# Package 'RgnTX'

May 11, 2024

**Title** Colocalization analysis of transcriptome elements in the presence of isoform heterogeneity and ambiguity

Version 1.6.0

**Description** RgnTX allows the integration of transcriptome annotations so as to model the complex alternative splicing patterns. It supports the testing of transcriptome elements without clear isoform association, which is often the real scenario due to technical limitations. It involves functions that do permutaion test for evaluating association between features and transcriptome regions.

**License** Artistic-2.0 **Encoding** UTF-8

**Roxygen** list(markdown = TRUE)

RoxygenNote 7.2.0

**biocViews** AlternativeSplicing, Sequencing, RNASeq, MethylSeq, Transcription, SplicedAlignment

Imports GenomeInfoDb, GenomicFeatures, GenomicRanges, ggplot2, graphics, IRanges, methods, regioneR, S4Vectors, stats, TxDb.Hsapiens.UCSC.hg19.knownGene

**Depends** R (>= 4.2.0)

**Suggests** BiocStyle, rmarkdown, knitr, testthat (>= 3.0.0)

VignetteBuilder knitr

Config/testthat/edition 3

git\_url https://git.bioconductor.org/packages/RgnTX

git\_branch RELEASE\_3\_19

git\_last\_commit b54e1c6

git\_last\_commit\_date 2024-04-30

**Repository** Bioconductor 3.19

**Date/Publication** 2024-05-10

Author Yue Wang [aut, cre],

Jia Meng [aut]

2 calculateShift

# **Contents**

	ulateShift Calculate positional shifting over transcriptome	
Index		31
	vector2GRangesDist	50
	vector2GRangesList	
	shiftTx	29
	shiftedZScoreTx	28
	randomizeTx	27
	randomizeTransByOrder	26
	randomizeFeaturesTxIA	25
	randomizeFeaturesTx	24
	plotShiftedZScoreTx	24
	plotPermResults	23
	permTestTx_customPick	21
	permTestTx_customAll	20
	permTestTxIA_customPick	18
	permTestTxIA	16
	permTestTx	15
	overlapWidthTx	14
	overlapCountsTxIA	13
	overlapCountsTx	12
	GRangesList2GRanges	12
	GRanges2GRangesList	10
	getStopCodon	9 10
	getPvalZscore	9 9
	getPermSpaceByType	8
	getPermSpaceByTxID	7
	getPermSpaceByFeatures	6
	getFormatCorrect	6
	extractRegions	5
	distanceTx	4
	calculateShift	2

# Description

The first step of calculating positional shift over transcriptome regions.

## Usage

```
calculateShift(regions, disp, direction = "right", strand = "+")
```

calculateShift 3

### Arguments

regions A feature set, which should be a GRangesList object.

disp A data frame object. It should have three columns, which are start: starting

positions. Each value represents a starting position in each input feature; width: widths. Each value represents a width of each region to be picked from each

feature; names: corresponding transcript ids.

direction Either to be character "left" or "right", which means the direction to which the

starting position is shifting. The former means moving to the direction of 5'

while the latter means moving to 3'.

strand Either to be "+" or "-".

#### Value

A GRanges object.

#### See Also

extractRegions

```
# Take five transcripts.
# Extract the last 200 nt regions from their CDS part.
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
trans.id.pstv <- c("170", "782", "974", "1364", "1387")
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene</pre>
# Download the CDS part of all transcriptome
cds.tx0 <- cdsBy(txdb, use.names = FALSE)</pre>
# pick the CDS part of these five transcripts
cds.p <- cds.tx0[trans.id.pstv]</pre>
width <- 200
disp.p.l <- data.frame(</pre>
    start = as.numeric(max(end(cds.p))),
    distance = width - 1,
    names = trans.id.pstv
)
R.p.l <- calculateShift(</pre>
    regions = cds.p, disp = disp.p.l,
    direction = "left", strand = "+"
)
```

4 distanceTx

distanceTx

Evaluation function

### **Description**

Evaluation function. This function calculates the mean of the distance from each region of set RS1 to the closest region in RS2.

### Usage

```
distanceTx(A, B, beta = 0.2, ...)
```

### **Arguments**

A Region set 1. A Granges or GRangesList object.

B Region set 2. A Granges or GRangesList object.

beta It is a user-defined argument that can filter out the corresponding percent of largest distance values. Default value is 0.2.

... Any additional parameters needed.

#### Value

A numeric object.

