from input data and corresponding identifiers as names (if no match is found, the original identifier from argument `stats` is used)

parseCMapID *Parse CMap identifier*

Description

Parse CMap identifier

Usage

```
parseCMapID(id, cellLine = FALSE)
```

Arguments

id	Character: CMap identifier
cellLine	Boolean: if TRUE, return cell line information from CMap identifier; else, return the CMap identifier without the cell line

Value

Character vector with information from CMap identifiers

See Also

Other functions related with the ranking of CMap perturbations: as.table.referenceComparison(), filterCMapMetadata(), getCMapConditions(), getCMapPerturbationTypes(), loadCMapData(), loadCMapZscores(), plot.perturbationChanges(), plot.referenceComparison(), plotTargetingDrugsVSsimilarP prepareCMapPerturbations(), print.similarPerturbations(), rankSimilarPerturbations()

Examples

```
id <- c("CVD001_HEPG2_24H:BRD-K94818765-001-01-0:4.8",
        "CVD001_HEPG2_24H:BRD-K96188950-001-04-5:4.3967",
        "CVD001_HUH7_24H:BRD-A14014306-001-01-1:4.1")
parseCMapID(id, cellLine=TRUE)
parseCMapID(id, cellLine=FALSE)
```

performDifferentialExpression *Perform differential gene expression based on ENCODE data*

Description

Perform differential gene expression based on ENCODE data

Usage

```
performDifferentialExpression(counts)
```

Arguments

counts	Data frame: gene expression

Value

Data frame with differential gene expression results between knockdown and control

See Also

Other functions related with using ENCODE expression data: downloadENCODEknockdownMetadata(), loadENCODEsamples(), prepareENCODEgeneExpression()

Examples

```
if (interactive()) {
  # Download ENCODE metadata for a specific cell line and gene
  cellLine <- "HepG2"
  gene <- "EIF4G1"
  ENCODEmetadata <- downloadENCODEknockdownMetadata(cellLine, gene)

  # Download samples based on filtered ENCODE metadata
  ENCODEsamples <- loadENCODEsamples(ENCODEmetadata)[[1]]

  counts <- prepareENCODEgeneExpression(ENCODEsamples)

  # Remove low coverage (at least 10 counts shared across two samples)
  minReads   <- 10
  minSamples <- 2
  filter <- rowSums(counts[ , -c(1, 2)] >= minReads) >= minSamples
  counts <- counts[filter, ]

  # Convert ENSEMBL identifier to gene symbol
  counts$gene_id <- convertGeneIdentifiers(counts$gene_id)

  # Perform differential gene expression analysis
  diffExpr <- performDifferentialExpression(counts)
}
```

---

performGSEA *Perform GSEA*

Description

Perform GSEA

Usage

```
performGSEA(pathways, stats)
```

Value

List with results of running GSEA

`plot.perturbationChanges` *Operations on a* `perturbationChanges` *object*

Description

Operations on a `perturbationChanges` object

Usage

```
## S3 method for class 'perturbationChanges'
plot(
  x,
  perturbation,
  input,
  method = c("spearman", "pearson", "gsea"),
  geneSize = 150,
  genes = c("both", "top", "bottom"),
  ...,
  title = NULL
)

## S3 method for class 'perturbationChanges'
x[i, j, drop = FALSE, ...]

## S3 method for class 'perturbationChanges'
dim(x)

## S3 method for class 'perturbationChanges'
dimnames(x)
```

Arguments

`x`	`perturbationChanges` object
`perturbation`	Character (perturbation identifier) or a `similarPerturbations` table (from which the respective perturbation identifiers are retrieved)
`input`	`Named numeric vector` of differentially expressed genes whose names are gene identifiers and respective values are a statistic that represents significance and magnitude of differentially expressed genes (e.g. t-statistics); or `character` of gene symbols composing a gene set that is tested for enrichment in reference data (only used if `method` includes `gsea`)
`method`	Character: comparison method (`spearman`, `pearson` or `gsea`; multiple methods may be selected at once)

geneSize	Numeric: number of top up-/down-regulated genes to use as gene sets to test for enrichment in reference data; if a 2-length numeric vector, the first index is the number of top up-regulated genes and the second index is the number of down-regulated genes used to create gene sets; only used if method includes gsea and if input is not a gene set
genes	Character: when plotting gene set enrichment analysis (GSEA), plot most up-regulated genes (genes = "top"), most down-regulated genes (genes = "bottom") or both (genes = "both"); only used if method = "gsea" and geneset = NULL
...	Extra arguments
title	Character: plot title (if NULL, the default title depends on the context; ignored when plotting multiple perturbations)
i, j	Character or numeric indexes specifying elements to extract
drop	Boolean: coerce result to the lowest possible dimension?