#### See Also

```
overlapWidthTx, overlapCountsTx
```

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
trans.ids <- c("170", "782", "974", "1364", "1387")
A <- randomizeTx(
    txdb, trans.ids,
    random_num = 20,
    random_length = 100
)
B <- randomizeTx(
    txdb, trans.ids,
    random_num = 20,
    random_num = 20,
    random_length = 100
)
distanceTx(A, B, beta = 0.2)</pre>
```

extractRegions 5

extractRegions	Extract	regions
extractives tons	Extract	regions

### **Description**

This function receives three arguments: the scope region set, the target region set and the type of strand. It returns a subset of target region set, which is the intersection of the target region set and the scope region set.

#### Usage

```
extractRegions(regions_A, R, strand = "+")
```

#### **Arguments**

regions\_A The scope region set. A GRangesList object. The name of each list element

should be the transcript id that it pertains to.

R The target region set. A GRanges object.

strand The strand type of the transcripts. It has options "+" and "-".

#### Value

A GRangesList object.

### See Also

```
calculateShift
```

```
# Take five transcripts.
# Extract the last 200 nt regions from their CDS part.
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
trans.id.pstv <- c("170", "782", "974", "1364", "1387")
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene

# download the CDS part of all transcriptome
cds.tx0 <- cdsBy(txdb, use.names = FALSE)

# pick the CDS part of these five transcripts
cds.p <- cds.tx0[trans.id.pstv]

width <- 200
disp.p.l <- data.frame(
    start = as.numeric(max(end(cds.p))),
    distance = width - 1,
    names = trans.id.pstv
)</pre>
```

```
R.p.l <- calculateShift(regions = cds.p, disp = disp.p.l, direction = "left", strand = "+")
R.cds.last200 <- extractRegions(regions_A = cds.p, R = R.p.l, strand = "+")</pre>
```

getFormatCorrect

getFormatCorrect

### Description

This function makes sure the two input region sets are in the correct format required by RgnTX evaluation functions.

### Usage

```
getFormatCorrect(A, B)
```

### **Arguments**

A Region set 1. A Granges or GRangesList object.

B Region set 2. A Granges or GRangesList object.

#### Value

A list object.

getPermSpaceByFeatures

Get permutation space for features

## Description

This function returns a default permutation space for features with isoform ambiguity. The default permutation space of a feature is the aggregate of the multiple transcripts it may overlap with. It requires the input feature to be GRanges format.

### Usage

```
getPermSpaceByFeatures(features, txdb, type = "mature")
```

#### **Arguments**

 $\begin{array}{ll} \text{features} & A \text{ GRanges object.} \\ \text{txdb} & A \text{ TxDb object.} \end{array}$ 

type A character object. Default is "mature". It accepts options "mature", "full",

"fiveUTR", "CDS" or "threeUTR", with which one can get corresponding types

of regions over transcriptome.

#### Value

A list object, which contains two elements.

- perm. space: A GRangesList object that includes all the transcripts input features may overlap with.
- index: It contains a series of numbers indicating which feature these transcripts are respectively associated with.

#### See Also

```
getPermSpaceByTxID, getPermSpaceByType
```

### **Examples**

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
file <- system.file(package="RgnTX", "extdata/m6A_sites_data.rds")
m6A_sites_data <- readRDS(file)
permSpace <- getPermSpaceByFeatures(features = m6A_sites_data[1:100], txdb)</pre>
```

getPermSpaceByTxID

Get permutation space by specifying transcript ids

#### **Description**

This function returns 5'UTR/CDS/3'UTR/mRNA/full part of transcriptome regions grouped by corresponding transcript ids.

### Usage

```
getPermSpaceByTxID(trans_ids = "all", txdb, type = "mature")
```

#### **Arguments**

trans\_ids A character object. The transcript ids. Default is "all". If it takes the default

value "all", the space that users get will be the whole transcriptome.

txdb A TxDb object.

type A character object. Default is "mature". It accepts options "mature", "full",

"fiveUTR", "CDS" or "threeUTR", with which one can get corresponding types

of transcriptome regions.

#### Value

A GRangesList object.

### See Also

```
getPermSpaceByType, getPermSpaceByFeatures
```

### **Examples**

```
trans.ids <- c("170", "782", "974", "1364", "1387")
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
permspace <- getPermSpaceByTxID(trans.ids, txdb)</pre>
```

getPermSpaceByType

Get permutation space by specifying type

### **Description**

This function can return 5'UTR/CDS/3'UTR/mRNA/full part of transcriptome regions, following the format required by the main permutation test functions.

### Usage

```
getPermSpaceByType(txdb, type = "mature")
```

### **Arguments**

txdb A TxDb object.

type A character object. Default is "mature". It accepts options "mature", "full",

"fiveUTR", "CDS" or "threeUTR", with which one can get corresponding types

of transcriptome regions.

### Value

A GRangesList object.

#### See Also

getPermSpaceByTxID, getPermSpaceByFeatures

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
permSpace <- getPermSpaceByType(txdb, type = "CDS")</pre>
```

getPvalZscore 9

### Description

Calculate a p-value and z-score based on observed value and random evaluation values.

### Usage

```
getPvalZscore(orig.ev, rand.ev, pval_z = FALSE)
```

### Arguments

orig.ev Observed value.

rand.ev Random evaluation values.

pval\_z Boolean. Default is FALSE. If FALSE, the p-value is calculated based on the

number of random evaluations is larger or less than the initial evaluation. If

TRUE, the p-value is calculated based on a z-test.

#### Value

A p-value and a z-score.

|--|--|

### **Description**

Get stop codon regions for input transcripts. This is an example of customPick function.

### Usage

```
getStopCodon(trans_ids, txdb, ...)
```

### **Arguments**

trans\_ids A character object containing transcript ids.

txdb A TxDb object.

... Any additional parameters needed.

### Value

A numeric object.

10 getTransInfo

### **Examples**

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
trans.ids <- c("170", "782", "974", "1364", "1387")
RS2 <- getStopCodon(trans.ids, txdb)</pre>
```

getTransInfo

Get transcript information

### **Description**

Generate a data frame object that contains information about input genomic feature set and its mapping results over the transcriptome.

### Usage

```
getTransInfo(A, txdb)
```

### **Arguments**

A Genomic feature set, which should be a GRanges object.

txdb A TxDb object.

#### Value

A data. frame object containing the following components:

- index\_trans: The label of transcripts.
- index\_features: The label of genomic features.
- segnames: The chr name.
- features\_pos: The starting coordinate of each genomic feature.
- width\_features: The width of each genomic feature.
- strand: The strand type of each genomic feature.
- trans\_ID: The ids of the transcripts that each feature can be mapped to.

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
file <- system.file(package="RgnTX", "extdata/m6A_sites_data.rds")
m6A_sites_data <- readRDS(file)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
getTransInfo(A = m6A_sites_data[1:100], txdb)</pre>
```

GRanges2GRangesList

 ${\tt GRanges2GRangesList}$ 

Convert a GRanges object to a GRangesList object

### **Description**

Convert a GRanges object to a GRangesList object. The output region set follows the format required by the main permutation test functions.

### Usage

```
GRanges2GRangesList(A = NULL)
```

### **Arguments**

Α

A GRanges object.

#### **Details**

If input GRanges object has a metadata named as "group", ranges having the same group number represent a region. If not, a range is a region. A region in the input set will be outputted as a list element IN returned GRangesList object.

### Value

A GRangesList object.

#### See Also

GRanges2GRangesList

```
library(GenomicRanges)
GRanges.object <- GRanges(
    Rle(c("chr2", "chr2", "chr1", "chr3")),
    IRanges(1:4, width = 5)
)
# Assign the first and the second ranges to the same element.
GRanges.object$group <- c(1, 1, 2, 3)
GRangesList.object <- GRanges2GRangesList(GRanges.object)</pre>
```

12 overlapCountsTx

GRangesList2GRanges

Convert a GRangesList object to a GRanges object

### **Description**

Convert a GRangesList object to a GRanges object. The output region set follows the format required by the RgnTX permutation test functions, which should have metadata columns 'group' and 'transcriptsHits'.

#### Usage

```
GRangesList2GRanges(A = NULL)
```

### **Arguments**

Α

A GRangesList object.

#### Value

A GRanges object. Its transcript ids (if available) should be contained in a metadata column named "transcriptsHits", which are provided by the names of input GRangesList object.

#### See Also

GRanges2GRangesList

### **Examples**

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
trans.ids <- c("170", "782", "974", "1364", "1387")
RS1 <- randomizeTx(txdb, trans.ids, random_num = 100, random_length = 100)
RS1 <- GRangesList2GRanges(RS1)</pre>
```

overlapCountsTx

**Evaluation function** 

### **Description**

This function receives two region sets and returns the number of their overlaps.

### Usage

```
overlapCountsTx(A, B, count_once = TRUE, over_trans = TRUE, ...)
```

overlapCountsTxIA 13

### **Arguments**

A	Region set 1. A GRangesList object.
В	Region set 2. A GRangesList object.
count_once	Whether the overlap of multiple B regions with a single A region should be counted once or multiple times.
over_trans	Whether the overlapping is counted over the transcriptome or over the genome.
	Any additional parameters needed.

#### Value

A numeric object.