Value

Subset, plot or return dimensions or names of a perturbationChanges object

See Also

Other functions related with the ranking of CMap perturbations: as.table.referenceComparison(), filterCMapMetadata(), getCMapConditions(), getCMapPerturbationTypes(), loadCMapData(), loadCMapZscores(), parseCMapID(), plot.referenceComparison(), plotTargetingDrugsVSsimilarPerturbations(prepareCMapPerturbations(), print.similarPerturbations(), rankSimilarPerturbations()

Examples

```
data("diffExprStat")
data("cmapPerturbationsKD")

compareKD <- rankSimilarPerturbations(diffExprStat, cmapPerturbationsKD)
EIF4G1knockdown <- grep("EIF4G1", compareKD[[1]], value=TRUE)
plot(cmapPerturbationsKD, EIF4G1knockdown, diffExprStat, method="spearman")
plot(cmapPerturbationsKD, EIF4G1knockdown, diffExprStat, method="pearson")
plot(cmapPerturbationsKD, EIF4G1knockdown, diffExprStat, method="gsea")

data("cmapPerturbationsCompounds")
pert <- "CVD001_HEPG2_24H:BRD-A14014306-001-01-1:4.1"
plot(cmapPerturbationsCompounds, pert, diffExprStat, method="spearman")
plot(cmapPerturbationsCompounds, pert, diffExprStat, method="pearson")
plot(cmapPerturbationsCompounds, pert, diffExprStat, method="gsea")

# Multiple cell line perturbations
pert <- "CVD001_24H:BRD-A14014306-001-01-1:4.1"
plot(cmapPerturbationsCompounds, pert, diffExprStat, method="spearman")
plot(cmapPerturbationsCompounds, pert, diffExprStat, method="pearson")
plot(cmapPerturbationsCompounds, pert, diffExprStat, method="gsea")
```

plot.referenceComparison *Plot data comparison*

Description

If `element = NULL`, comparison is plotted based on all elements. Otherwise, show scatter or GSEA plots for a single element compared with previously given differential expression results.

Usage

```
## S3 method for class 'referenceComparison'
plot(
  x,
  element = NULL,
  method = c("spearman", "pearson", "gsea", "rankProduct"),
  n = c(3, 3),
  showMetadata = TRUE,
  plotNonRankedPerturbations = FALSE,
  alpha = 0.3,
  genes = c("both", "top", "bottom"),
  ...,
  zscores = NULL,
  title = NULL
)
```

Arguments

`x`	`referenceComparison` object: obtained after running `rankSimilarPerturbations()` or `predictTargetingDrugs()`
`element`	Character: identifier in the first column of `x`
`method`	Character: method to plot results; choose between `spearman`, `pearson`, `gsea` or `rankProduct` (the last one is only available if `element = NULL`)
`n`	Numeric: number of top and bottom genes to label (if a vector of two numbers is given, the first and second numbers will be used as the number of top and bottom genes to label, respectively); only used if `element = NULL`
`showMetadata`	Boolean: show available metadata information instead of identifiers (if available)? Only used if `element = NULL`
`plotNonRankedPerturbations`	Boolean: plot non-ranked data in grey? Only used if `element = NULL`
`alpha`	Numeric: transparency; only used if `element = NULL`
`genes`	Character: when plotting gene set enrichment analysis (GSEA), plot most up-regulated genes (`genes = "top"`), most down-regulated genes (`genes = "bottom"`) or both (`genes = "both"`); only used if `method = "gsea"` and `geneset = NULL`
`...`	Extra arguments currently not used

zscores	Data frame (GCTX z-scores) or character (respective filepath to load data from file)
title	Character: plot title (if NULL, the default title depends on the context; ignored when plotting multiple perturbations)

Value

Plot illustrating the reference comparison

See Also

Other functions related with the ranking of CMap perturbations: as.table.referenceComparison(), filterCMapMetadata(), getCMapConditions(), getCMapPerturbationTypes(), loadCMapData(), loadCMapZscores(), parseCMapID(), plot.perturbationChanges(), plotTargetingDrugsVSsimilarPerturbations(prepareCMapPerturbations(), print.similarPerturbations(), rankSimilarPerturbations()

Other functions related with the prediction of targeting drugs: as.table.referenceComparison(), listExpressionDrugSensitivityAssociation(), loadExpressionDrugSensitivityAssociation(), plotTargetingDrugsVSsimilarPerturbations(), predictTargetingDrugs()

Examples

```
# Example of a differential expression profile
data("diffExprStat")

## Not run:
# Download and load CMap perturbations to compare with
cellLine <- "HepG2"
cmapMetadataKD <- filterCMapMetadata(
  "cmapMetadata.txt", cellLine=cellLine,
  perturbationType="Consensus signature from shRNAs targeting the same gene")

cmapPerturbationsKD <- prepareCMapPerturbations(
  cmapMetadataKD, "cmapZscores.gctx", "cmapGeneInfo.txt", loadZscores=TRUE)

## End(Not run)

# Rank similar CMap perturbations
compareKD <- rankSimilarPerturbations(diffExprStat, cmapPerturbationsKD)

# Plot ranked list of CMap perturbations
plot(compareKD, method="spearman")
plot(compareKD, method="spearman", n=c(7, 3))
plot(compareKD, method="pearson")
plot(compareKD, method="gsea")

# Plot results for a single perturbation
pert <- compareKD[[1, 1]]
plot(compareKD, pert, method="spearman", zscores=cmapPerturbationsKD)
plot(compareKD, pert, method="pearson", zscores=cmapPerturbationsKD)
plot(compareKD, pert, method="gsea", zscores=cmapPerturbationsKD)
```

```
# Predict targeting drugs based on a given differential expression profile
gdsc <- loadExpressionDrugSensitivityAssociation("GDSC 7")
predicted <- predictTargetingDrugs(diffExprStat, gdsc)

# Plot ranked list of targeting drugs
plot(predicted, method="spearman")
plot(predicted, method="spearman", n=c(7, 3))
plot(predicted, method="pearson")
plot(predicted, method="gsea")

# Plot results for a single targeting drug
drug <- predicted$compound[[4]]
plot(predicted, drug, method="spearman")
plot(predicted, drug, method="pearson")
plot(predicted, drug, method="gsea")
```

`plotDrugSetEnrichment` *Plot drug set enrichment*

Description

Plot drug set enrichment

Usage

```
plotDrugSetEnrichment(
  sets,
  stats,
  col = "rankProduct_rank",
  selectedSets = NULL,
  keyColSets = NULL,
  keyColStats = NULL
)
```

Arguments

`sets`	Named list of characters: named sets containing compound identifiers (obtain drug sets by running `prepareDrugSets()`)
`stats`	Named numeric vector or either a `similarPerturbations` or a `targetingDrugs` object (obtained after running `rankSimilarPerturbations` or `predictTargetingDrugs`, respectively)
`col`	Character: name of the column to use for statistics (only required if class of `stats` is either `similarPerturbations` or `targetingDrugs`)
`selectedSets`	Character: drug sets to plot (if `NULL`, plot all)
`keyColSets`	Character: column from `sets` to compare with column `keyColStats` from `stats`; automatically selected if `NULL`
`keyColStats`	Character: column from `stats` to compare with column `keyColSets` from `sets`; automatically selected if `NULL`

Value

List of GSEA plots per drug set

See Also

Other functions for drug set enrichment analysis: `analyseDrugSetEnrichment()`, `loadDrugDescriptors()`, `prepareDrugSets()`

Examples

```
descriptors <- loadDrugDescriptors()
drugSets <- prepareDrugSets(descriptors)

# Analyse drug set enrichment in ranked targeting drugs for a differential
# expression profile
data("diffExprStat")
gdsc      <- loadExpressionDrugSensitivityAssociation("GDSC")
predicted <- predictTargetingDrugs(diffExprStat, gdsc)

plotDrugSetEnrichment(drugSets, predicted)
```

---

plotESplot *Render GSEA enrichment plot*

Description

Render GSEA enrichment plot

Usage

```
plotESplot(enrichmentScore, gseaStat, compact = FALSE)
```

Value

GSEA enrichment plot

---

plotGSEA *Plot gene set enrichment analysis (GSEA)*

Description

Plot gene set enrichment analysis (GSEA)

Usage

```
plotGSEA(
  stats,
  geneset,
  genes = c("both", "top", "bottom"),
  title = "GSEA plot",
  gseaParam = 1,
  compact = FALSE
)
```

Arguments

stats	Named numeric vector: statistics
genes	Character: when plotting gene set enrichment analysis (GSEA), plot most up-regulated genes (genes = "top"), most down-regulated genes (genes = "bottom") or both (genes = "both"); only used if method = "gsea" and geneset = NULL
title	Character: plot title (if NULL, the default title depends on the context; ignored when plotting multiple perturbations)
gseaParam	Numeric: GSEA-like parameter
compact	Boolean: render a compact version of the GSEA plot?