### See Also

```
overlapCountsTx
```

### **Examples**

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
trans.ids <- c("170", "782", "974", "1364", "1387")
exons.tx0 <- exonsBy(txdb)
regions.A <- exons.tx0[trans.ids]
A <- randomizeTransByOrder(regions.A, random_length = 200)
B <- randomizeTransByOrder(regions.A, random_length = 200)
overlapCountsTx(A, B)</pre>
```

overlapCountsTxIA

Evaluation function

### Description

Evaluation function. This function receives a feature set (with isoform ambiguity) and a transcriptome region set (without isoform ambiguity), and returns a weighted number of overlaps between them.

### Usage

```
overlapCountsTxIA(A, B, ...)
```

### Arguments

A A feature set, which should be GRanges.

B A region set, which should be GRangesList.

. . . Any additional parameters needed.

14 overlapWidthTx

#### Value

A numeric object.

#### See Also

```
overlapWidthTx, distanceTx, overlapCountsTx
```

#### **Examples**

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
file <- system.file(package="RgnTX", "extdata/m6A_sites_data.rds")
m6A_sites_data <- readRDS(file)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
RS1 <- m6A_sites_data[1:100]

trans.info <- getTransInfo(RS1, txdb)
trans.ids <- trans.info[, "trans_ID"]

RS2 <- getStopCodon(trans.ids, txdb = txdb)

# Evaluation step.
orig.ev <- overlapCountsTxIA(RS1, RS2)</pre>
```

overlapWidthTx

Evaluation function

### **Description**

Evaluation function. This function returns the sum of widths of each overlap between two region sets, i.e., the total number of overlapping nucleotides between two input region sets.

### Usage

```
overlapWidthTx(A, B, ...)
```

### **Arguments**

A Region set 1. A Granges or GRangesList object.
 B Region set 2. A Granges or GRangesList object.
 ... Any additional parameters needed.

#### Value

A numeric object.

### See Also

```
overlapCountsTx, distanceTx
```

permTestTx 15

### **Examples**

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
trans.ids <- c("170", "782", "974", "1364", "1387")
A <- randomizeTx(
    txdb, trans.ids, random_num = 20,
    random.length = 100
)
B <- randomizeTx(
    txdb, trans.ids = trans.ids, random_num = 20,
    random_length = 100
)
overlapWidthTx(A, B)</pre>
```

permTestTx

Perform permutation test

### **Description**

Perform permutation test for evaluating spatial association between a feature set and a region set.

#### Usage

```
permTestTx(RS1 = NULL, RS2 = NULL, txdb = NULL, type = "mature",
ntimes = 50, ev_function_1 = overlapCountsTx, ev_function_2 = overlapCountsTx,
pval_z = FALSE, ...)
```

### **Arguments**

RS1	The region set to be randomized. It should be in the GRanges or GRangesList format.
RS2	The region set to be compared with. It should be in the GRanges or GRangesList format.
txdb	A TxDb object.
type	A character object. Default is "mature". It accepts options "mature", "full", "fiveUTR", "CDS" or "threeUTR", with which one can get corresponding types of transcriptome regions.
ntimes	Randomization times.
ev_function_1	Evaluation function defines what statistic to be tested between RS1 and RS2. Default is overlapCountsTx.
ev_function_2	Evaluation function defines what statistic to be tested between each element in RSL and RS2. Default is overlapCountsTx.
pval_z	Boolean. Default is FALSE. If FALSE, the p-value is calculated based on the number of random evaluations is larger or less than the initial evaluation. If TRUE, the p-value is calculated based on a z-test.
	Any additional parameters needed.

16 permTestTxIA

#### **Details**

permTestTxIA only needs users to input two region sets. It will automatically randomize the first region set into transcriptome.

#### Value

A list object, which is defined to be permTestTx.results class. It contains the following items:

- RSL: Randomized region sets of RS1.
- RS1: The feature set to be randomized.
- RS2: The region set to be compared with the feature set.
- orig.ev: The value of overlapping counts between RS1 and RS2.
- rand.ev: The values of overlapping counts between each element in RSL and RS2.
- pval: p-value of the test.
- zscore: Standard score of the test.

#### See Also

```
plotPermResults
```

### **Examples**

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
exons.tx0 <- exonsBy(txdb)
trans.ids <- sample(names(exons.tx0), 500)

A <- randomizeTx(txdb, trans.ids, random_num = 100, random_length = 100)
B <- c(randomizeTx(txdb, trans.ids, random_num = 75, random_length = 100), A[1:25])
permTestTx_results <- permTestTx(A, B, txdb, ntimes = 5)</pre>
```

permTestTxIA

Perform permutation test

### **Description**

Perform permutation test for evaluating spatial association between some features (with isoform ambiguity) and a region set. It randomizes the features and compares it with the region set to see if there is an association between the features and the region set. The difference between this function and permTestTx is that it is for RNA-related genomic features that have isoform ambiguity, i.e., features that one does not know which transcript they comes from.