Value

Grid of plots illustrating a GSEA plot

---

plotMetricDistribution

Plot metric distribution

Description

Plot metric distribution

Usage

```
plotMetricDistribution(stat, compact = FALSE)
```

Value

Metric distribution plot

plotSingleCorr *Render scatter plot to show a single relationship*

Description

Render scatter plot to show a single relationship

Usage

```
plotSingleCorr(perturbation, ylabel, diffExprGenes, title = NULL)
```

Arguments

perturbation	List of named numeric vectors containing the differential expression profile score per gene for a perturbation; each perturbation of the list will be rendered with a different colour
ylabel	Character: Y axis label
diffExprGenes	Named numeric vector
title	Character: plot title (if NULL, the default title depends on the context; ignored when plotting multiple perturbations)

Value

Scatter plot

plotTargetingDrugsVSsimilarPerturbations *Plot similar perturbations against predicted targeting drugs*

Description

Plot similar perturbations against predicted targeting drugs

Usage

```
plotTargetingDrugsVSsimilarPerturbations(
  targetingDrugs,
  similarPerturbations,
  column,
  labelBy = "pert_iname",
  quantileThreshold = 0.25,
  showAllScores = FALSE,
  keyColTargetingDrugs = NULL,
  keyColSimilarPerturbations = NULL
)
```

Arguments

targetingDrugs	targetingDrugs object
similarPerturbations	similarPerturbations object
column	Character: column to plot (must be available in both databases)
labelBy	Character: column in as.table(similarPerturbations) or as.table(targetingDrugs) to be used for labelling
quantileThreshold	Numeric: quantile (between 0 and 1) to highlight values of interest
showAllScores	Boolean: show all scores? If FALSE, only the best score per compound will be plotted
keyColTargetingDrugs	Character: column from targetingDrugs to compare with column keyColSimilarPerturbations from similarPerturbations; automatically selected if NULL
keyColSimilarPerturbations	Character: column from similarPerturbations to compare with column keyColTargetingDrugs from targetingDrugs; automatically selected if NULL

Value

ggplot2 plot

See Also

Other functions related with the ranking of CMap perturbations: as.table.referenceComparison(), filterCMapMetadata(), getCMapConditions(), getCMapPerturbationTypes(), loadCMapData(), loadCMapZscores(), parseCMapID(), plot.perturbationChanges(), plot.referenceComparison(), prepareCMapPerturbations(), print.similarPerturbations(), rankSimilarPerturbations()

Other functions related with the prediction of targeting drugs: as.table.referenceComparison(), listExpressionDrugSensitivityAssociation(), loadExpressionDrugSensitivityAssociation(), plot.referenceComparison(), predictTargetingDrugs()

Examples

```
# Rank similarity against CMap compound perturbations
similarPerts <- rankSimilarPerturbations(diffExprStat,
                                         cmapPerturbationsCompounds)

# Predict targeting drugs
gdsc <- loadExpressionDrugSensitivityAssociation("GDSC 7")
predicted <- predictTargetingDrugs(diffExprStat, gdsc)

plotTargetingDrugsVSsimilarPerturbations(predicted, similarPerts,
                                         "spearman_rank")
```

predictTargetingDrugs *Predict targeting drugs*

Description

Identify compounds that may target the phenotype associated with a user-provided differential expression profile by comparing such against a correlation matrix of gene expression and drug sensitivity.

Usage

```
predictTargetingDrugs(
  input,
  expressionDrugSensitivityCor,
  method = c("spearman", "pearson", "gsea"),
  geneSize = 150,
  isDrugActivityDirectlyProportionalToSensitivity = NULL,
  threads = 1,
  chunkGiB = 1,
  verbose = FALSE
)
```

Arguments

input	Named numeric vector of differentially expressed genes whose names are gene identifiers and respective values are a statistic that represents significance and magnitude of differentially expressed genes (e.g. t-statistics); or character of gene symbols composing a gene set that is tested for enrichment in reference data (only used if method includes gsea)
expressionDrugSensitivityCor	Matrix or character: correlation matrix of gene expression (rows) and drug sensitivity (columns) across cell lines or path to file containing such data; see loadExpressionDrugSensitivityAssociation().
method	Character: comparison method (spearman, pearson or gsea; multiple methods may be selected at once)
geneSize	Numeric: number of top up-/down-regulated genes to use as gene sets to test for enrichment in reference data; if a 2-length numeric vector, the first index is the number of top up-regulated genes and the second index is the number of down-regulated genes used to create gene sets; only used if method includes gsea and if input is not a gene set
isDrugActivityDirectlyProportionalToSensitivity	Boolean: are the values used for drug activity directly proportional to drug sensitivity? If NULL, the argument expressionDrugSensitivityCor must have a non-NULL value for attribute isDrugActivityDirectlyProportionalToSensitivity.
threads	Integer: number of parallel threads

chunkGiB	Numeric: if second argument is a path to an HDF5 file (.h5 extension), that file is loaded and processed in chunks of a given size in gibibytes (GiB); lower values decrease peak RAM usage (see details below)
verbose	Boolean: print additional details?

Value

Data table with correlation and/or GSEA score results

Process data by chunks

If a file path to a valid HDF5 (.h5) file is provided instead of a data matrix, that file can be loaded and processed in chunks of size chunkGiB, resulting in decreased peak memory usage.

The default value of 1 GiB (1 GiB = 1024^3 bytes) allows loading chunks of ~10000 columns and 14000 rows (10000 * 14000 * 8 bytes / 1024^3 = 1.04 GiB).

GSEA score

When method = "gsea", weighted connectivity scores (WTCS) are calculated (https://clue.io/connectopedia/cmap_algorithms).

See Also

Other functions related with the prediction of targeting drugs: as.table.referenceComparison(), listExpressionDrugSensitivityAssociation(), loadExpressionDrugSensitivityAssociation(), plot.referenceComparison(), plotTargetingDrugsVSsimilarPerturbations()

Examples

```
# Example of a differential expression profile
data("diffExprStat")

# Load expression and drug sensitivity association derived from GDSC data
gdsc <- loadExpressionDrugSensitivityAssociation("GDSC 7")

# Predict targeting drugs on a differential expression profile
predictTargetingDrugs(diffExprStat, gdsc)
```

---

prepareCMapPerturbations

Prepare CMap perturbation data

Description

Prepare CMap perturbation data

Usage

```
prepareCMapPerturbations(
  metadata,
  zscores,
  geneInfo,
  compoundInfo = NULL,
  ...,
  loadZscores = FALSE
)
```

Arguments

metadata	Data frame (CMap metadata) or character (respective filepath to load data from file)
zscores	Data frame (GCTX z-scores) or character (respective filepath to load data from file)
geneInfo	Data frame (CMap gene info) or character (respective filepath to load data from file)
compoundInfo	Data frame (CMap compound info) or character (respective filepath to load data from file)
...	Arguments passed on to filterCMapMetadata cellLine Character: cell line (if NULL, all values are loaded) timepoint Character: timepoint (if NULL, all values are loaded) dosage Character: dosage (if NULL, all values are loaded) perturbationType Character: type of perturbation (if NULL, all perturbation types are loaded)
loadZscores	Boolean: load matrix of perturbation z-scores? Not recommended in systems with less than 30GB of RAM; if FALSE, downstream functions will load and process the file directly chunk by chunk, resulting in a lower memory footprint

Value

CMap perturbation data attributes and filename

See Also

Other functions related with the ranking of CMap perturbations: as.table.referenceComparison(), filterCMapMetadata(), getCMapConditions(), getCMapPerturbationTypes(), loadCMapData(), loadCMapZscores(), parseCMapID(), plot.perturbationChanges(), plot.referenceComparison(), plotTargetingDrugsVSsimilarPerturbations(), print.similarPerturbations(), rankSimilarPerturbations()

Examples

```
metadata <- loadCMapData("cmapMetadata.txt", "metadata")
metadata <- filterCMapMetadata(metadata, cellLine="HepG2")
## Not run:
prepareCMapPerturbations(metadata, "cmapZscores.gctx", "cmapGeneInfo.txt")
```

```
## End(Not run)
```

`prepareDrugSets` *Prepare drug sets from a table with compound descriptors*

Description

Create a list of drug sets for each character and numeric column. For each character column, drugs are split across that column's unique values (see argument `maxUniqueElems`). For each numeric column, drugs are split across evenly-distributed bins.

Usage

```
prepareDrugSets(
  table,
  id = 1,
  maxUniqueElems = 15,
  maxBins = 15,
  k = 5,
  minPoints = NULL
)
```

Arguments

`table`	Data frame: drug descriptors
`id`	Integer or character: index or name of the identifier column
`maxUniqueElems`	Numeric: ignore character columns with more unique elements than `maxUniqueElems`
`maxBins`	Numeric: maximum number of bins for numeric columns
`k`	Numeric: constant; the higher the constant, the smaller the bin size (check `minpts`)
`minPoints`	Numeric: minimum number of points in a bin (if `NULL`, the minimum number of points is the number of non-missing values divided by `maxBins` divided by `k`)

Value

Named list of characters: named drug sets with respective compound identifiers as list elements

See Also

Other functions for drug set enrichment analysis: `analyseDrugSetEnrichment()`, `loadDrugDescriptors()`, `plotDrugSetEnrichment()`

Examples

```
descriptors <- loadDrugDescriptors("NCI60")
prepareDrugSets(descriptors)
```

prepareENCODEgeneExpression *Load ENCODE gene expression data*

Description

Load ENCODE gene expression data

Usage

```
prepareENCODEgeneExpression(samples)
```

Arguments

samples	List of loaded ENCODE samples

Value

Data frame containing gene read counts

See Also

convertGeneIdentifiers()

Other functions related with using ENCODE expression data: downloadENCODEknockdownMetadata(), loadENCODEsamples(), performDifferentialExpression()

Examples

```
if (interactive()) {
  # Load ENCODE metadata for a specific cell line and gene
  cellLine <- "HepG2"
  gene <- "EIF4G1"
  ENCODEmetadata <- downloadENCODEknockdownMetadata(cellLine, gene)

  # Load samples based on filtered ENCODE metadata
  ENCODEsamples <- loadENCODEsamples(ENCODEmetadata)[[1]]

  prepareENCODEgeneExpression(ENCODEsamples)
}
```

prepareExpressionDrugSensitivityAssociation *Prepare gene expression and drug sensitivity correlation matrix*

Description

Prepare gene expression and drug sensitivity correlation matrix

Usage

```
prepareExpressionDrugSensitivityAssociation(
  dataset = c("GDSC 7", "CTRP 2.1", "NCI60"),
  method = "spearman"
)
```

Arguments

dataset	Character: dataset to use (CTRP, GDSC or NCI60)
method	Character: correlation method to use between gene expression and drug sensitivity

Details

If path directs to non-existing files, data will be downloaded.