permTestTxIA 17

### Usage

### **Arguments**

RS1	The feature set to be randomized. It should be in the GRanges or GRangesList format.
RS2	The region set to be compared with. It should be in the GRanges or GRangesList format.
txdb	A TxDb object.
type	A character object. Default is "mature". It accepts options "mature", "full", "fiveUTR", "CDS" or "threeUTR", with which one can get corresponding types of transcriptome regions.
ntimes	Randomization times.
ev_function_1	Evaluation function defines what statistic to be tested between RS1 and RS2. Default is overlapCountsTx.
ev_function_2	Evaluation function defines what statistic to be tested between each element in RSL and RS2. Default is overlapCountsTx.
pval_z	Boolean. Default is FALSE. If FALSE, the p-value is calculated based on the number of random evaluations is larger or less than the initial evaluation. If TRUE, the p-value is calculated based on a z-test.
	Any additional parameters needed.

#### **Details**

permTestTxIA only needs users to input two region sets. It will automatically randomize the first region set into transcriptome.

### Value

A list object, which is defined to be permTestTx.results class. It contains the following items:

- RSL: Randomized region sets of RS1.
- RS1: The feature set to be randomized.
- RS2: The region set to be compared with the feature set.
- $\bullet\,$  orig.ev: The value of overlapping counts between RS1 and RS2.
- rand. ev: The values of overlapping counts between each element in RSL and RS2.
- pval: p-value of the test.
- zscore: Standard score of the test.

#### See Also

```
plotPermResults
```

### **Examples**

permTestTxIA\_customPick

Perform permutation test

### **Description**

Perform permutation test for evaluating spatial association between RNA features and a specified kind of regions. The latter is defined by the customPick\_function argument input by users. The difference between this function and permTestTx\_customPick is that it is for RNA-related genomic features that have isoform ambiguity, i.e., features that one does not know which transcript they comes from.

#### Usage

```
permTestTxIA_customPick(RS1 = NULL, txdb = NULL, type = 'mature',
customPick_function = NULL, ntimes = 50,
ev_function_1 = overlapCountsTxIA, ev_function_2 = overlapCountsTx, pval_z = FALSE, ...)
```

#### **Arguments**

RS1 The region set to be randomized. It should be in the GRanges or GRangesList

format.

txdb A TxDb object.

type A character object. Default is "mature". It accepts options "mature", "full",

"fiveUTR", "CDS" or "threeUTR", with which one can get corresponding types

of transcriptome regions.

customPick\_function

A custom function needs to be inputted by users. The customPick function should have two arguments: a TxDb object and a character object of transcript ids. It returns a specified region over each transcript.

ntimes	Randomization times.
ev_function_1	Evaluation function defines what statistic to be tested between RS1 and RS2. Default is overlapCountsTxIA.
ev_function_2	Evaluation function defines what statistic to be tested between each element in RSL and RS2. Default is overlapCountsTx.
pval_z	Boolean. Default is FALSE. If FALSE, the p-value is calculated based on the number of random evaluations is larger or less than the initial evaluation. If TRUE, the p-value is calculated based on a z-test.
	Any additional parameters needed.

#### **Details**

permTestTxIA\_customPick will assess the test statistic between RS1 and each region in RSL, and the relation between RS1 and RS2. Each RNA feature is only mapped with a part of region on its transcript (picked by the customPick\_function). The output orig.ev is the weighted counts between RS1 and RS2. Each feature in RS1 related to n1 isoforms in RS2 and overlapped with n2 RS2 regions will contribute a value of n2/n1 to the total number of overlaps. This test function also randomizes input features per transcript. The set of randomized results is outputted as RSL. The overlapping counts between each set in RSL with RS2 is outputted as rand.ev.

#### Value

A list object, which is defined to be permTestTx.results class. It contains the following information:

- RSL: Randomized region sets of RS1.
- RS1: The feature set to be randomized.
- RS2: The region set to be compared with the feature set.
- orig.ev: The value of overlapping counts between RS1 and RS2.
- rand.ev: The values of overlapping counts between each element in RSL and RS2.
- pval: p-value of the test.
- zscore: Standard score of the test.

#### See Also

```
plotPermResults
```

```
type = 'mature',
customPick_function = getStopCodon,
ntimes = 5)
```

```
permTestTx_customAll Perform permutation test
```

### **Description**

Perform permutation test for evaluating spatial association between region sets. This permutation test function receives two region sets and a set of randomized region sets of one of them. It evaluates if there is an association between these two region sets.