Value

Correlation matrix between gene expression and drug sensitivity

prepareGSEAgenesets *Prepare GSEA gene sets*

Description

Prepare GSEA gene sets

Usage

```
prepareGSEAgenesets(input, geneSize)
```

Arguments

input	Named numeric vector of differentially expressed genes whose names are gene identifiers and respective values are a statistic that represents significance and magnitude of differentially expressed genes (e.g. t-statistics); or character of gene symbols composing a gene set that is tested for enrichment in reference data (only used if method includes gsea)
geneSize	Numeric: number of top up-/down-regulated genes to use as gene sets to test for enrichment in reference data; if a 2-length numeric vector, the first index is the number of top up-regulated genes and the second index is the number of down-regulated genes used to create gene sets; only used if method includes gsea and if input is not a gene set

Value

List of gene sets

---

prepareSetsCompoundInfo

Get drug sets' compound info

Description

Get drug sets' compound info

Usage

```
prepareSetsCompoundInfo(sets)
```

Arguments

sets	Named list of characters: named sets containing compound identifiers (obtain drug sets by running prepareDrugSets())

Value

List containing drug sets' compound info

prepareStatsCompoundInfo *Prepare stats' compound information*

Description

Prepare stats' compound information

Usage

```
prepareStatsCompoundInfo(stats)
```

Arguments

stats	Named numeric vector or either a similarPerturbations or a targetingDrugs object (obtained after running rankSimilarPerturbations or predictTargetingDrugs, respectively)

Value

List containing stats' compound info

prepareWordBreak *Create word break opportunities (for HTML) using given characters*

Description

Create word break opportunities (for HTML) using given characters

Usage

```
prepareWordBreak(
  str,
  pattern = c(".", "-", "\\", "/", "_", ",", " ", "+", "="),
  html = TRUE
)
```

Arguments

str	Character: text
pattern	Character: pattern(s) of interest to be used as word break opportunities
html	Boolean: convert to HTML?

Value

String containing HTML elements

print.similarPerturbations *Print a* similarPerturbations *object*

Description

Print a similarPerturbations object

Usage

```
## S3 method for class 'similarPerturbations'
print(x, perturbation = NULL, ...)
```

Arguments

x	similarPerturbations object
perturbation	Character (perturbation identifier) or numeric (perturbation index)
...	Extra parameters passed to print

Value

Information on perturbationChanges object or on specific perturbations

See Also

Other functions related with the ranking of CMap perturbations: as.table.referenceComparison(), filterCMapMetadata(), getCMapConditions(), getCMapPerturbationTypes(), loadCMapData(), loadCMapZscores(), parseCMapID(), plot.perturbationChanges(), plot.referenceComparison(), plotTargetingDrugsVSsimilarPerturbations(), prepareCMapPerturbations(), rankSimilarPerturbations()

processByChunks *Process data by chunks*

Description

Process data by chunks

Usage

```
processByChunks(
  data,
  FUN,
  num,
  ...,
  threads = 1,
  chunkGiB = 1,
  verbose = FALSE
)
```

Arguments

data	Character containing a HDF5 file path (allowing partial loading) or data matrix (processed as single chunk if data matrix)
FUN	Function: function to run for each chunk
num	Numeric: numbers of methods to run per chunk
...	Arguments passed to FUN
threads	Integer: number of parallel threads
chunkGiB	Numeric: size (in gibibytes) of chunks to load reference file; only if argument reference is a file path
verbose	Boolean: print additional details?

Value

Results of running FUN

Note

All rows from file are currently loaded when processing chunks.