### Usage

```
permTestTx_customAll(RSL = NULL, RS1 = NULL, RS2 = NULL,
ev_function_1 = overlapCountsTx, ev_function_2 = overlapCountsTx, pval_z = FALSE, ...)
```

#### **Arguments**

RSL	Randomized region sets of RS1. It should be a list object and each element should be in the GRanges or GRangesList format.
RS1	The region set. It should be in the GRanges or GRangesList format.
RS2	The region set to be compared with. It should be in the GRanges or GRangesList format.
ev_function_1	Evaluation function defines what statistic to be tested between RS1 and RS2. Default is overlapCountsTx.
ev_function_2	Evaluation function defines what statistic to be tested between each element in RSL and RS2. Default is overlapCountsTx.
pval_z	Boolean. Default is FALSE. If FALSE, p-value is calculated based on the number of random evaluations is larger or less than the initial evaluation. If TRUE, p-value is calculated based on a z-test.
	Any additional parameters needed.

#### **Details**

permTestTx\_customAll will use evaluation function ev\_function\_1 to calculate the test statistic between RS1 and RS2, and use ev\_function\_2 to evaluate the statistic between RSL and RS2. It will also return a p-value and a z-score.

#### Value

A list object, which is defined to be permTestTx.results class. It contains the following items:

- RSL: Randomized region sets of RS1.
- RS1: The feature set to be randomized.
- RS2: The region set to be compared with the feature set.
- orig.ev: The value of overlapping counts between RS1 and RS2.
- rand.ev: The values of overlapping counts between each element in RSL and RS2.
- pval: p-value of the test.
- zscore: Standard score of the test.

#### **Examples**

permTestTx\_customPick Perform permutation test

#### **Description**

Perform permutation test for evaluating spatial association between a feature set and the customPick regions. The latter is defined by the customPick\_function argument provided by users.

#### Usage

```
permTestTx_customPick(RS1 = NULL, txdb = NULL, type = "mature",
customPick_function = NULL, ntimes = 50, ev_function_1 = overlapCountsTx,
ev_function_2 = overlapCountsTx, pval_z = FALSE, ...)
```

### **Arguments**

RS1 The feature set to be randomized. It should be in the GRanges or GRangesList format.

txdb A TxDb object.

type A character object. Default is "mature". It accepts options "mature", "full",

"fiveUTR", "CDS" or "threeUTR", with which one can get corresponding types

of transcriptome regions.

customPick\_function

A custom function needs to be inputted by users. The custom function should have two arguments: a TxDb object and a character object of transcript ids. It

returns a part of region of each transcript.

ntimes Randomization times.

ev\_function\_1 Evaluation function defines what statistic to be tested between RS1 and RS2.

 $Default\ is\ {\tt overlapCountsTx}.$ 

ev\_function\_2 Evaluation function defines what statistic to be tested between each element in

RSL and RS2. Default is overlapCountsTx.

pval\_z Boolean. Default is FALSE. If FALSE, the p-value is calculated based on the

number of random evaluations is larger or less than the initial evaluation. If

TRUE, the p-value is calculated based on a z-test.

... Any additional parameters needed.

#### **Details**

Each feature in RS1 is only mapped with the customPick regions over its transcript (picked by the customPick\_function). The output orig.ev is the number of features that have overlap with its customPick region. The set of randomized region sets is outputted as RSL. The overlapping counts between each set in RSL with RS2 is outputted as rand.ev.

### Value

A list object, which is defined to be permTestTx.results class. It contains the following items:

- RSL: Randomized region sets of RS1.
- RS1: The feature set to be randomized.
- RS2: The region set to be compared with the feature set.
- orig.ev: The value of overlapping counts between RS1 and RS2.
- rand.ev: The values of overlapping counts between each element in RSL and RS2.
- pval: p-value of the test.
- zscore: Standard score of the test.

### See Also

plotPermResults

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
exons.tx0 <- exonsBy(txdb)
trans.ids <- sample(names(exons.tx0), 100)</pre>
```

plotPermResults 23

```
RS1 <- randomizeTx(txdb, trans.ids, random_num = 100,
random_length = 200, type = 'CDS')
getCDS = function(txdb, trans.id){
cds.tx0 <- cdsBy(txdb, use.names=FALSE)
    cds.names <- as.character(intersect(names(cds.tx0), trans.id))
    cds = cds.tx0[cds.names]
    return(cds)
}

permTestTx_results <- permTestTx_customPick(RS1,txdb,
customPick_function = getCDS, ntimes = 5)</pre>
```

plotPermResults

Plot permutation test results

### **Description**

Show a graphical representation of permutation test.

### Usage

```
plotPermResults(permTestTx_results = NULL, breaks = 15, alpha = 0.05,
test_type = "one-sided", binwidth = NULL)
```

#### **Arguments**

permTestTx\_results

A permTestTx.results list object, which can be generated by the permTestTx

function.

breaks Histogram breaks. Default is 15.

alpha Significance level.

test\_type The type of the test. This argument only receives either two options "one-sided"

or "two-sided". Default is "one-sided".

binwidth Histogram binwidth.

### Value

A plot object.

#### See Also

```
permTestTx
```

```
file <- system.file(package="RgnTX", "extdata", "permTestTx_results.rds")
permTestTx_results <- readRDS(file)
p_a <- plotPermResults(permTestTx_results, binwidth = 1)
p_a</pre>
```

24 randomizeFeaturesTx

### Description

Plot shifted z scores for permutation test results.

### Usage

```
plotShiftedZScoreTx(shitedZScoresTx_results)
```

### **Arguments**

```
\label{eq:shitedZScoresTx_results} A \ shitedZScoreTx.results \ object.
```

#### Value

A plot.

#### See Also

shiftedZScoreTx

### **Examples**

```
file <- system.file(package="RgnTX", "extdata", "shiftedZScoreTx_results1.rds")
shiftedZScoreTx_results <- readRDS(file)
p1 <- plotShiftedZScoreTx(shiftedZScoreTx_results)
p1</pre>
```

randomizeFeaturesTx

Randomize features into transcriptome

### **Description**

Randomize features into transcriptome.

### Usage

```
randomizeFeaturesTx(RS, txdb, type = "mature", N = 1, ...)
```

randomizeFeaturesTxIA 25

### **Arguments**

RS	The feature to be randomized. It should be a GRanges or GRangesList object.
txdb	A TxDb object.
type	A character object. Default is "mature". It accepts options "mature", "full", "fiveUTR", "CDS" or "threeUTR", with which one can get corresponding types of transcriptome regions.
N	The number of iterations.
	Any additional parameters needed.

#### Value

A GRangesList object. The name of each element is the id of the transcript where the corresponding range is located.

#### See Also

 $randomize Trans By Order, \ randomize Features TxIA, \ randomize TxIA \$ 

### **Examples**

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
trans.ids <- c("170", "782", "974", "1364", "1387")
RS1 <- randomizeTx(txdb, trans.ids, random_num = 100, random_length = 100)
RS <- randomizeFeaturesTx(RS1, txdb, N = 1)</pre>
```

randomizeFeaturesTxIA Randomize features into transcriptome

### **Description**

Randomize features into transcriptome, especially for the features that have isoform ambiguity.

### Usage

```
randomizeFeaturesTxIA(RS, txdb, type = "mature", N = 1, ...)
```

### **Arguments**

RS	The feature being randomized. It should be a GRanges or GRangesList object.
txdb	A TxDb object.
type	A character object. Default is "mature". It accepts options "mature", "full", "fiveUTR", "CDS" or "threeUTR", with which one can get corresponding types of transcriptome regions.
N	Randomization times.
	Any additional parameters needed.

#### Value

A GRangesList object. The name of each element is the id of the transcript where the corresponding range is located.

#### See Also

randomizeTransByOrder, randomizeFeaturesTx, randomizeTx

### **Examples**

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
file <- system.file(package="RgnTX", "extdata/m6A_sites_data.rds")
m6A_sites_data <- readRDS(file)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
RS1 <- m6A_sites_data[1:100]
RS <- randomizeFeaturesTxIA(RS1, txdb, N = 1)</pre>
```

randomizeTransByOrder Randomize features into transcriptome

### Description

This function receives a GRangesList object and picks a random region within each list element of this object. The length of the region to be picked is decided by the input random\_length argument.

#### Usage

```
randomizeTransByOrder(regions_A, random_length = 20)
```

### Arguments

regions\_A A GRangesList object. The name of each list element should be the correspond-

ing transcript id.

random\_length A numeric object.

#### Value

A GRangesList object. The name of each list element should be the corresponding transcript id.

#### See Also

randomizeTx, randomizeFeaturesTx, randomizeFeaturesTxIA

randomizeTx 27

#### **Examples**

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
exons.tx0 <- exonsBy(txdb)
trans.ids <- sample(names(exons.tx0), 500)
regions.A <- exons.tx0[trans.ids]
RS <- randomizeTransByOrder(regions.A, random_length = 20)</pre>
```

randomizeTx

Get randmized regions over transcriptome

### **Description**

Pick random regions over specified transcripts.

### Usage

```
randomizeTx(txdb, trans_ids = 'all',
random_num = 100, random_length = 20, type = 'mature', N = 1, ...)
```

#### **Arguments**

txdb A TxDb object.

trans\_ids The ids of transcripts, which should be a character object. Random regions will

be picked from these transcripts. If this argument takes the default value 'all',

the scope of picking random regions will be the whole transcriptome.

random\_num The number of regions to be picked.

random\_length The length of regions to be picked.

type A character object. Default is "mature". It accepts options "mature", "full",

"fiveUTR", "CDS" or "threeUTR", with which one can get corresponding types

of transcriptome regions.