---

processIds *Return a subset of requested GCTX row/column ids out of the universe of all ids*

Description

Return a subset of requested GCTX row/column ids out of the universe of all ids

Usage

```
processIds(ids, all_ids, type = "rid")
```

Arguments

ids	vector of requested ids. If NULL, no subsetting is performed
all_ids	vector of universe of ids
type	flag indicating the type of ids being processed

Details

This is a low-level helper function which most users will not need to access directly

Value

a list with the following elements ids: a character vector of the processed ids idx: an integer list of their corresponding indices in all_ids

Source

https://github.com/cmap/cmapR

See Also

Other GCTX parsing functions: fix.datatypes(), readGctxIds(), readGctxMeta()

---

rankAgainstReference *Compare multiple methods and rank against reference accordingly*

Description

Compare multiple methods and rank against reference accordingly

Usage

```
rankAgainstReference(
  input,
  reference,
  method = c("spearman", "pearson", "gsea"),
  geneSize = 150,
  cellLines = NULL,
  cellLineMean = "auto",
  rankByAscending = TRUE,
  rankPerCellLine = FALSE,
  threads = 1,
  chunkGiB = 1,
  verbose = FALSE
)
```

Arguments

input	Named numeric vector of differentially expressed genes whose names are gene identifiers and respective values are a statistic that represents significance and magnitude of differentially expressed genes (e.g. t-statistics); or character of gene symbols composing a gene set that is tested for enrichment in reference data (only used if method includes gsea)
reference	Data matrix or character object with file path to CMap perturbations (see prepareCMapPerturbations()) or gene expression and drug sensitivity association (see loadExpressionDrugSensitivityAssociation())
method	Character: comparison method (spearman, pearson or gsea; multiple methods may be selected at once)
geneSize	Numeric: number of top up-/down-regulated genes to use as gene sets to test for enrichment in reference data; if a 2-length numeric vector, the first index is the number of top up-regulated genes and the second index is the number of down-regulated genes used to create gene sets; only used if method includes gsea and if input is not a gene set

cellLines	Integer: number of unique cell lines
cellLineMean	Boolean: add rows with the mean of method across cell lines? If cellLineMean = "auto" (default), rows will be added when data for more than one cell line is available.
rankByAscending	Boolean: rank values based on their ascending (TRUE) or descending (FALSE) order?
rankPerCellLine	Boolean: rank results based on both individual cell lines and mean scores across cell lines (TRUE) or based on mean scores alone (FALSE)? If cellLineMean = FALSE, individual cell line conditions are always ranked.
threads	Integer: number of parallel threads
chunkGiB	Numeric: if second argument is a path to an HDF5 file (.h5 extension), that file is loaded and processed in chunks of a given size in gibibytes (GiB); lower values decrease peak RAM usage (see details below)
verbose	Boolean: print additional details?

Value

Data table with correlation and/or GSEA score results

Process data by chunks

If a file path to a valid HDF5 (.h5) file is provided instead of a data matrix, that file can be loaded and processed in chunks of size chunkGiB, resulting in decreased peak memory usage.

The default value of 1 GiB (1 GiB = 1024^3 bytes) allows loading chunks of ~10000 columns and 14000 rows (10000 * 14000 * 8 bytes / 1024^3 = 1.04 GiB).

GSEA score

When method = "gsea", weighted connectivity scores (WTCS) are calculated (https://clue.io/connectopedia/cmap_algorithms).

---

rankColumns *Rank columns in a dataset*

Description

Rank columns in a dataset

Usage

```
rankColumns(table, rankingInfo, rankByAscending = TRUE, sort = FALSE)
```

Arguments

`table`	Data table: data; first column must be identifiers
`rankingInfo`	Data table: boolean values of which rows to rank based on columns (column names to be ranked must exactly match those available in argument `table`); first column must be identifiers
`rankByAscending`	Boolean: rank values based on their ascending (`TRUE`) or descending (`FALSE`) order?
`sort`	Boolean: sort data based on rank product's rank (if multiple methods are available) or by available ranks

Details

The rank product's rank is calculated if more than one method is ranked.

Value

Data table with the contents of `table` and extra columns with respective rankings

Note

The first column of `data` and `rankingInfo` must contain common identifiers.

`rankSimilarPerturbations` *Rank differential expression profile against CMap perturbations by similarity*

Description

Compare differential expression results against CMap perturbations.

Usage

```
rankSimilarPerturbations(
  input,
  perturbations,
  method = c("spearman", "pearson", "gsea"),
  geneSize = 150,
  cellLineMean = "auto",
  rankPerCellLine = FALSE,
  threads = 1,
  chunkGiB = 1,
  verbose = FALSE
)
```

Arguments

input	Named numeric vector of differentially expressed genes whose names are gene identifiers and respective values are a statistic that represents significance and magnitude of differentially expressed genes (e.g. t-statistics); or character of gene symbols composing a gene set that is tested for enrichment in reference data (only used if method includes gsea)
perturbations	perturbationChanges object: CMap perturbations (check prepareCMapPerturbations())
method	Character: comparison method (spearman, pearson or gsea; multiple methods may be selected at once)
geneSize	Numeric: number of top up-/down-regulated genes to use as gene sets to test for enrichment in reference data; if a 2-length numeric vector, the first index is the number of top up-regulated genes and the second index is the number of down-regulated genes used to create gene sets; only used if method includes gsea and if input is not a gene set
cellLineMean	Boolean: add rows with the mean of method across cell lines? If cellLineMean = "auto" (default), rows will be added when data for more than one cell line is available.
rankPerCellLine	Boolean: rank results based on both individual cell lines and mean scores across cell lines (TRUE) or based on mean scores alone (FALSE)? If cellLineMean = FALSE, individual cell line conditions are always ranked.
threads	Integer: number of parallel threads
chunkGiB	Numeric: if second argument is a path to an HDF5 file (.h5 extension), that file is loaded and processed in chunks of a given size in gibibytes (GiB); lower values decrease peak RAM usage (see details below)
verbose	Boolean: print additional details?