N Randomization times.

... Any additional parameters needed.

#### Value

A GRangesList object. The name of each element is the id of the transcript where the corresponding range is located.

#### See Also

randomizeTransByOrder, randomizeFeaturesTx, randomizeFeaturesTxIA

28 shiftedZScoreTx

#### **Examples**

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
trans.ids <- c("170", "782", "974", "1364", "1387")
RS1 <- randomizeTx(txdb, trans.ids, random_num = 100, random_length = 100)</pre>
```

shiftedZScoreTx

Calculate shifted z scores

#### **Description**

Calculate shifted z scores for permutation test results.

### Usage

```
shiftedZScoreTx(permTestTx_results = NULL, txdb = NULL,
window = 200, step = 20, ev_function_1 = overlapCountsTx, ...)
```

#### **Arguments**

permTestTx\_results

A permTestTx.results object.

txdb A TxDb object.

window The window of the whole shifting.

step The step of each shifting.

ev\_function\_1 Evaluation function. Default is overlapCountsTx.

... Any additional parameters needed.

### **Details**

see examples in plotShiftedZScoreTx

#### Value

A list object, which is defined to be shitedZScore.results class. It contains the following items:

- shifted.z.scores: Standard z-scores after shifting.
- window: Window of the whole shifting.
- step: Step of each shifting.
- original.z.score: Original standard score.

### See Also

```
plotShiftedZScoreTx
```

shiftTx 29

### **Examples**

shiftTx

Shift over transcripts

### **Description**

Calculate positional shifting over transcript regions. This function accepts a feature set and outputs a region set from it. Each output region is from each input feature.

#### Usage

```
shiftTx(regions, start, width, direction, strand)
```

### **Arguments**

regions	A feature set following the format indicated in vignette section 3. Either to be GRanges or GRangesList.
start	Starting positions. Each value represents a starting position in each input feature.
width	Widths. Each value represents a width of each region to be picked from each feature.
direction	Either to be character "left" or "right", which means the direction to which the starting position is shifting. The former means moving to the direction of 5' while the latter means moving to 3'.
strand	The strand type of the transcripts. It receives "+" or "-".

#### Value

A Granges object.

30 vector2GRangesList

#### **Examples**

```
# Take five transcripts.
# Extract the last 200 nt regions from their CDS part.
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
trans.id.pstv <- c("170", "782", "974", "1364", "1387")
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene

# download the CDS part of all transcriptome
cds.tx0 <- cdsBy(txdb, use.names = FALSE)

# pick the CDS part of these five transcripts
cds.p <- cds.tx0[trans.id.pstv]

width <- 200
start <- as.numeric(max(end(cds.p)))
R.cds.last200 <- shiftTx(cds.p, start = start, width = width, direction = 'left', strand = "+")</pre>
```

vector2GRangesList vector2GRangesList

### **Description**

Generate GRangesList object from vectors. The output region set follows the format required by the main permutation test functions.

### Usage

```
vector2GRangesList(RefSeqID, targetName, strand, blockSizes, targetStart)
```

### **Arguments**

RefSeqID The name of each element.

targetName The seqnames of each range.

strand The strand of each range.

blockSizes The width of each range.

targetStart The stard coordinate of each range.

#### Value

A GRangesList object.

### See Also

GRanges2GRangesList

# **Index**

```
calculateShift, 2, 5
distanceTx, 4, 14
extractRegions, 3, 5
getFormatCorrect, 6
getPermSpaceByFeatures, 6, 7, 8
getPermSpaceByTxID, 7, 7, 8
getPermSpaceByType, 7, 8
getPvalZscore, 9
getStopCodon, 9
getTransInfo, 10
GRanges2GRangesList, 11, 11, 12, 30
GRangesList2GRanges, 12
overlapCountsTx, 4, 12, 13, 14
overlapCountsTxIA, 13
overlapWidthTx, 4, 14, 14
permTestTx, 15, 16, 23
permTestTx_customAll, 20
permTestTx_customPick, 18, 21
permTestTxIA, 16
permTestTxIA\_customPick, 18
plotPermResults, 16, 18, 19, 22, 23
plotShiftedZScoreTx, 24, 28
randomizeFeaturesTx, 24, 26, 27
randomizeFeaturesTxIA, 25, 25, 26, 27
randomizeTransByOrder, 25, 26, 26, 27
randomizeTx, 25, 26, 27
shiftedZScoreTx, 24, 28
shiftTx, 29
{\tt vector2GRangesList, 30}
```