Value

Data table with correlation and/or GSEA score results

Process data by chunks

If a file path to a valid HDF5 (.h5) file is provided instead of a data matrix, that file can be loaded and processed in chunks of size chunkGiB, resulting in decreased peak memory usage.

The default value of 1 GiB (1 GiB = 1024^3 bytes) allows loading chunks of ~10000 columns and 14000 rows (10000 * 14000 * 8 bytes / 1024^3 = 1.04 GiB).

GSEA score

When method = "gsea", weighted connectivity scores (WTCS) are calculated (https://clue.io/connectopedia/cmap_algorithms).

See Also

Other functions related with the ranking of CMap perturbations: as.table.referenceComparison(), filterCMapMetadata(), getCMapConditions(), getCMapPerturbationTypes(), loadCMapData(), loadCMapZscores(), parseCMapID(), plot.perturbationChanges(), plot.referenceComparison(), plotTargetingDrugsVSsimilarPerturbations(), prepareCMapPerturbations(), print.similarPerturbations()

Examples

```
# Example of a differential expression profile
data("diffExprStat")

## Not run:
# Download and load CMap perturbations to compare with
cellLine <- c("HepG2", "HUH7")
cmapMetadataCompounds <- filterCMapMetadata(
    "cmapMetadata.txt", cellLine=cellLine, timepoint="24 h",
    dosage="5 \u00B5M", perturbationType="Compound")

cmapPerturbationsCompounds <- prepareCMapPerturbations(
    cmapMetadataCompounds, "cmapZscores.gctx", "cmapGeneInfo.txt",
    "cmapCompoundInfo_drugs.txt", loadZscores=TRUE)

## End(Not run)
perturbations <- cmapPerturbationsCompounds

# Rank similar CMap perturbations (by default, Spearman's and Pearson's
# correlation are used, as well as GSEA with the top and bottom 150 genes of
# the differential expression profile used as reference)
rankSimilarPerturbations(diffExprStat, perturbations)

# Rank similar CMap perturbations using only Spearman's correlation
rankSimilarPerturbations(diffExprStat, perturbations, method="spearman")
```

---

readGctxIds *Read GCTX row or column ids*

Description

Read GCTX row or column ids

Usage

```
readGctxIds(gctx_path, dimension = "row")
```

Arguments

gctx_path	path to the GCTX file
dimension	which ids to read (row or column)

Value

a character vector of row or column ids from the provided file

Source

https://github.com/cmap/cmapR

See Also

Other GCTX parsing functions: fix.datatypes(), processIds(), readGctxMeta()

---

readGctxMeta *Parse row or column metadata from GCTX files*

Description

Parse row or column metadata from GCTX files

Usage

```
readGctxMeta(
  gctx_path,
  dimension = "row",
  ids = NULL,
  set_annot_rownames = TRUE
)
```

Arguments

gctx_path	the path to the GCTX file
dimension	which metadata to read (row or column)
ids	a character vector of a subset of row/column ids for which to read the metadata
set_annot_rownames	a boolean indicating whether to set the rownames attribute of the returned data.frame to the corresponding row/column ids.

Value

a data.frame of metadata

Source

https://github.com/cmap/cmapR

See Also

Other GCTX parsing functions: fix.datatypes(), processIds(), readGctxIds()

stripStr *Strip non-alpha-numeric characters from a string*

Description

Strip non-alpha-numeric characters from a string

Usage

```
stripStr(str)
```

Arguments

str	Character

Value

Character without non-alphanumeric values

subsetData *Subset data by rows and/or columns*

Description

Subset data by rows and/or columns

Usage

```
subsetData(x, i, j, rowAttr, colAttr, nargs, ...)
```

Value

Subset data

subsetDim *Subset rows or columns based on a given index*

Description

Subset rows or columns based on a given index

Usage

```
subsetDim(k, dims, nargs, areCols = TRUE)
```

Value

Subset rows/columns

subsetToIds *Do a robust* data.frame *subset to a set of ids*

Description

Do a robust data.frame subset to a set of ids

Usage

```
subsetToIds(df, ids)
```

Arguments

df	data.frame to subset
ids	the ids to subset to

Value

a subset version of df

Source

https://github.com/cmap/cmapR

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